# *REVIEW Article*

# **Complexities of Dengue Fever: Pathogenesis, Clinical Features and Management Strategies**

*Maheen Nasir 1, #, \*, Javeria Irfan 1, # , Aimen Binte Asif 1, # , Qudsia Umaira Khan <sup>1</sup> ,* 

*Haleema Anwar <sup>1</sup>*

<sup>1</sup> CMH Lahore Medical and Dental College, Pakistan

*#* These authors contributed equally to this work

*\* Corresponding author*: *Maheen Nasir,* CMH Lahore Medical and Dental College, Pakistan, maheennasir\_@hotmail.com

*Submitted:* May 05, 2024; *Revised:* June 30, 2024; *Accepted:* June 30, 2024.

*Citation:* Nasir M, Irfan J, Asif AB, Khan QU, Anwar H. Complexities of Dengue Fever: Pathogenesis, Clinical Features and Management Strategies. *Discoveries* 2024, e189. DOI: 10.15190/d.2024.8

# **ABSTRACT**

Dengue fever, transmitted through the bite of infected Aedes mosquitos, poses a significant global threat, particularly in the tropical and subtropical region. In this review, we aim to summarize the existent literature on dengue virus infection and to enlighten the reader on recent advances and knowledge. Dengue virus infection can cause a spectrum of clinical manifestations, ranging from asymptomatic or mild illness to more severe and potentially lifethreatening complications. Pathogenesis of dengue is based on viral and host factors. Viral factors include NS1 antigen and genomic factors. Host factors include antibody dependent enhancement, anti-NS1 antibodies, cytokines, cross reactive T-Cell response, HLA allele variation and non-HLA mediated polymorphisms. The clinical picture of dengue is described on the basis of WHO 1997 and 2009 criteria. It is classified into dengue fever, dengue hemorrhagic fever (DHF), and dengue shock syndrome (DSS). Life-threatening complications can develop in severe cases, and this includes renal complications such as acute kidney injury (AKI) and hepatic complications such as hepatic dysfunction and in rare cases, fulminant hepatic failure. Neurological complications, cardiac complications and respiratory distress syndrome have also been reported. Treatment methods include targeting the dengue vector and Carica papaya, a natural remedy

with antiviral properties. Additionally, the role of corticosteroids, intravenous immunoglobulins, and mast cell inhibitors has been explored in dengue treatment, aiming to reduce severity. Novel approaches involve drugs targeting dengue proteins and host factors necessary for the virus's life cycle, offering potential avenues for more targeted therapeutic interventions. In recent years, significant progress has been made in the development of vaccines against dengue, with Sanofi Pasteur's Dengvaxia being the first licensed vaccine approved for use. Utilizing various approaches such as recombinant proteins, viral vectors and viral like particles, various alternatives have been provided which aim to be safer substitutes to Dengvaxia while maintaining the effectiveness. A review on dengue is essential for clinicians and healthcare professionals to stay updated on diagnostics, treatment protocols and prevention strategies.

#### **SUMMARY**

- *1. Introduction*
- *2. Pathogenesis*
	- *2.1. Host Factors 2.2. Viral Factors*
- *3. Clinical features*
	- *3.1. WHO Classification*
	- *3.2. Clinical Course*
- *4. Complications*
- *5. Treatment*
	- *5.1. Fluid Management*
	- *5.2. Drugs Targeting Dengue Proteins*
	- *5.3. Alternative Methods*
	- *5.4. Symptomatic Management*
- *6. Prevention: Vaccines*
	- *6.1. Advancements in vaccine development*
	- *6.2. Types of vaccines*
	- *6.3. Notable Examples*
	- *6.4. Challenges in the development of vaccine*
	- *6.5. Future of vaccine development*
- *7. Conclusion*

#### **Abbreviations**

Dengue virus (DENV), Dengue fever (DF), Dengue Hemorrhagic Fever (DHF), Dengue Shock Syndrome (DSS), Acute Kidney Injury (AKI), World Health Organization (WHO), Antibody-dependent Enhancement (ADE), Soluble Non Structural Protein 1 Antigen (sNS1), Membrane Attack Complex (MAC), Subgenomic Flavivirus RNA (sfRNA), Human Leukocyte Antigen (HLA), Vitamin D receptor (VDR), Human Platelet Antigen (HPA), Tumor Necrosis Factor (TNF), Acute Transverse Myelitis (ATM), Acute Coronary Syndrome (ACS), Acute Respiratory Distress Syndrome (ARDS), RNA-dependent RNA polymerase (RdRP), Chloroquine (CQ) Recombinant factor VIIa (rFVIIa), Intravenous Immunoglobulins (IVIG), Idiopathic Thrombocytopenic Purpura (ITP), Tetravalent Dengue Vaccine (TAK-003)

#### **Keywords**

Dengue, dengue fever, dengue virus, dengue virus receptor, dengue vaccine, dengue hemorrhagic fever.

# **1. Introduction**

Dengue virus (DENV) is a single-stranded RNA virus of the Flaviviridae family and the Flavivirus genus<sup>1,2</sup>. The distinctive feature of arboviruses, or arthropodborne viruses, is their ability to spread from arthropod vectors to vertebrate hosts despite their taxonomically different nature. They are categorized based on replicative methods, morphology, and antigenic connections.

Arboviruses belong to the Togaviridae, Flaviviridae, Bunyaviridae, Rhabdoviridae, Orthomyxoviridae, and Reoviridae virus families<sup>6</sup>. This virus is spread through the bite of an Aedes species female mosquito, primarily Aedes aegypti but also infrequently Aedes albopictus<sup>3</sup>. Four dengue virus serotypes (DEN-1, DEN-2, DEN-3, and DEN-

4) belonging to the Flavivirus genus cause dengue infection in humans.

Dengue fever (DF), dengue hemorrhagic fever (DHF), and dengue shock syndrome (DSS) are the three categories into which symptomatic dengue virus infection has been divided, according to the WHO classification from 1997 as shown in Figure 1. Dengue without warning signs, dengue with warning signs (abdominal pain, persistent vomiting, fluid accumulation, mucosal bleeding, lethargy, liver enlargement, increasing hematocrit with decreasing platelets), and severe dengue are the three categories of dengue patients according to the revised WHO classification of 2009<sup>4</sup> . The symptoms of DF, an acute febrile illness, include headaches, leukopenia, rash, and pains in the muscles, joints, and bones. The four main clinical signs of DHF are high fever, bleeding, frequently accompanied by hepatomegaly, and, in extreme situations, circulatory collapse. A significant amount of plasma leakage may cause hypovolemic shock in some of the affected people<sup>5</sup>. Whereas in DSS, there is a risk of serious bleeding, shock, and up to 20% fatality if treatment is not received<sup>6</sup>.



**Figure 1. Spectrum of clinical manifestations seen with dengue virus infection5-7**

Dengue fever, a mosquito-borne infectious disease prevalent in tropical and subtropical regions, affects an estimated 3.6 billion individuals globally. The dengue virus (DENV) causes a substantial burden, with up to 390 million infections and 96 million symptomatic cases each year. The first reports of dengue or dengue-like disease date back to 1780 in Madras, India; however, the first virologically confirmed dengue fever epidemic in India took place in Calcutta and along the Eastern Coast of India in 1963–1964. The Philippines was the first place where DHF, a severe illness derived from DF patients, was initially documented in 1953. Eight DHF was theorized to be the result of numerous DENV infections because patients in the Philippines in 1956 had different serotypes (DENV-2, 3 and 4) isolated from them. During an epidemic in Bangkok, Thailand in 1958, people with multiple DENV infections were also isolated<sup>9</sup>. The incubation period ranges from three to fourteen days, with an average duration of five to seven days, during which viremic hosts can transmit the virus to mosquitoes for five to twelve days. Notably, asymptomatic cases, although capable of spreading the virus, often go undetected in global surveillance systems.

Dengue is endemic in approximately 125 countries, with transmission reported across all World Health Organization (WHO) regions. International travel facilitates the importation of cases into both endemic and non-endemic countries. In non-tropical areas, Aedes albopictus serves as the primary vector, with a broader host range, while Aedes aegypti predominates in tropical regions, posing a risk to travelers through mosquito bites<sup>7</sup> Frequent outbreaks and a high prevalence of dengue disease put a severe strain on health services and the nation's economy. The three primary methods for preventing and controlling the spread of the dengue virus are vector control, case management, and case detection. There is currently a dengue vaccine on the market, and more vaccines are being developed. To properly use current and new preventive and control techniques, judgments on the burden, prevalence, incidence, and geographic distribution of dengue disease must be made. Considering this, we estimated the illness burden of dengue fever by a systematic study<sup>8</sup>.

# **2. Pathogenesis**

Dengue fever is a mosquito-borne disease caused by one of the four antigenically distinguishable serotypes i.e. DENV-1 to DENV-4. Pathogenesis is mediated by viral and host factors as shown in Table 1. Viral factors include NS1 antigen, subgenomic flavivirus RNA and other genomic factors while host factors include cross-reactive T-Cell response,

antibody-dependent enhancement, anti-NS1 antibodies, cytokines, HLA and non-HLA mediated polymorphisms.

<b>Host Factors</b>	<b>Viral Factors</b>
Antibody dependent	NS1 antigen
enhancement	
Cross reactive T cell response	sfRNA
Anti-NS1 antibodies	Genotypic variation
<b>HLA</b> alleles	
Non HLA polymorphisms	
Cytokines	

**Table 1. Viral and host factors mediating pathogenesis of dengue fever11,12,19,20**

# *2.1 Cross-Reactive T Cell Response*

On infection with DENV, the ability of dendritic cells to mature and migrate will be interrupted by NS1 protein through regulation of associated gene  $expression<sup>10</sup>$ . Dendritic cells then express the target antigen to CD8+ AND CD4+ T-cells. CD4+ cells act on structural and NS1 proteins thus assisting in humoral response, T-cell mediated memory response and producing cytotoxic effect. Whereas CD8+ cells act on non-structural proteins and produce cytolytic effects on infected cells directly.

During a secondary DENV infection, CD8+ T-Cells with high affinity for the infecting virus are selectively activated and produce large numbers of inflammatory cytokines like IFN-γ, IL-13 and TNFα. Those with low affinity for the heterologous infecting virus, however, are favorably multiplied. They lose their cytotoxic ability so viral clearance is prolonged but produces large numbers of inflammatory cytokines which play a major role in increased vascular permeability<sup>11</sup>.

Moreover, the heterologous DENV infection activates memory T-cells selective for the primary DENV strain rather than I T-cells. This phenomenon is called original antigenic  $sin<sup>12</sup>$ .

# *2.2 Antibody-Dependent Enhancement (Ade)*

Infection with one DENV serotype provides lifetime protection against it while only short-term crossreactive protection from other serotypes. When a secondary infection occurs, sub or non-neutralizing antibodies bind to DENV and ease its entrance into host cells by phagocytosis thus enhancing the virulence of the heterotypic strain. ADE occurs in

infants as well due to the interaction between maternal antibodies and primary infection. ADE causes a high risk of severe infection in those who were infected primarily because of the production of low levels of sub-neutralizing antibodies against other serotypes<sup>13</sup>.

It has two mechanisms:

- Intrinsic ADE acts on the innate immune system to down-regulate levels of type-1 interferon, interleukin-12, interferon-γ and TNF thereby reducing the antiviral capability of DENV. This increases the burst size i.e. viral release from host  $\text{cells}^{14}$ .
- Extrinsic ADE refers to the increased number of infected cells when antibodies fall below their neutralizing ability<sup>15</sup>.

# *2.3 Soluble Non-Structural Protein 1 Antigen (Sns1)*

sNS1 protein is a glycoprotein existing as a dimer on the surface of DENV and secreted as a hexamer from infected cells. It can be detected in the bloodstream acute dengue so is used as a diagnostic marker<sup>16</sup>. It plays a part in viral morphogenesis and replication. Its role in the pathogenesis of dengue includes:

- sNS1 binds to endothelium and increases production of heparanidase and sialidase that disturb its normal structure. It also breaks the intracellular junctions by clathrin-mediated internalization or phosphorylation. This causes vessel damage and plasma leakage<sup>18</sup>.
- Direct binding to TLR-4 increases expression of various cytokines and vasoactive amines e.g. interleukin 6 or TNF- alpha which increase the risk of vascular disorders leading to severe  $d$ engue<sup>17</sup>.
- It causes fixation of many complement system components like membrane attack complex (MAC) and mannose-binding lectin, decreasing its ability to neutralize viruses and stopping activation by the lectin pathway<sup>18,23</sup>.

# *2.4 Anti-Ns1 Antibodies*

The presence of these antibodies causes immune activation and release of many inflammatory mediators like interleukin-6, interleukin-8 and MCP-1. They produce effects that may lead to  $DHF<sup>19</sup>$ . In severe dengue, hepatic damage is seen due to anti-NS1 antibodies as demonstrated in mice, the liver of

whom showed the antibodies deposited in vascular endothelium and macrophages when immunized passively. When actively immunized the mice's liver showed hepatic fibrosis, and fatty liver and liver enzymes were raised<sup>20</sup>.

# *2.5 Genomic Factors*

Certain DENV strains produce a greater risk of epidemics due to increased replicative potential in humans or mosquitoes. Further, the sequence of infection may affect the pathogenic ability of a specific serotype<sup>21</sup>. For example, DENV-1 results in higher viremia in contrast to DENV-2 and DENV-3. The Southeast Asian genotype of DENV-2 is more virulent and replicates at a higher titer as compared to the indigenous American DENV-2 genotype and leads to a more severe disease. Higher levels of dengue virus RNA are observed in in DHF patients. Moreover, subgenomic flavivirus RNA (sfRNA) can inhibit interferon-related translation<sup>19</sup>.

Human Leukocyte Antigen type 1 (HLA-1) causes increased severity of dengue while HLA class type 2 (HLA-2) and HLA-DR4 are protective against DHF. Factors not related to HLA include Fcγ receptor II (FcγRII, CD32), Vitamin D receptor (VDR) and Human Platelet Antigen (HPA). Othergenetic polymorphisms such as the tumor necrosis factor (TNF)-308A allele increased the risk of severe dengue whereas the tumor necrosis factor (TNF)- 238 allele was protective<sup>21</sup>.

# *2.6 Cytokines*

The excessive production of cytokines like TNFalpha and interferon-γ from memory T cells during secondary infection activate polymorphonuclear monocytes. These then produce more inflammatory cytokines that increase vascular permeability<sup>22</sup>. Cytokines including CCL2, CCL5, CCL20 and CXCL1 are found to be raised in dengue. They disrupt lipid metabolism and alter membrane permeability. HMGB-1 and ICAM-1 were found to be correlated to the severity of the disease. HMGB-1 increases amounts of other inflammatory cytokines while ICAM-1 is an intercellular adhesion molecule<sup>23</sup>.

# **3. Clinical Findings**

*3.1 Classification of Dengue*

According to 1997 WHO guidelines, the classification of dengue is divided into  $24$ :

- Dengue fever (DF): Fever associated with two or more of the given features: headache, myalgia, retro-orbital pain, arthralgia, rash, leucopenia or hemorrhagic indications.
- Dengue hemorrhagic fever (DHF): Hemorrhages, low platelet counts and plasma leakage.
- Dengue shock syndrome (DSS): Hemorrhages, low platelet count, plasma leakage along with circulatory failure i.e. tachycardia, hypotension, pedal oedema, shortness of breath.

The WHO guidelines in 2009 revised the criteria and divided dengue into non-severe and severe forms $^{25}$ . Non-severe dengue is further divided into those without warning signs and those with warning signs. Manifestations of non-severe dengue without warning signs include fever with nausea and vomiting, rash, body pain, positive tourniquet test or leucopenia. Warning signs included abdominal pain, mucosal bleeding, vomiting, hepatomegaly and findings of fluid accumulation. Severe dengue was presented as hepatomegaly, abdominal pain, bleeding, pleural effusion, ascites, increased hematocrit, high AST levels and decreased platelet count. This implied impending plasma leakage leading to decreased tissue perfusion and shock $26$ .

# *3.2 Clinical Course*

The clinical course of the disease follows three stages $^{27}$ :

- Febrile phase: It lasts 2 to 7 days. It presents with sudden high fever, headache, arthralgia, myalgia, facial flushing, blanching macular skin rash, sore throat, nausea, vomiting, elevated liver enzymes and anorexia<sup>28</sup>. The temperature pattern is biphasic.
- Critical phase. It does not occur in patients with DF but in those with DHF. It is marked by increased capillary permeability with increased hematocrit. Decreasing platelet count and progressive leukopenia occur before significant plasma loss. Significant plasma leakage leads to hypovolemic shock which presents with warning signs like subnormal temperature, sweaty hands and rapid thread-like pulse. Extended shock causes organ dysfunction, metabolic acidosis and

disseminated intravascular coagulation thus, leading to severe dengue<sup>29</sup>.

Recovery phase: Lasts for 48 to 72 hours. It is characterized by reabsorption of extravascular fluid back into the vessels. The appetite is improved, gastrointestinal symptoms are relieved, and bradycardia, diuresis, pruritus and a generalized rash are seen (islands of white in a sea of red $)^{30}$ .

Expanded dengue syndrome occurs when multiple organs are involved other than the plasma leakage. They do not fit into DHF or DSS. Organ systems involved include cardiovascular, kidneys, hematological, and gastrointestinal. Other than these, rhabdomyolysis and upper limb compartment syndrome have also been reported in several cases $^{31}$ .

# **4. Complications of Dengue Fever**

Dengue fever, caused by the dengue virus and transmitted by Aedes mosquitoes, generally causes a mild, self-limiting illness which resolves on its own with supportive care. However, in some cases, especially in areas where it is endemic, the illness can rapidly progress to severe dengue which can be recognised by persistent vomiting, abdominal pain and bleeding under the skin (Table 1).

# *4.1 Neurological complications*

Neurological complications of dengue, although incredibly rare, can occur in severe cases and are increasingly being recognized. Neurological syndromes of dengue, a neurotropic virus, occur due to direct invasion of the virus or due to the systemic effects caused by the infection $32$ .

Most common neurological complications of dengue are encephalopathy and encephalitis<sup>33</sup>. Encephalopathy occurs in severe cases of dengue and signifies generalized brain dysfunction. It presents with altered levels of consciousness, seizures, confusion and irritability and may be associated with metabolic disturbances in the body, hypoxia or direct viral involvement. Similarly, encephalitis occurs due to direct invasion of the virus into the brain tissue which causes inflammation of the brain, resulting in symptoms such as seizures, headaches, and focal neurologic deficits. Among the rare neurologic complications of dengue is acute transverse myelitis (ATM) characterized by inflammation of the spinal cord and typically presents with sensory deficits,

motor weakness and dysfunction of the bladder and the bowel<sup>34</sup>. Sudden neurological deficits including weakness in one half of the body, slurred speech and loss of consciousness can occur due to cerebral hemorrhage, the risk of which is increased in dengue due to impaired blood clotting<sup>34</sup>. Guillain Barre, an autoimmune disease can occur following the viral infection and it presents with ascending paralysis, starting from the legs and progressively moving upwards $35$ . In some cases, dengue fever also has longterm neurological sequelae leading to cognitive impairment, memory deficits and altered behavior.

#### *4.2 Gastrointestinal complications*

Gastrointestinal complications of dengue include gastrointestinal bleeding, intestinal perforation, acute pancreatitis, hepatitis and in severe cases, fulminant hepatic failure. Among the early signs of dengue fever are nausea and vomiting which may persist throughout illness and may cause dehydration if fluids are not adequately replenished. Abdominal pain in dengue fever may be due to inflammation of the gastrointestinal system, liver failure or collection of fluid in the abdomen. Gastrointestinal bleeding is a manifestation of dengue hemorrhagic fever or dengue shock syndrome and indicates severe progression of the disease<sup>36</sup>. It may present with hematemesis or melena, blood in vomit or black tarry stools, respectively. If severe, excessive gastrointestinal bleeding can cause intestinal perforation which is a surgical emergency and requires urgent exploratory surgery to prevent the development of peritonitis and septic shock. Involvement of the pancreas and gallbladder is rare in dengue infection but it may occur in severe cases, manifested as pancreatitis and acalculous cholecystitis, respectively<sup>37,38</sup>. Hepatic involvement in dengue is signified by elevated liver enzymes, jaundice, enlarged liver and accumulation of fluid in the abdominal cavity. Hepatic dysfunction occurs due to direct viral damage to the hepatic cells or secondary effects of response to the virus. Hepatic involvement due to dengue infection is characterized by higher levels of AST as compared to ALT due to the release of AST from hepatocytes damaged by the dengue infection <sup>39</sup>. Liver damage was most associated with serotypes DENV-3 and DENV-4 and was more common in women<sup>40</sup>.

# *4.3 Cardiac complications*

Cardiac complications of dengue fever are rare and include myocarditis, arrhythmias, pericardial

effusion, acute coronary syndrome (ACS), hypotension and shock. Arrhythmias associated with DHF have been reported and are due to the disruption of the normal electrical activity of the heart. Arrhythmias associated with dengue fever include atrioventricular block, atrial fibrillation, tachycardia and bradycardia, which manifest as irregular heartbeats<sup>41</sup>. Arrhythmias generally resolve with the infection but can further increase the risk of complications such as stroke or heart failure. Another commonly reported cardiac complication of dengue fever is myocarditis which is due to inflammation of the heart muscle and presents with shortness of breath, chest pain and fatigue<sup>42</sup>. Pericardial involvement in dengue fever involves pericardial effusion and pericarditis which are due to inflammation of the pericardium, the surrounding outer covering of the heart. Pericarditis is characterized by sharp stabbing pain in the chest and pericardial effusion may present with signs of cardiac tamponade if the amount of fluid around the heart is significant. In severe cases, individuals may suffer from long-term cardiac complications which include persistent arrhythmias and chronic dysfunction of the heart muscle known as cardiomyopathy.

# *4.4 Respiratory complications*

In severe cases, the progression of dengue fever can lead to the development of acute respiratory distress syndrome (ARDS). ARDS is a life-threatening condition and is due to severe inflammation of the lung and concomitant impaired oxygenation <sup>41</sup>. ARDS is an emergency and can lead to respiratory failure, requiring ventilatory support. Severe dengue can lead to increased vascular permeability leading to extravasation of fluid (pulmonary edema) which impairs gas exchange and causes difficulty breathing and distress. The respiratory distress in dengue may be increased due to exacerbation of asthma, development of viral or bacterial pneumonia because of the impaired immune system, and accumulation of fluid in pleural space leading to pleural effusion. In a few cases, particularly in the pediatric population, dengue fever can cause inflammation of small airways leading to the development of bronchiolitis which presents with wheezing and signs of respiratory distress.

# *4.5 Hematological complications*

The most prominent features of dengue fever, particularly in severe cases are the hematological

complications. The hallmark hematological complication of dengue is thrombocytopenia, and decreased platelet count, which is due to both decreased production and increased destruction of platelets. Thrombocytopenia leads to bleeding tendencies and predisposes a patient to bleed of various types, such as petechiae, ecchymosis and more severe life-threatening manifestations of hemorrhage. In severe cases of dengue, due to endothelial injury which affects hemostasis, there is concomitant activation of coagulation due to the release of cytokines and fibrinolysis due to the formation of fibrin. A relative imbalance in the coagulation system can lead to disseminated intravascular coagulation which can cause widespread thrombosis and bleeding. Microvascular thrombosis can cause multi-organ ischemia and eventually lead to failure<sup>43,44</sup>. Dengue fever can cause suppression of the bone marrow leading to exacerbation of thrombocytopenia, development of anemia and dysfunction of the immune system. Another hematological complication is the development of hypovolemic shock the development of which is contributed by the leakage of plasma leading to hemoconcentration.

# *4.6 Renal complications*

Hypovolemia due to plasma leakage, decreased blood supply to the kidneys and direct viral effect can lead to impairment of the kidney function and development of acute kidney injury (AKI)<sup>45</sup>. AKI leads to the retention of waste products and fluid imbalance in the body. Damage to the kidney can

**Table 2. Complications of Dengue Fever41,43,45**

affect the filtration barrier and lead to proteinuria and severe bleeding and thrombocytopenia can lead to hematuria. Formation of small blood clots, as seen in severe cases of dengue, known as thrombotic microangiopathy can lead to impairment of kidney function and AKI. Electrolyte imbalances such as hyponatremia, hyperkalemia and acid-base abnormalities can further deteriorate kidney function and lead to renal complications. In cases of recurrent severe dengue or those with AKI, there is a risk of long-term kidney damage and the eventual development of chronic kidney disease.

# **5. Treatment**

# *5.1 Fluid Management*

Patients can be divided into four distinct categories (adapted from WHO, 2012).

- A normal heart rate and blood pressure of more than 20 mmHg.
- A pulse pressure or hypotension of less than 20 mmHg.
- Shock.
- Shock despite crystalloid solution fluid resuscitation.

The main form of therapy for groups 1-3 is crystalloid solution, which has certain limitations due to its increased capacity to leak into the third space, which may result in pulmonary edema, ascites, or pleural effusion. According to a study conducted on Vietnamese children (Wills et al., 2005), severe shock



should be treated with starch rather than dextran, whereas moderately severe shock should be treated with fluid therapy using Ringer's lactate solution<sup>46</sup>. Colloids have the potential to expand volume in addition to the actual fluid volume infused. Colloid molecules encourage fluid retention in the intravascular compartment and raise plasma oncotic pressure. The average molecular weight of the colloid molecule controls the effect's size, while circulation retention time controls how long the effect lasts. The plasma-volume-expanding capacity of crystalloids is correlated with sodium content. In principle, Ringer's lactate (131 mM) may not perform as well as NaCl 0.9% (154 mM) $47,48$ . Furthermore, there is a chance that infusing significant amounts of Ringer's lactate will exacerbate tissue acidosis and lactate buildup. When treating children with DSS, Wills et al. tested Ringer's lactate with two colloids and found no discernible benefit of the colloid over lactate infusion<sup>49</sup>.

#### *5.2 Drugs targeting dengue proteins*

#### *5.2.1 Entry fusion inhibitors*

The E protein, which binds to a variety of cellular receptors, is the main method by which the Dengue Virus (DENV) penetrates host cells. The virus is internalised by clathrin-mediated endocytosis once it has been bound. The E protein experiences conformational changes in the acidic environment of the cell, exposing the fusion loop and promoting membrane fusion, which permits viral RNA to enter the cytoplasm. In the interim between further optimisation and clinical research, membrane fusion inhibitors that target various viral protein areas have been created and are being assessed for possible therapeutic application<sup>50</sup>.

#### *5.2.2 Replication and transcription inhibitors*

Balapiravir, which is generated from the nucleoside analogue R1479, has been shown in a randomized, double-blind, placebo-controlled experiment to be effective in treating dengue virus infection by selectively targeting the RNA-dependent RNA polymerase (RdRP) domain of the viral NS5 protein. When balapiravir was given to adult dengue patients within 48 hours of the onset of the disease, it did not considerably lower viremia, NS1 antigenemia, or fever clearance time in comparison to the placebo group. The trial found no significant differences in whole blood transcriptional patterns or plasma cytokine profiles between the treatment and placebo groups, despite the medication being well tolerated. The trial's overall failure to establish balapiravir as a viable treatment option for dengue virus infection highlights the difficulties in creating antiviral



**Figure 2. Fluid management for Dengue47,48**

treatments that specifically target conserved viral proteins like NS5<sup>51</sup>.

#### *5.2.3 Methyltransferase inhibitors*

Because of its critical function in RNA synthesis and capping, which are necessary for viral stability and immune evasion, dengue virus NS5 is a prime target for treatments. Although specific inhibitors for the NS5 methyltransferase domain have been found, specificity and toxicity issues have made clinically meaningful capping inhibitors difficult to find. The review suggests creative enzyme engineering techniques to create therapeutic proteins that target NS5 to remedy this. Viral RNA may be more susceptible to host immune detection if engineered methyltransferases improve viral RNA binding and cause aberrant methylation. By lowering viral load and boosting immune response, these modified proteins may be administered using cutting-edge technologies like nanoparticles or chimeric proteins, potentially leading to better dengue treatment outcomes<sup>52</sup>.

#### *5.2.4 RNA-dependent RNA polymerase inhibitors*

Ivermectin, a well-known antiparasitic drug, has shown promise against dengue virus in vitro by inhibiting host nuclear import proteins crucial for the nuclear localization of the dengue NS5 protein, which has RNA-dependent RNA polymerase (RdRp) function. A phase  $2/3$  randomized, double-blind, placebo-controlled trial investigated the efficacy of ivermectin at a once-daily dose of 400 μg/kg for 2–3 days in adult dengue patients<sup>53</sup>. The study found that while ivermectin treatment led to faster clearance of NS1 antigenemia, there was no significant difference in viremia, viral clearance, or any beneficial clinical outcomes such as fever, dengue hemorrhagic fever (DHF) incidence, hospitalization, pleural effusion, hemoconcentration, or fluid requirements. In order to assess ivermectin's potential as a dengue treatment, more study is required to comprehend its pharmacodynamics and mechanism of action with regard to NS1, as elevated NS1 levels are a risk factor for DHF.

# *5.2.5 Nucleoside analog (NITD008)*

Both in vitro and in vivo, NITD-008, a nucleoside inhibitor that targets RNA-dependent RNA polymerase, demonstrated a potent inhibitory impact against all four DENV serotypes. However, the acute

renal toxicity seen in preclinical tests put a stop to its  $d$ evelopment $54$ .

# *5.2.6 Helicase inhibitors*

The NS3 protein's C-terminal domain, which spans amino acids 180–618, performs the activities of a non-processive NTPase/helicase. This helicase is a member of the "SF2" helicase superfamily. For DENV, YFV, and JEV, helicase/NTPase activity and crystal structures have been described. Between subdomains 1 and 2, where ATP substrate binding predominates, is the location of the ATPase active  $site^{55}$ .

# *5.2.7 Protease inhibitors*

Potential protease inhibitors for dengue virus NS2B/NS3 protease (PR) include diaryl (thio)ethers. With low micromolar IC50 values, benzothiazole derivatives selectively and noncompetitively suppress the dengue virus PR serotypes 2 and 3 in vitro and in cells. Specifically, these drugs target DENV replication while leaving HCV and HIV-1 unaffected. According to molecular docking, binding occurs at an allosteric location. Cell-based experiments demonstrate their potential to limit dengue virus multiplication at low or submicromolar concentrations by confirming their inhibitory ecacy<sup>56</sup>.

# *5.2.8 NS4B inhibitor*

As a direct-acting treatment for dengue virus, Janssen Pharmaceuticals' NS4B inhibitor JNJ-A07 has demonstrated encouraging preclinical results. JNJ-A07, which is derived from a ketoindole molecule, works by severing the link between the NS4B and NS3 proteins to prevent the development of a DENV replication complex. In vitro, it demonstrates antiviral efficacy against several clinical isolates including all four dengue serotypes. Even with a delayed start of therapy, it quickly lowers viremia and the amount of virus in organs in murine models and lowers proinflammatory cytokine level while raising survival rates<sup>57</sup>. However, more research is required to determine any potential negative consequences, such as decreased antibody production that could result in severe dengue in subsequent infections. Phase 2 clinical trials are now being conducted on JNJ-64281802, an analogue of JNJ-A07, for dengue prophylaxis in healthy individuals and therapeutic usage in patients with confirmed dengue fever. The purpose of these trials is to evaluate its safety and effectiveness in actual use.

*5.2.9 Drugs targeting host factors required by DENV to complete its life cycle*

#### *5.2.9.1 Chloroquine*

The major aim of clinical studies for the widely used medication chloroquine (CQ), which is used to treat inflammatory illnesses such as lupus and rheumatoid arthritis, was the reduction of viral load. Research conducted in vitro indicates that CQ obstructs the pHdependent phases of viral replication, meaning it prevents the growth of coronaviruses, retroviruses, and flaviviruses. CQ suppresses TNF-α and IL-6 production, which in turn prevents endosomal fusion and furin-dependent viral maturation during dengue virus (DENV) infection. But in a randomized, doubleblind, placebo-controlled study, 307 adult Vietnamese patients infected with DENV were shown to have a longer viremia duration and no discernible decrease in the development of severe dengue hemorrhagic fever (DHF) in those treated with CQ. Additionally, there was a noticeable increase in vomiting that was linked to CQ treatment. A smaller, more recent trial on 37 dengue-positive patients revealed that although CQ treatment lessened the intensity of pain symptoms, it was unable to shorten the length of the illness or the intensity of the fever. These results raise questions regarding the possible negative effects of CQ and point to its limited effectiveness in treating dengue infection<sup>58</sup>

# *5.2.9.2 Elgosiver*

The alpha-glucosidase antagonist celgosivir protected mice against a deadly DENV challenge and showed abnormalities in the folding and ensnaring of viral NS1 in the endoplasmic reticulum. A phase 1b randomized placebo-controlled trial could not clearly demonstrate a benefit in lowering viremia or the course of the disease, despite the fact that DENVinfected mice treated with celgosivir showed reduced viremia and a mortality benefit. Further research on the pharmacokinetics and dose of the medication revealed that raising the dosage could enhance the medication's effectiveness<sup>59</sup>.

#### *5.2.9.3. Statins*

A class of medications known as statins prevents the synthesis of cholesterol by blocking the enzyme HMG-CoA reductase. Statins have anti-inflammatory characteristics in addition to their ability to decrease cholesterol. Since membrane lipids are essential to the

Flavivirus life cycle, it was thought that statins would prevent the virus from replicating. This is corroborated by research showing statins have antiviral and anti-inflammatory qualities in a dengueinfected animal model. But a study evaluating lovastatin in DENV infection was unable to show any improvement in viremia or the duration of the illness<sup>60</sup>.

# *5.3 Alternative methods*

*5.3.1 Revolutionary Methods for Aiming at the Vector* The introduction of genetically modified male mosquitoes, which sterilize the wild-type female population and lower egg production and the size of the following generation's population that could be available for possible dengue virus transmission, is one of the new vector-control strategies<sup>61</sup>. An alternate tactic is to introduce strains of the obligatory intracellular bacteria Wolbachia into A. aegypti during the embryonic stage. Remarkably, Aegypti with Wolbachia infection exhibits partial resistance to dengue virus infection and can infiltrate native populations of A.aegypti, indicating the potential for inducing pervasive biologic immunity against dengue viruses in A. aegypti populations $62,63,64$ .

#### *5.3.2 Sofosbuvir*

Due to shortcomings with current interventions such as the Dengvaxia vaccination, research into dengue fever treatments has increased. Nucleoside inhibitors that target the dengue virus polymerase are one line of inquiry; one such inhibitor is sofosbuvir, a prodrug known to be anti-HCV. Its active metabolite, GS-461203, has been found to exhibit strong viral suppression in both in vitro and in silico investigations, as well as a high binding affinity to the dengue viral polymerase's catalytic motif. The study also highlights the need for particular liver enzymes to activate sofosbuvir into its active form, highlighting potential complications in its therapeutic use against dengue. These results provide encouraging opportunities for the development of successful anti dengue treatments by highlighting the need for additional study into sofosbuvir's effectiveness and potential as a DENV polymerase inhibitor in human subjects<sup>65</sup>.

#### *5.3.3 Carica papaya*

Plenty of companies are creating or formulating C. papaya leaf extract products that are sold in stores.

Rochway, Herbal Papaya, SidoMuncul Herbal, and more companies prepare supplements. They are utilizing fermentation, micronization, and liquid extraction in the formulation of supplements<sup>66.</sup> Numerous plant species may elevate platelet count. In several animal dengue fever models, C. papaya leaf extract has been shown to increase platelet count and shorten the clotting time in thrombocytopenic rats<sup>67,68</sup>. When fed orally to AG129 dengue-infected mice, freeze-dried C. papaya leaves also elevated plasma monocyte chemoattractant protein-1 (MCP-1) levels during the peak of viremia, revealing the potential immunomodulatory activity of this plant during DENV infection $69,70$ .

#### *5.3.4 Azadirachta indica*

A real-time polymerase chain reaction (RT-PCR) test using an aqueous extract of Azadirachta indica leaves at a maximum non-toxic concentration of 20– 30 mg/mL revealed complete inhibition of viral replication, as did the absence of virus-specific 511 bp amplicon and DENV-related symptoms in DENVinfected suckling mice $71,72$ .

#### *5.3.5 Vitamin E supplementation*

Clinical investigations have demonstrated the benefits of vitamin E administration in addition to standard therapy, as it can reduce liver function abnormalities and promote platelet count recovery during dengue illness<sup>73</sup>.

#### *5.4 Symptomatic management*

It is advised to give large amounts of oral fluids and administer paracetamol as needed for antipyretic treatment during the febrile period. Avoid using any other nonsteroidal anti-inflammatory medications<sup>74</sup>. The patient can be maintained at home with daily full blood counts if there is a nearby medical facility available to them. Severe prostration, early bleeding symptoms, or excessive vomiting or diarrhoea that causes dehydration are all signs that the patient needs to be admitted to the hospital for close supervision<sup>75</sup>.

# *5.4.1 Antiviral Research in cure of Dengue*

Since there are currently no effective treatments for dengue, research into the development of antiviral medicines has accelerated in recent years. Small molecule inhibitors have traditionally targeted proteins such as NS3 protease, NS3 helicase, NS4B, and NS5. The NS3 and NS5 proteins are considered to be the most important targets for the development of antivirals due to their essential enzymatic roles in the process of viral replication. While NS5 possesses methyltransferase (MTase) and RNA-dependent RNA polymerase (RdRp) activities,NS3 has a variety of enzymatic activities, such as serine protease, nucleoside triphosphatase (NTPase), 5′-RNA triphosphatase, and helicase activities<sup>76-78</sup>.

### *5.4.2 Corticosteroids*

Corticosteroids have been debated for their potential role in dengue shock syndrome and the early administration to prevent progression to severe illness; however, there is insuffiecnt evidence for the support of this claim..Both natural immunological responses of phagocytes and a wide variety of immune responses mediated by T and B cells are inhibited by corticosteroids. High-dose corticosteroids are helpful in a variety of immunological abnormalities, including autoimmune illnesses like systemic lupus erythematosus. Corticosteroids used in the early acute phase of dengue infection did not affect shock, plasma leakage, or the recovery of platelet count in dengue patients, according to a double-blind, placebocontrolled trial. Neither the kinetics of dengue virological indicators nor the amounts of plasma cytokines were altered by it.

# *5.4.3 Blood products*

A common characteristic of Dengue Virus (DENV) infection is thrombocytopenia, which is defined as a platelet count below  $150 \times 10^{9}/l$ . Thrombocytopenia usually occurs between days 3 and 8 after symptom onset. As the hematocrit rises and platelet counts decrease, the disease is moving towards its critical phase. During defervescence (days 3–6), nadir platelet counts are attained, and then there is a gradual, spontaneous rebound. Mucosal or petechiae bleeding is common and typically goes away on its own without treatment. To reduce the risk of bleeding, severe thrombocytopenia ( $\leq 20 \times 10^9/1$ ) may require rigorous bed rest, avoidance of nonsteroidal anti-inflammatory medications, and intramuscular injections. Although preemptive platelet transfusion is frequently used in the treatment of sepsis, there is no proof of its effectiveness in dengue infections.Severe bleeding, frequently in the gastrointestinal or vaginal tract, is linked to metabolic acidosis, protracted shock, liver or renal failure, and certain drugs. Prompt transfusion of platelets, fresh frozen plasma, and packed red blood cells may be

life-saving in such situations. However, because there are no long-term advantages and several serious concerns, including fluid overload and transfusionrelated problems, prophylactic platelet transfusion is not advised.

#### *5.4.4 Alternative Measures for Treating Dengue Patients' Bleeding*

Recombinant factor VIIa (rFVIIa) was shown to decrease bleeding temporarily in a research study including 25 children with dengue hemorrhagic fever (grade II–III) and active bleeding, but there was no overall effect. While there is some indication that intravenous anti-D globulin can improve platelet counts in dengue patients, there is not enough data to support the use of intravenous immunoglobulin (IVIg), interleukin-1 (IL-1), or tranexamic acid.

#### *5.4.5 Intravenous immunoglobulins*

One recognised treatment Recombinant factor VIIa (rFVIIa is intravenous immunoglobulins (IVIG). Platelet-associated IgG (PAIgG) autoantibodies are responsible for thrombocytopenia in ITP. Platelets coated with PAIgG are rapidly cleared by means of Fcγ receptors that are found on mononuclear phagocytic cells. IVIG most likely occurs through ligation of inhibitory receptors or competitive inhibition of Fcγ receptors.(79) Thirty six dengue patients participated in a randomized, controlled study to assess IVIG treatment in relation to the amount and frequency used to treat ITP over the course of four days. The effectiveness of IVIG in increasing platelet recovery was not demonstrated by the trial $^{80}$ .

# *5.4.6 Mast cell inhibitors*

The pathophysiology of dengue-vascular leakage and hemorrhage has been linked to mast cell (MC) activation, according to recent research by St John et al.(48)In a mouse model of DENV infection, it was demonstrated that activated MC releases a variety of proteases into the serum, most notably tryptase and chymase, which causes vascular integrity to be lost. The authors also demonstrated a correlation between serum chymase levels and dengue severity in people participating in a prospective research, which is in line with their observation in mice. Subsequently, the group demonstrated that, in spite of a little (but not statistically significant) rise in mouse viremia, MCstabilizing drugs, such as cromolyn, montelukast, and ketotifen, decreased vascular leakage in the wild-type mice model of DENV assault [67]. All of these findings point to the possibility of using MCs as therapeutic targets to reduce DENV pathogenesis $47$ .

# *6. Dengue Vaccine*

The lack of specific antiviral treatment against dengue has placed a significant emphasis on preventative measures which include vector control and most importantly, vaccination. In combating and adequately controlling this disease, the development of vaccines has been an essential focus.

# *6.1 Advancements in the vaccine development*

Considerable progress has been made in the development of vaccines against dengue in the last few decades. Most notably, the first licensed dengue vaccine known as Dengavaxia by San Pasteur's became approved in various countries where dengue is an epidemic. Dengvaxia is a live-attenuated vaccine and is formulated to provide protection and immunity against all four serotypes of dengue<sup>88</sup>).

Additional advancements have led to the development of alternatives to Dengvaxia and aim to become established as safer and more effective contenders while simultaneously addressing concerns related to the safety profile of Dengvaxia. These advancements have been domadey utilizing various advanced approaches such as viral vectors, recombinant proteins and DNA vaccines.

# *6.2 Types of vaccines*

# *6.2.1 Live attenuated vaccines*

Using weakened forms of viruses which are attenuated to reduce their virulence, live-attenuated vaccines can produce an immune response in the host. Sanofi Pasteur's Dengvaxia is an example of a live atta enuated vaccine and is approved to provide immunity against all four serotypes of dengue. Dengvaxia was developed by substituting the pRM/E RNAs of the yellow fever vaccine strain with equivalent sequences from the various serotypes. The vaccine has been licensed by several countries where dengue is endemic such as in Asia and Latin America88,89. Dengvaxia provides an immunity to the host which lasts for more than four years and clinical trials have revealed it to be more effective in individuals over the age of 9 (90).

# *6.2.2 Recombinant protein vaccines*

The production of recombinant protein vaccines ininvolveshe use of various genetic engineering techniques for the production of viral proteins, mainly the envelope proteins. These proteins are then modified to produce an immune response against dengue virus in the host. Several vaccine candidates have been developed utilizing this approach ancludincludingakeda Tetravalent Dengue Vaccine (TAK-003). In comparison to the more widely used live-attenuated vaccines, recombinant protein vaccines are capable of inducing a more fitting immune response while simultaneously lowering the incidence of antibody-dependent enhancement $91,92$ .

#### *6.2.3 Viral vector vaccines*

To stimulate immune response, viral vector vaccines use genetically engineered viruses such as adenovirus or chimeric viruses to deliver the dengue virus into the body. Examples include NIH the live attenuated teta-valent vaccine (TV003/TV005) and IMCB's chimeric dengue vaccine. Using a viral vector vaccine remains superior at inducing cellular immunity and has the potential to become better at causing robust humoral immune response<sup>93</sup>.

### *6.2.4 DNA vaccine*

Several dengue DNA vaccines are underway because they are stable and relatively easy to produce  $94,95$ .

# *6.3 Notable Examples*

#### *6.3.1 Dengue Virus (DENV) Vaccines Dengvaxia*

This technology facilitated the development of four chimeric YF-DEN viruses, which were utilized in the development of a tetravalent DENV vaccine. The vaccine is based on the 17D strain of yellow fever virus, where the pre-membrane and envelope proteins of yellow fever virus have been substituted by the corresponding genes from each of the four DENV serotypes. These serotypes were originally obtained from DENV isolates collected in Thailand and Indonesia between 1978 and 198819,20 After one injection of the vaccine, either at a high or low dose, in cynomolgus macaques, it produced seroconversion and significant neutralizing antibody responses against all four DENV serotypes and limited viremias in comparison to the parental DENV strains. It's interesting to note that 92% of the immunized monkeys were shielded from a challenge with wildtype DENV 1-4 by challenge experiments<sup>96</sup>

### *6.3.2 TV003/TV005*

Given the significance of untranslated regions (UTRs) in the replication of the DENV genome, the first attenuation method concentrated on removing 30 consecutive nucleotides  $(172-143)$  from the 3 -UTR of DENV-4 (rDEN4 $\Delta$ 30) in the TL2 stem-loop<sup>(97</sup>. For DENV-1, rDEN1∆30—a mutant lacking the same homologous genomic region—was also created. When confronted with wild types of DENV-1 and DENV-4, both mutants showed an attenuated phenotype as evidenced by their decreased infectivity and their ability to elicit robust neutralizing antibody responses in rhesus macaques, which coincided with the protection<sup>98</sup>).

Continued efforts towards development of tetravalent DENV vaccine included vrrating an attenuated DENV-2 component. This was achieved by utilizing rDEN4∆30 backbone to create two attenuated chimeric viruses: one with the membrane and envelope genes (rDEN2/4 ∆30 (ME)), and another with the capsid, membrane, and envelope genes (rDENV2/4 ∆30 (CME)) similarly substituted<sup>99</sup>.

#### *6.3.3 TAK-003 (DENVax)*

When scientists at Mahidol University in Bangkok, Thailand, identified a DENV-2 strain (DENV-2 16681) from a patient's serum, they began developing the DENVax vaccine in the late 1980s. The DENV-2 PDK-53-V strain was obtained by attenuating the DENV-2 16681 strain in primary dog kidney cells (PDK cells) through 53 serial passages. This strain differs from the parental DENV-2 PDK-53 strain in that it has an additional non-synonymous mutation in the NS3 gene, while attenuation-related mutations are present in the 5 ́UTR and NS1 gene. The DENV-2 PDK53-V strain was also the foundation for the development of the DENVax vaccines, exhibiting decreased neurovirulence in nursing mice and decreased replication rates in  $C6/36$  cells<sup>100,101</sup>.

#### *6.4 Challenges in the development of vaccine*

Despite the recent advancements and progress, the development of a vaccine against dengue faces many challenges. A significant challenge is the requirement of induction of simultaneous immunity against all four serotypes of dengue. Vaccine development is further complicated by the phenomenon of antibodydependent enhancement, in which prior infection with one serotype may amplify and exacerbate disease

upon subsequent infection with another serotype of dengue. Furthermore, safety concerns have been raised against Dengvaxia because of reports of severe dengue infection in vaccinated individuals without any prior history of exposure and infection. These reports have impacted the trust of the public in the dengue vaccination programs and have prompted regular scrutiny. Maintaining efficacy and addressing safety concerns at the same time is a critical challenge for the developers.

Additional challenges in vaccine development are posed by the complex epidemiology of the disease, with varying prevalence of serotypes in different populations and variable transmission dynamics. This requires tailored vaccination programs for different populations and equitable access to vaccines in endemic areas, particularly low- and middle-income countries where prevalence of dengue is the highest. This represents additional challenges to the infrastructure of the healthcare system, distribution and economic affordability.

# *6.5 Future of vaccine development*

Despite the multiple challenges, ongoing research has offered promising results to the development of safe and effective vaccine development coupled with a better understanding of the epidemiology and immunology of the disease that provide help in overcoming the different hurdles. Addressing these challenges requires collaboration and efforts of researchers, policymakers and healthcare professionals. Additionally, innovative strategies should be employed to minimize antibody-dependent enhancement, to increase immunogenicity and to provide cross-protective efficacy.

# **7. Conclusion**

In conclusion, this comprehensive review provides a deeper understanding of the complexities and multifaceted nature of this infectious disease through an exploration of its pathogenesis, clinical features and emerging insights. Amidst the challenges posed by the wide spectrum of clinical presentations and complications, ongoing research efforts have yielded valuable insights into the immunological mechanisms underlying the viral infection, role of host genetics in disease susceptibility and the development of therapeutic interventions. Additionally, the development of dengue vaccine offers hope for

reducing the burden of dengue fever worldwide. A review on dengue is essential for clinicians and healthcare professionals to stay updated and informed about the pathogenesis, treatment protocols and prevention strategies i.e., vaccines.

# **Acknowledgements**

This review study received no grant from any funding agency in the public, commercial or not-for-profit sectors. Ethical approval was not required for this study.

PO conceptualized the research idea and framed the title. She determined the exclusion and inclusion criteria for the papers used by the other authors in writing the sections of the article. She wrote the abstract. GNEH wrote the introduction and was in charge of compiling the data, reviewing, supervising and preparing the manuscript. GNEH, RHA and SM were in charge of editing the content of the paper along with the other authors. AM, Apurva P, Aayushi P, SR and RHA made the diagrams and tables that are used in the article. All the authors significantly contributed to writing the sections of the article and approved the manuscript.

# **Conflict of Interest**

The authors would like to disclose no conflict of interest related to the publication of the article.

# **Acknowledgment**

This review did not receive any sort of funding from any institution or any individual. No acknowledgments which the authors would like to mention.

# **References**

- 1. Leong AS, Wong KT, Leong TY, Tan PH, Wannakrairot P. The pathology of dengue hemorrhagic fever. InSeminars in diagnostic pathology 2007 Nov 1 (Vol. 24, No. 4, pp. 227-236). WB Saunders.
- 2. Byard RW. Lethal dengue virus infection: a forensic overview. The American journal of forensic medicine and pathology. 2016 Jun 1;37(2):74-8.
- 3. Kayesh ME, Khalil I, Kohara M, Tsukiyama-Kohara K. Increasing dengue burden and severe dengue risk in Bangladesh: an overview. Tropical Medicine and Infectious Disease. 2023 Jan 3;8(1):32.
- 4. Ganeshkumar P, Murhekar MV, Poornima V, Saravanakumar V, Sukumaran K, Anandaselvasankar A, John D, Mehendale SM. Dengue infection in India:

A systematic review and meta-analysis. PLoS neglected tropical diseases. 2018 Jul 16;12(7):e0006618.

- 5. Wang WH, Urbina AN, Chang MR, Assavalapsakul W, Lu PL, Chen YH, Wang SF. Dengue hemorrhagic fever–A systemic literature review of current perspectives on pathogenesis, prevention and control. Journal of Microbiology, Immunology and Infection. 2020 Dec 1;53(6):963-78.
- 6. Harapan H, Michie A, Sasmono RT, Imrie A. Dengue: a minireview. Viruses. 2020 Jul 30;12(8):829.
- 7. Gwee XW, Chua PE, Pang J. Global dengue importation: a systematic review. BMC infectious diseases. 2021 Dec;21:1-0.
- 8. Ganeshkumar P, Murhekar MV, Poornima V, Saravanakumar V, Sukumaran K, Anandaselvasankar A, John D, Mehendale SM. Dengue infection in India: A systematic review and meta-analysis. PLoS neglected tropical diseases. 2018 Jul 16;12(7):e0006618.
- 9. Wang WH, Urbina AN, Chang MR, Assavalapsakul W, Lu PL, Chen YH, Wang SF. Dengue hemorrhagic fever–A systemic literature review of current perspectives on pathogenesis, prevention and control. Journal of Microbiology, Immunology and Infection. 2020 Dec 1;53(6):963-78.
- 10. Kok BH, Lim HT, Lim CP, Lai NS, Leow CY, Leow CH. Dengue virus infection - a review of pathogenesis, vaccines, diagnosis and therapy. Virus Res. 2023 Jan 15;324:199018. doi: 10.1016/j.virusres.2022.199018. Epub 2022 Dec 7. PMID: 36493993; PMCID: PMC10194131.
- 11. Martina BE, Koraka P, Osterhaus AD. Dengue virus pathogenesis: an integrated view. Clin Microbiol Rev. 2009 Oct;22(4):564-81. doi: 10.1128/CMR.00035-09. PMID: 19822889; PMCID: PMC2772360.
- 12. Nikin-Beers R, Ciupe SM. Modelling original antigenic sin in dengue viral infection. Math Med Biol. 2018 Jun 13;35(2):257-272. doi: 10.1093/imammb/dqx002. PMID: 28339786.
- 13. Wong JM, Adams LE, Durbin AP, Muñoz-Jordán JL, Poehling KA, Sánchez-González LM, Volkman HR, Paz-Bailey G. Dengue: A Growing Problem With New Interventions. Pediatrics. 2022 Jun 1;149(6):e2021055522. doi: 10.1542/peds.2021- 055522. PMID: 35543085.
- 14. Narayan R and Tripathi S (2020) Intrinsic ADE: The Dark Side of Antibody Dependent Enhancement During Dengue Infection. Front. Cell. Infect. Microbiol. 10:580096. doi: 10.3389/fcimb.2020.580096
- 15. Harapan H, Michie A, Sasmono RT, Imrie A. Dengue: A Minireview. Viruses. 2020 Jul 30;12(8):829. Doi: 10.3390/v12080829. PMID: 32751561; PMCID: PMC7472303.
- 16. Alcalá AC, Ludert JE. The dengue virus NS1 protein; new roles in pathogenesis due to similarities with and affinity for the high-density lipoprotein (HDL)? PLoS Pathog. 2023 Aug 24;19(8):e1011587. Doi: 10.1371/journal.ppat.1011587. PMID: 37616216; PMCID: PMC10449462.
- 17. Jayathilaka D, Gomes L, Jeewandara C, Jayarathna GSB, Herath D, Perera PA, Fernando S, Wijewickrama A, Hardman CS, Ogg GS, Malavige GN. Role of NS1 antibodies in the pathogenesis of acute secondary dengue infection. Nat Commun. 2018 Dec 7;9(1):5242. Doi: 10.1038/s41467-018-07667-z. PMID: 30531923; PMCID: PMC6286345.
- 18. Lebeau G, Lagrave A, Ogire E, Grondin L, Seriacaroupin S, Moutoussamy C, Mavingui P, Hoarau JJ, Roche M, Krejbich-Trotot P, Desprès P, Viranaicken W. Viral Toxin NS1 Implication in Dengue Pathogenesis Making It a Pivotal Target in Development of Efficient Vaccine. Vaccines (Basel). 2021 Aug 25;9(9):946. Doi: 10.3390/vaccines9090946. PMID: 34579183; PMCID: PMC8471935.
- 19. Bhatt P, Sabeena SP, Varma M, Arunkumar G. Current Understanding of the Pathogenesis of Dengue Virus Infection. Curr Microbiol. 2021 Jan;78(1):17- 32. Doi: 10.1007/s00284-020-02284-w. Epub 2020 Nov 24. PMID: 33231723; PMCID: PMC7815537.
- 20. Lin CF, Wan SW, Chen MC, Lin SC, Cheng CC, Chiu SC et al. Liver injury caused by antibodies against dengue virus nonstructural protein 1 in a murine model. Lab Invest. 2008;88(10):1079–1089. [PubMed] [Google Scholar] [Ref list]
- 21. Murugesan A, Manoharan M. Dengue Virus. Emerging and Reemerging Viral Pathogens. 2020:281–359. Doi: 10.1016/B978-0-12-819400- 3.00016-8. Epub 2019 Sep 20. PMCID: PMC7149978.
- 22. Soe HJ, Manikam R, Raju CS, Khan MA, Sekaran SD. Correlation of host inflammatory cytokines and immune-related metabolites, but not viral NS1 protein, with disease severity of dengue virus infection. PLoS One. 2020 Aug 7;15(8):e0237141. Doi: 10.1371/journal.pone.0237141. PMID: 32764789; PMCID: PMC7413495.
- 23. Glasner DR, Puerta-Guardo H, Beatty PR, Harris E. The Good, the Bad, and the Shocking: The Multiple Roles of Dengue Virus Nonstructural Protein 1 in Protection and Pathogenesis. Annu Rev Virol. 2018 Sep 29;5(1):227-253. Doi: 10.1146/annurev-virology-

101416-041848. Epub 2018 Jul 25. PMID: 30044715; PMCID: PMC6311996.

- 24. Raafat N, Blacksell SD, Maude RJ. A review of dengue diagnostics and implications for surveillance and control. Trans R Soc Trop Med Hyg. 2019 Nov 1;113(11):653-660. Doi: 10.1093/trstmh/trz068. PMID: 31365115; PMCID: PMC6836713.
- 25. Hadinegoro SR. The revised WHO dengue case classification: does the system need to be modified? Paediatr Int Child Health. 2012 May;32 Suppl 1(s1):33-8. Doi: 10.1179/2046904712Z.00000000052. PMID: 22668448; PMCID: PMC3381438.
- 26. Tsheten T, Clements ACA, Gray DJ, Adhikary RK, Furuya-Kanamori L, Wangdi K. Clinical predictors of severe dengue: a systematic review and meta-analysis. Infect Dis Poverty. 2021 Oct 9;10(1):123. DOI: 10.1186/s40249-021-00908-2. PMID: 34627388; PMCID: PMC8501593.
- 27. Kalayanarooj S. Clinical Manifestations and Management of Dengue/DHF/DSS. Trop Med Health. 2011 Dec;39(4 Suppl):83-7. Doi: 10.2149/tmh.2011- S10. Epub 2011 Dec 22. PMID: 22500140; PMCID: PMC3317599.
- 28. Schaefer TJ, Panda PK, Wolford RW. Dengue Fever. [Updated 2022 Nov 14]. In: StatPearls [Internet].

Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: https://www.ncbi.nlm.nih.gov/books/NBK430732/

- 
- 29. Dengue: Guidelines for Diagnosis, Treatment, Prevention and Control: New Edition. Geneva: World
- Health Organization; 2009. 2, Clinical Management and Delivery of Clinical Services. Available from: https://www.ncbi.nlm.nih.gov/books/NBK143161/
- 30. Tayal A, Kabra SK, Lodha R. Management of Dengue: An Updated Review. Indian J Pediatr. 2023 Feb;90(2):168-177. Doi: 10.1007/s12098-022-04394- 8. Epub 2022 Dec 27. PMID: 36574088; PMCID: PMC9793358.
- 31. Arif A, Abdul Razzaque MR, Kogut LM, Tebha SS, Shahid F, Essar MY. Expanded dengue syndrome presented with rhabdomyolysis, compartment syndrome, and acute kidney injury: A case report. Medicine (Baltimore). 2022 Feb 18;101(7):e28865. Doi: 10.1097/MD.0000000000028865. PMID: 35363190; PMCID: PMC9281986.
- 32. Soares CN, Cabral-Castro MJ, Peralta JM, Freitas MRG, Puccioni-Sohler M. Meningitis determined by oligosymptomatic dengue virus type 3 infection: Report of a case. International Journal of Infectious Diseases. 2010;14(2). DOI: 10.1016/j.ijid.2009.03.016
- 33. Trivedi S, Chakravarty A. Neurological Complications of Dengue Fever. Vol. 22, Current Neurology and Neuroscience Reports 2022.
- 34. Jafri L, Hameed S, Shakeel E, Shaikh N, Kanwar D. Transverse myelitis with positive dengue virus serology: a case report. Egyptian Journal of Neurology, Psychiatry and Neurosurgery. 2022;58(1). DOI: 10.1186/s41983-022-00564-9
- 35. Puccioni-Sohler M, Rosadas C, Cabral-Castro MJ. Neurological complications in dengue infection: A review for clinical practice. Vol. 71, Arquivos de Neuro-Psiquiatria 2013.
- 36. Dalugama C, Shelton J, Ekanayake M, Gawarammana IB. Dengue fever complicated with Guillain-Barré syndrome: A case report and review of the literature. J Med Case Rep. 2018;12(1). DOI: 10.1186/s13256- 018-1626-y
- 37. Chien Y-W, Chuang H-N, Wang Y-P, Perng GC. Risk of gastrointestinal bleeding after acute dengue virus infection. International Journal of Infectious Diseases. 2020;101. DOI: 10.1016/j.ijid.2020.11.054
- 38. Gulati S, Maheshwari A. Atypical manifestations of dengue. Vol. 12, Tropical Medicine and International Health 2007.
- 39. Nguyen THT, Nguyen HQ. A Rare Case of Acute Pancreatitis as Dengue Complication. Case Rep Infect Dis. 2023;2023. DOI: 10.1155/2023/2619785
- 40. de Souza LJ, Gonçalves Carneiro H, Souto Filho JTD, Ferreira de Souza T, Azevedo Côrtes V, Neto CG, et al. Hepatitis in dengue shock syndrome. Braz J Infect Dis. 2002;6(6). DOI: 10.1590/s1413-86702002000600010
- 41. Naya PA, Permatananda K, Ayu P, Kasih Permatananda N. Dengue Complication in Children Article in. International Journal of Science and Research. 2020;
- 42. Lee IK, Lee WH, Liu JW, Yang KD. Acute myocarditis in dengue hemorrhagic fever: A case report and review of cardiac complications in dengueaffected patients. International Journal of Infectious Diseases. 2010;14(10). DOI: 10.1016/j.ijid.2010.06.011
- 43. Van Gorp ECM, Setiati TE, Mairuhu ATA, Suharti C, Ten Cate H, Dolmans WMV, et al. Impaired fibrinolysis in the pathogenesis of dengue hemorrhagic fever. J Med Virol. 2002;67(4). DOI: 10.1002/jmv.10137
- 44. C.A. L, W.G. P, S. B, M. D, M.A. K, R. K, et al. Clinical perspectives of emerging pathogens in bleeding disorders. Vol. 367, Lancet 2006.
- 45. Bignardi PR, Pinto GR, Boscarioli MLN, Lima RAA, Delfino VDA. Acute kidney injury associated with

dengue virus infection: a review. Vol. 42, Brazilian Journal of Nephrology 2022.

- 46. Wongsa A. Fluid and hemodynamic management in severe dengue. Southeast Asian J Trop Med Public Health. 2015 Jan 1;46(Suppl 1):123-7.
- 47. St John AL, Rathore AP, Raghavan B, Ng ML, Abraham SN. Contributions of mast cells and vasoactive products, leukotrienes and chymase, to dengue virus-induced vascular leakage. elife. 2013 Apr 30;2:e00481.
- 48. St. John AL, Rathore AP, Yap H, Ng ML, Metcalfe DD, Vasudevan SG, Abraham SN. Immune surveillance by mast cells during dengue infection promotes natural killer (NK) and NKT-cell recruitment and viral clearance. Proceedings of the National Academy of Sciences. 2011 May 31;108(22):9190-5.
- 49. Pierson TC, Diamond MS. Vaccine development as a means to control dengue virus pathogenesis: do we know enough? Annual Review of Virology. 2014 Sep 29;1:375-98
- 50. De La Guardia C, Lleonart R. Progress in the identification of dengue virus entry/fusion inhibitors.

BioMed research international. 2014 Oct;2014.

- 51. Nguyen NM, Tran CN, Phung LK, Duong KT, Huynh HL, Farrar J, Nguyen QT, Tran HT, Nguyen CV, Merson L, Hoang LT. A randomized, double-blind placebo controlled trial of balapiravir, a polymerase inhibitor, in adult dengue patients. The Journal of infectious diseases. 2013 May 1;207(9):1442-50.
- 52. Vashishta K, Pandeya KC. A review on future perspectives in targeting the RNA capping in dengue virus by NS5 methyltransferase. Acta Scientifica Malaysia (ASM). 2023;7(1):17-22.
- 53. Zou J, Lee LT, Wang QY, Xie X, Lu S, Yau YH, Yuan Z, Geifman Shochat S, Kang C, Lescar J, Shi PY. Mapping the interactions between the NS4B and NS3 proteins of dengue virus. Journal of virology. 2015 Apr 1;89(7):3471-83.
- 54. Yin Z, Chen YL, Schul W, Wang QY, Gu F, Duraiswamy J, Kondreddi RR, Niyomrattanakit P, Lakshminarayana SB, Goh A, Xu HY. An adenosine nucleoside inhibitor of dengue virus.
- Proceedings of the National Academy of Sciences. 2009 Dec 1;106(48):20435-9.
- 55. Lim SP, Wang QY, Noble CG, Chen YL, Dong H, Zou B, Yokokawa F, Nilar S, Smith P, Beer D, Lescar J. Ten years of dengue drug discovery: progress and prospects. Antiviral research. 2013 Nov 1;100(2):500- 19.
- 56. Wu H, Bock S, Snitko M, Berger T, Weidner T, Holloway S, Kanitz M, Diederich WE, Steuber H, Walter C, Hofmann D. Novel dengue virus NS2B/NS3 protease inhibitors. Antimicrobial agents and chemotherapy. 2015 Feb;59(2):1100-9.
- 57. Wagstaff KM, Sivakumaran H, Heaton SM, Harrich D, Jans DA. Ivermectin is a specific inhibitor of importin  $\alpha/\beta$ -mediated nuclear import able to inhibit replication of HIV-1 and dengue virus. Biochemical Journal. 2012 May 1;443(3):851-6
- 58. Tricou V, Minh NN, Van TP, Lee SJ, Farrar J, Wills B, Tran HT, Simmons CP. A randomized controlled trial of chloroquine for the treatment of dengue in Vietnamese adults. PLoS neglected tropical diseases. 2010 Aug 10;4(8):e785.
- 59. Low JG, Sung C, Wijaya L, Wei Y, Rathore AP, Watanabe S, Tan BH, Toh L, Chua LT, Chow
- A, Howe S. Efficacy and safety of celgosivir in patients with dengue fever (CELADEN): a phase 1b, randomised, double-blind, placebo-controlled, proofof-concept trial. The Lancet infectious diseases. 2014 Aug 1;14(8):706-15.
- 60. Neufeldt CJ, Cortese M, Acosta EG, Bartenschlager R. Rewiring cellular networks by members of the Flaviviridae family. Nature Reviews Microbiology. 2018 Mar;16(3):125-42.
- 61. Wise de Valdez MR, Nimmo D, Betz J, Gong HF, James AA, Alphey L, Black WC 4th. Genetic elimination of dengue vector mosquitoes. Proc Natl Acad Sci U S A. 2011 Mar 22;108(12):4772-5. doi: 10.1073/pnas.1019295108. Epub 2011 Mar 7. PMID: 21383140; PMCID: PMC3064365Moreira LA, Iturbe-Ormaetxe I, Jeffery JA, Lu G, Pyke AT, Hedges LM, Rocha BC, Hall-Mendelin S, Day A, Riegler M, Hugo LE. A Wolbachia symbiont in Aedes aegypti limits infection with dengue, Chikungunya, and Plasmodium. Cell. 2009 Dec 24;139(7):1268-78.
- 62. Walker TJ, Johnson PH, Moreira LA, Iturbe-Ormaetxe I, Frentiu FD, McMeniman CJ, Leong YS, Dong Y, Axford J, Kriesner P, Lloyd AL. The w Mel Wolbachia strain blocks dengue and invades caged Aedes aegypti populations. Nature. 2011 Aug 25;476(7361):450-3.
- 63. Hoffmann AA, Montgomery BL, Popovici J, Iturbe-Ormaetxe I, Johnson PH, Muzzi F, Greenfield M, Durkan M, Leong YS, Dong Y, Cook H. Successful establishment of Wolbachia in Aedes populations to suppress dengue transmission. Nature. 2011 Aug 25;476(7361):454-7.
- 64. Gan CS, Lim SK, Chee CF, Yusof R, Heh CH. Sofosbuvir as treatment against dengue? Chemical biology & drug design. 2018 Feb;91(2):448-55.
- 65. Mandal SU, Jaiswal DV, Shiva K. A review on marketed Carica papaya leaf extract (CPLE) supplements for the treatment of dengue fever with thrombocytopenia and its drawback. International Journal of Pharmaceutical Research. 2020 Jul 1;12(3).
- 66. Patil S, Shetty S, Bhide R, Narayanan S. Evaluation of platelet augmentation activity of Carica papaya leaf aqueous extract in rats. Journal of Pharmacognosy and phytochemistry. 2013;1(5):57-60.
- 67. Arollado EC. Platelet augmentation activity of selected Philippine plants. International Journal of Pharmaceutical and Phytopharmacological Research. 2014 Feb 8;3(2).
- 68. Coloma AJ, Casilla AS, Estolero BH, Ulalan MG, Veloso GD, Man R. Thrombocytotic Efficacy of Tawa-Tawa, Papaya and MalunggayAmong Aspirin-Induced Laboratory Rabbits. ICHAMS Health Care J. 2015;5(1):8-16.
- 69. Saleh MS, Kamisah Y. Potential medicinal plants for the treatment of dengue fever and severe acute respiratory syndrome-coronavirus. Biomolecules. 2020 Dec 30;11(1):42.
- 70. Ichsyani M, Ridhanya A, Risanti M, Desti H, Ceria R, Putri DH, Sudiro TM, Dewi BE. Antiviral effects of Curcuma longa L. against dengue virus in vitro and in vivo. InIOP Conference Series: Earth and Environmental Science 2017 Dec 1 (Vol. 101, No. 1, p. 012005). IOP Publishing.
- 71. Parida MM, Upadhyay C, Pandya G, Jana AM. Inhibitory potential of neem (Azadirachta indica Juss) leaves on dengue virus type-2 replication. Journal of ethnopharmacology. 2002 Feb 1;79(2):273-8.
- 72. Chathurangana PW, Samaranayake DB, Quienters VG, Wickramasinghe VP. Effects of vitamin E supplementation on the clinical outcome of dengue fever and dengue haemorrhagic fever in children. Asian Pac J Trop Dis. 2017;7:645-9.
- 73. Gupta E, Dar L, Kapoor G, Broor S. The changing epidemiology of dengue in Delhi, India. Virology journal. 2006 Dec;3:1-5.
- 74. Wiwanitkit V. Dengue fever: diagnosis and treatment. Expert review of anti-infective therapy. 2010 Jul 1;8(7):841-5.
- 75. Apte-Sengupta S, Sirohi D, Kuhn RJ. Coupling of replication and assembly in flaviviruses. Current opinion in virology. 2014 Dec 1;9:134-42.
- 76. Zou G, Chen YL, Dong H, Lim CC, Yap LJ, Yau YH, Shochat SG, Lescar J, Shi PY. Functional analysis of two cavities in flavivirus NS5 polymerase. Journal of Biological Chemistry. 2011 Apr 22;286(16):14362- 72.
- 77. Maltsev OV, Kasyanenko KV, Zhdanov KV, Malyshev NA, Kolomoets EV, Konomou VK. The experience in treatment of dengue fever using antiviral drug riamilovir in the Republic of Guinea (case report). Terapevticheskii arkhiv. 2023 Feb 24;95(1):85-9.
- 78. Pierson TC, Diamond MS. Vaccine development as a means to control dengue virus pathogenesis: do we know enough?. Annual Review of Virology. 2014 Sep 29;1:375-98.
- 79. Dimaano EM, Saito M, Honda S, Miranda EA, Alonzo MT, Valerio MD, Mapua CA, Inoue S, Kumaori A, Matias R, Natividad FF. Lack of efficacy of high-dose intravenous immunoglobulin treatment of severe thrombocytopenia in patients with secondary dengue virus infection.
- 80. Deng SQ, Yang X, Wei Y, Chen JT, Wang XJ, Peng HJ. A review on dengue vaccine development. Vol. 8, Vaccines 2020.
- 81. Guy B, Noriega F, Ochiai RL, L'azou M, Delore V, Skipetrova A, et al. A recombinant live attenuated tetravalent vaccine for the prevention of dengue. Expert Rev Vaccines. 2017;16(7). DOI: 10.1080/14760584.2017.1335201
- 82. Pang EL, Loh HS. Towards development of a universal dengue vaccine – How close are we? Vol. 10, Asian Pacific Journal of Tropical Medicine 2017.
- 83. Shukla R, Rajpoot RK, Arora U, Poddar A, Swaminathan S, Khanna N. Pichia pastoris-expressed bivalent virus-like particulate vaccine induces domain III-focused bivalent neutralizing antibodies without antibody-dependent enhancement in vivo. Front Microbiol. 2018;8(JAN). DOI: 10.3389/fmicb.2017.02644
- 84. Whitehead SS, Blaney JE, Durbin AP, Murphy BR. Prospects for a dengue virus vaccine. Vol. 5, Nature Reviews Microbiology 2007.
- 85. Kochel T, Wu SJ, Raviprakash K, Hobart P, Hoffman S, Porter K, et al. Inoculation of plasmids expressing the dengue-2 envelope gene elicit neutralizing antibodies in mice. Vaccine. 1997;15(5). DOI: 10.1016/S0264-410X(97)00215-6
- 86. Capeding MR, Tran NH, Hadinegoro SRS, Ismail HIHM, Chotpitayasunondh T, Chua MN, et al. Clinical efficacy and safety of a novel tetravalent dengue vaccine in healthy children in Asia: A phase 3, randomised, observer-masked, placebo-controlled trial. The Lancet. 2014;384(9951). DOI: 10.1016/S0140-6736(14)61060-6
- 87. Men R, Bray M, Clark D, Chanock RM, Lai CJ. Dengue type 4 virus mutants containing deletions in the 3'noncoding region of the RNA genome: analysis

of growth restriction in cell culture and altered viremia pattern and immunogenicity in rhesus monkeys. Journal of virology. 1996 Jun;70(6):3930-7.

- 88. Whitehead SS, Falgout B, Hanley KA, Blaney Jr JE, Markoff L, Murphy BR. A live, attenuated dengue virus type 1 vaccine candidate with a 30-nucleotide deletion in the 3′ untranslated region is highly attenuated and immunogenic in monkeys. Journal of virology. 2003 Jan 15;77(2):1653-7.
- 89. Blaney Jr JE, Sathe NS, Goddard L, Hanson CT, Romero TA, Hanley KA, Murphy BR, Whitehead SS. Dengue virus type 3 vaccine candidates generated by introduction of deletions in the 3′ untranslated region (3′-UTR) or by exchange of the DENV-3 3′-UTR with that of DENV-4. Vaccine. 2008 Feb 6;26(6):817-28.
- 90. Blaney Jr JE, Hanson CT, Firestone CY, Hanley KA, Murphy BR, Whitehead SS. Genetically modified, live attenuated dengue virus type 3 vaccine candidates. American Journal of Tropical Medicine and Hygiene. 2004 Dec 1;71(6):811-21.
- 91. Kinney RM, Butrapet S, Chang GJ, Tsuchiya KR, Roehrig JT, Bhamarapravati N, Gubler DJ. Construction of infectious cDNA clones for dengue 2 virus: strain 16681 and its attenuated vaccine derivative, strain PDK-53. Virology. 1997 Apr 14;230(2):300-8.
- 92. Torres-Flores JM, Reyes-Sandoval A, Salazar MI. Dengue vaccines: an update. BioDrugs. 2022 May;36(3):325-36.

*This article is an Open Access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited and it is not used for commercial purposes; 2024, Nasir M. et al., Applied Systems and Discoveries Journals.*