

REVIEW Article

The role of CD36 in renal and bladder cancer

Mihai Ioan Pavalean^{1,*}, Victor Lucian Madan^{1,2}, Mihaela Cristina Pavalean¹, Laura Cristina Ceafalan^{1,3}, Mihail Eugen Hinescu^{1,3}

¹ Faculty of Medicine, Carol Davila University of Medicine and Pharmacy, 050474 Bucharest, Romania

² Emergency University Central Military Hospital, 010825 Bucharest, Romania

³ Victor Babeş National Institute of Pathology, 050096 Bucharest, Romania

* *Corresponding authors: Mihai Ioan Pavalean*, Faculty of Medicine, Carol Davila University of Medicine and Pharmacy, 050474 Bucharest, Romania; email: mihai-ioan.pavalean@drd.umfcd.ro (M.I.P.), ORCID: 0000-0003-0245-0905

Submitted: Mar. 25, 2026; Revised: Mar. 31, 2026; Accepted: Mar. 31, 2026; Published: Mar. 31, 2026

Citation: Pavalean MI, Madan VL, Pavalean MC, Ceafalan LC, Hinescu ME. The role of CD36 in renal and bladder cancer. Discoveries 2026, 14(1): e225. DOI: 10.15190/d.2026.4

ABSTRACT

CD36 functions as both a lipid transporter and scavenger receptor, integrating metabolic and inflammatory signaling pathways. It plays a critical role in maintaining cellular homeostasis and influencing disease progression. This review summarizes the structure, ligands, functions, regulation, and clinical implications of CD36 in renal and bladder cancer. Increased CD36 expression promotes enhanced fatty acid uptake, which supports tumor cell proliferation, migration and survival, by mediating metabolic reprogramming and interacting with the tumor microenvironment. In renal cancer, most frequently clear cell renal carcinoma (ccRCC), which has a typical metabolic phenotype, CD36 is involved in lipid accumulation and oxidative stress pathways. Pathogenic mechanisms include hypoxia-inducible factor (HIF)-driven pathways and carnitine palmitoyl transferase 1A (CPT1A) via the PPAR α /CD36 axis, which phosphorylate Akt. By using fatty acid oxidation, CD36 lead to the production of reactive oxygen species and to transcription of genes mediating a pro-tumor function, inducing tumor-associated macrophages (TAM). In bladder cancer, CD36 is implicated in tumoral cells proliferation, survival, and adaptation to

metabolic stress, epithelial–mesenchymal transition (EMT) and influences the tumor microenvironment, through interactions with tumor-associated macrophages and inflammatory signaling pathways. Although multiple studies propose CD36 as a prognostic biomarker, inconsistencies across cohorts limit its clinical translation. Notably, advances have revealed the regulatory networks governing distinct physiological properties of CD36, thereby identifying targeting CD36 as a potential strategy for cancer treatment. Inhibition of CD36-mediated lipid metabolism and signaling pathways may reduce tumor growth and metastatic potential. However, further research is necessary to clarify its context-dependent functions and to develop effective CD36-targeted therapies. To our knowledge, this is the first review to systematically examine the role of CD36 across both renal and bladder cancer. It could be the first step toward identifying new mechanisms mediated by CD36 in these malignancies.

Keywords

CD36, renal cell cancer, bladder cancer, lipid metabolism, metastasis, prognostic biomarker.

Abbreviations

Long-chain acyl CoA synthetase 1 (ACSL1); advanced glycation end products (AGE); advanced oxidation protein products (AOPPs); bladder cancer (BC); clear cell renal carcinoma (ccRCC); cluster of differentiation 36 (CD36); circular RNAs (circRNAs); Carcinoma in situ (CIS); cytotoxic necrotizing factor 1 (CNF1); carnitine palmitoyl transferase (CPT1A); chemokine receptors 3 (CXCR3); dendritic cells (DC); diabetic nephropathy (DN); epithelial-to-mesenchymal transition (EMT); endoplasmic reticulum (ER); endostatin (ES); fatty acids (FA); FA uptake and consumption (FAO); hypoxia-inducible factor (HIF); interleukin (IL); ischemia-reperfusion injury (IRI); long non-coding RNA (lncRNA); liver X receptor β (LXR β); micropapillary variant of bladder cancer (MPBC); matrix metalloprotease (MMP); messenger RNA (mRNA); mitochondrial reactive oxygen species (mtROS); microvascular endothelial cells (MVECs); NLR family pyrin domain 3 (NLRP3); oxidized high-density lipoprotein (ox-HDL); oxidized low-density lipoprotein (ox-LDL); peroxisome proliferator-activated receptor α (PPAR α); renal cell carcinoma (RCC); reactive oxygen species (ROS); renal transplantation (RTx); subcutaneous adipose tissue (SAT); suppressor of mothers against decapentaplegic (SMAD); thrombospondin (TSP); thrombospondin type I repeats (TSR); time to progression (TTP); visceral adipose tissue (VAT); vascular endothelial growth factor (VEGF).

SUMMARY

1. An overview of the CD36 receptor
2. The tissue distribution of the CD36 receptor in the urinary system and its functions
 - 2.1. CD36 and the renal tissue
 - 2.2. CD36 and the urinary bladder tissue
3. The UroOnco spectrum of CD36
 - 3.1. Renal cell carcinoma
 - 3.2. Urothelial (transitional cell) carcinoma
4. Potential prognostic markers for ccRCC
 - 4.1. The role of CD36 in tumor progression and dissemination
 - 4.2. The prognostic value of CD36
5. Conclusion

1. An overview of the CD36 receptor

CD36, also known as SR-B2, was first recognised as a receptor for thrombospondin (TSP). It is considered the archetypal class B scavenger receptor¹. Scavenger receptors are currently categorized into ten classes (A–J) based on their sequence similarity or shared structural features. There is little to no sequence

homology between different classes of scavenger receptors^{2,3}. Mammalian class C scavenger receptors are currently not known, and the class C scavenger receptors have only been described in *Drosophila melanogaster*².

From a structural standpoint, CD36 has an apparent molecular mass of 88 kDa due to extensive glycosylation⁴. It is a 472 amino acid protein and comprises of five distinct regions: the carboxy-terminal intracellular domain (COOH-terminal), the amino-terminal intracellular domain (NH₂-terminal), an extracellular domain, and two transmembrane domains⁵. It contains two transmembrane domains and several palmitoylation sites that the C and N termini of CD36 are intracellular, and both ends are palmitoylated (3/7, 464/466). In addition to palmitoylation, the COOH terminus contains a set of ubiquitination sites (469/472)^{6,7} and has a motif of CXCX5K located on the cytosolic ends of the T cell co-receptors CD4 and CD8, which may be involved in the binding of src-related protein tyrosine kinases⁸.

The extracellular domains, recognized by the ligand, is a highly glycosylated hydrophobic ring containing ten glycosylation sites (79/102/134/163/205/220/235/247/321/417), two phosphorylation sites (92, 237), and three pairs of disulfide bonds (243-311/313-322/272-333)^{6,9-11}. CD36 has the ability to bind numerous ligands, including advanced glycation end products (AGE) - a family of compounds that are the products of nonenzymatic reactions between reducing sugars and proteins, lipids, or nucleic acids, oxidized low density lipoprotein (oxLDL), fatty acids (FAs), collagen, TSP, and anionic phospholipids¹²⁻¹⁶. In the extracellular region of rat CD36, there are ten possible glycosylation sites, eight of which are conserved across humans and rats¹⁷ (**Figure 1**).

The previously described structure is essential for the recognition and endocytic uptake of oxidized phospholipids and modified low-density lipoprotein (LDL). Additionally, CD36 activity extends to the clearance of apoptotic cells, binding of amyloid proteins¹⁸, and involvement in inflammatory processes in atherosclerosis and Alzheimer's disease¹⁹⁻²². CD36 also acts as a potent endogenous inhibitor of angiogenesis, including tumor neovascularization²³. In supporting this role, Hale et al. reported that deletion of CD36 in mice altered tumor growth and angiogenesis in the presence of thrombospondin type I repeat (TSR) proteins and demonstrated that Histidine-Rich Glycoprotein modulates this activity²³.

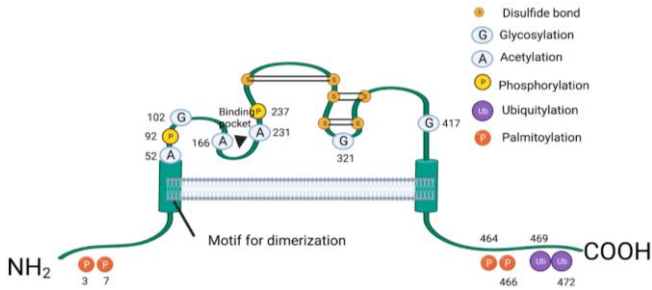


Figure 1. The structure of CD36 (Created in BioRender. Ceafalan, L. (2026) <https://BioRender.com/zqoonwp>)

2. The tissue distribution of the CD36 receptor in the urinary system and its functions

CD36 is expressed in microvascular endothelial cells (MVECs)²⁰, stromal cells²⁴, and immune cells²⁵, and its level varies across these cell types. In malignant epidermal tumor cells, such as in the cells of ovarian cancer²⁶, gastric cancer²⁷, glioblastoma (GBM)²⁸, and oral carcinoma^{29,30}, CD36 expression is upregulated. In tumor microvessels, which support tumor development and metastasis, CD36 expression is generally downregulated³¹.

CD36 demonstrates a distinct and cell-type-specific distribution within the urinary system, indicating its involvement in renal lipid handling, tubular reabsorption, and local immune responses. In the kidney, CD36 is primarily expressed in proximal tubular epithelial cells, podocytes, mesangial cells, and endothelial cells, where it facilitates fatty acid uptake and lipid accumulation^{6,32}. This distribution is functionally significant, as dysregulated CD36 expression has been linked to lipotoxicity, oxidative stress, and inflammation, which are key mechanisms in the development of renal disorders such as diabetic nephropathy and chronic kidney disease^{6,31}.

Beyond the kidney, recent studies indicate that CD36 is expressed in additional components of the urinary tract, including the urothelium, where it may contribute to epithelial integrity and the regulation of inflammatory responses²⁹. Alterations in CD36 expression and function have also been associated with tumorigenesis in urological malignancies, such as renal cell carcinoma and bladder cancer, highlighting its context-dependent biological significance^{33,34}.

Due to its diverse functional roles and heterogeneous tissue distribution, characterizing the spatial expression patterns of CD36 within the urinary

system is essential for clarifying its contributions to both normal physiology and disease.

2.1. CD36 and the renal tissue

CD36 serves as the primary uptake system for free fatty acids in the kidney and is highly expressed in epithelial cells of the proximal and distal tubules, as well as in podocytes and mesangial cells (**Table 1**)^{35,36}. Numerous functions of CD36 in renal tissue, healthy tissue distribution, and lipid metabolism have been described.

Table 1. CD36 ligands for every renal cell type and their effects

Cell type	CD36 ligand	Roles	
Proximal tubule epithelial cells	Fatty acids	ATP production, lipid accumulatio, apoptosis	32,37
	Albumin	Fibrosis	35
	AGEs	Apoptosis	32
	AOPPs	Inflammation, apoptosis	37
	Ox-LDL	Inflammation, apoptosis, ROS production	38–41
Monocytes and/or macrophages	Ox-LDL	Foam cell formation, ROS production, lipid accumulatio, apoptosis	42
Podocytes	Fatty acids	Lipid accumulatio, apoptosis, ROS production	43–45
Mesangial cells	Ox-HDL	Inflammation, apoptosis	38
Vascular endothelial cells	Fatty acids	ATP production	46
	Ox-LDL	Foam cell formation, lipid accumulatio, inflammation	42

CD36 functions as a critical upstream regulator of various inflammatory pathways in renal tubular epithelial cells, like NLR family pyrin domain-containing 3 (NLRP3) inflammasome activation by promoting mitochondrial reactive oxygen species (mtROS) generation. The NLRP3 inflammasome, a principal mediator of innate immune signalling, is increasingly recognized as a central contributor to

inflammatory injury in diabetic nephropathy (DN)⁴³. To demonstrate the mechanistic role of CD36 in diabetic nephropathy (DN), Hou et al. employed both *ex vivo* and *in vivo* (murine) models. They found that exposure to high glucose conditions induces significant metabolic reprogramming in renal tubular cells, characterized by a shift from oxidative phosphorylation (OXPHOS) to glycolysis, which promotes mtROS accumulation. Notably, CD36 exacerbates this process by inhibiting mitochondrial fatty acid oxidation (FAO), thereby reinforcing a feed-forward cycle of mitochondrial dysfunction and oxidative stress⁴⁴.

Taken together, these findings establish CD36 as a central nexus of metabolic and inflammatory pathways connecting high glucose-induced metabolic stress to NLRP3 inflammasome activation. Consequently, targeting the CD36-mtROS-NLRP3 axis may represent a promising therapeutic strategy to mitigate inflammation-driven progression in DN.

Additionally, studies have demonstrated that increased dietary fat consumption, in the absence of circulating proprotein convertase subtilisin/kexin type 9, promotes renal lipid accumulation and subsequent renal injury⁴⁷.

2.2 CD36 and the urinary bladder tissue

Although CD36 expression in bladder tissue is less extensively characterized than in renal structures, current evidence indicates that CD36 contributes to local lipid metabolism and innate immune surveillance. CD36 mediates the uptake of long-chain fatty acids, as a fatty acid translocase, into urothelial and stromal cells, thereby supporting membrane remodeling and cellular energy demands essential for maintaining urothelial barrier integrity and turnover^{48,49}.

Beyond its metabolic functions, CD36 also acts as a pattern recognition receptor that participates in host defense mechanisms within the bladder. It binds a diverse array of endogenous and exogenous ligands, including oxidized lipids and microbial components, thereby facilitating the detection of pathogens and damaged cells. CD36 activates intracellular signaling pathways, such as NF- κ B and MAPK, in cooperation with Toll-like receptors, which induce the production of pro-inflammatory cytokines and chemokines to coordinate the local immune response⁵⁰.

Experimental evidence from urinary tract infection models indicates that CD36 plays a

protective role in bladder immunity by promoting the clearance of bacteria and apoptotic neutrophils. Dysregulation of CD36 expression, including its downregulation by bacterial toxins, can impair pathogen clearance and intensify inflammation, leading to increased bacterial burden and tissue injury in the bladder⁴⁶.

Collectively, these findings demonstrate that CD36 in the urinary bladder integrates metabolic and immune functions, contributing to epithelial homeostasis and the regulation of inflammatory responses during infection. This dual role emphasizes its significance as a key modulator of bladder physiology, especially under conditions of immune activation and tissue stress.

3. The UroOnco spectrum of CD36

Urological cancers, including renal and bladder malignancies, constitute a biologically heterogeneous group of tumors defined by intricate interactions among metabolic reprogramming, immune modulation, and microenvironmental adaptation. Renal cell carcinoma (RCC), originating in the renal cortex, accounts for eighty to eighty-five percent of primary renal cancers. Transitional cell carcinoma of the renal pelvis is the second most common, comprising approximately eight percent of cases. Other parenchymal epithelial tumors, such as oncocytomas, collecting duct tumors, and kidney sarcomas, are rare⁵¹⁻⁵³. In the urinary tract, bladder cancer (BC) is the most prevalent and is increasingly recognized as a malignancy with significant metabolic reprogramming. Enhanced lipid uptake, storage, and β -oxidation, regulated in part by CD36 and PPAR signalling, contribute to tumor growth, survival, and therapeutic resistance²⁹.

Recent studies have demonstrated an association between CD36 and tumor progression and metastasis in both RCC and BC. Inflammation and dysregulated lipid metabolism, mediated by CD36, are central to the pathogenesis of these cancers. Alterations in CD36 expression and function via signalling pathways have been implicated in cancer development, progression, and dissemination^{32,33}.

3.1 Renal cell carcinoma

RCC has been classified into distinct subtypes:

1. Clear cell (seventy-five to eighty-five percent of tumors)-ccRCC,

2. Papillary (ten to fifteen percent),
3. Chromophobe (five to ten percent),
4. Oncocytic (three to seven percent),
5. Collecting duct (very rare),
6. Molecularly defined renal cell carcinomas (rare)⁵⁴⁻⁵⁶.

The most frequent histological subtype of renal cancer, clear cell renal cell carcinoma (ccRCC), is characterized by the accumulation of intracellular lipids and glycogen. This distinctive metabolic phenotype reflects alterations in lipid metabolism, which are now recognized as a hallmark of ccRCC biology.

To better understand the processes underlying CD36-mediated ccRCC development, several mechanisms have been described in studies. One new theoretical explanation for the pathogenesis of ccRCC is the regulatory role of carnitine palmitoyl transferase 1A (CPT1A) via the PPAR α /CD36 axis. CPT1A is located on the outer mitochondrial membrane and is a rate-limiting fatty acid oxidation (FAO) enzyme that transports fatty acids into mitochondria for oxidation by converting acyl-CoA into acyl-carnitines. CPT1A expression is crucial for lipid accumulation, which promotes ccRCC development. Therefore, CPT1A prevents cholesterol uptake and lipid accumulation by increasing peroxisome proliferator-activated receptor α (PPAR α) levels through regulation of CD36, with further control of Akt phosphorylation implicated in ccRCC growth. Also, Akt activation, which regulates cell survival under stress conditions in RCC, is linked to cancer survival and proliferation⁵⁷.

The accumulation and storage of lipids are critical for the management of oxidative and endoplasmic reticulum (ER) stress, and lipids promote metastasis in ccRCC tumors⁵⁸⁻⁶⁰.

Another explanation for the pathogenesis of ccRCC is the activation of the hypoxia-inducible factor (HIF) signaling pathways, which remains poorly understood. Hypoxic conditions further enhance CD36-mediated lipid remodeling. HIF1 α signaling increases the expression of lipid receptors, such as CD36 and ACVRL1, as well as genes involved in lipid transport (FABP7) and storage (PLIN2, HILPDA), thereby reinforcing a lipid-rich tumor phenotype⁶¹.

Additionally, Liao et al. demonstrated that hypoxia directly induces CD36 expression through HIF-2 α activation. Functional studies show that

CD36 knockdown reduces lipid accumulation and eliminates HIF-2 α -driven tumor-promoting effects, establishing CD36 as a key mediator of hypoxia-induced metabolic reprogramming^{62,63}.

Moreover, since CD36 is expressed not only in renal tumor cells but also in macrophages, some studies have focused on the tumor microenvironment (TME), suggesting potential links between ccRCC lipid metabolism and its immune infiltrate, particularly tumor-associated macrophages (TAM). Therefore, CD36 plays an important role in fatty acid transport, macrophage lipid accumulation, and immune modulation⁶⁴. In a murine model, Su et al. reported that CD36 plays a key role in TAMs for lipid accumulation and tumor development. It showed that TAMs express CD36 and use fatty acid oxidation, leading to production of reactive oxygen species and to transcription of genes mediating a pro-tumor function (M2-like phenotype)⁶⁵.

This CD36-mediated influence on macrophage polarization correlates with immunosuppression, thereby reducing the effectiveness of anti-tumor immunity in the kidney tumour microenvironment⁶⁴. In summary, these findings establish CD36 as a critical convergence point that integrates hypoxia signalling and lipid metabolism in ccRCC. By interacting with HIF-driven pathways and PI3K/Akt signalling, CD36 maintains a lipid-dependent oncogenic state, highlighting its potential as a therapeutic target in metabolically reprogrammed renal tumors.

3.2 Urothelial (transitional cell) carcinoma

Bladder cancer is the most prevalent neoplasm of the urinary tract and is classified into two types with distinct molecular characteristics. In 75% of cases, the disease is confined to the mucosa and is termed non-muscle-invasive bladder cancer (NMIBC). The remaining cases are classified as muscle-invasive bladder cancer (MIBC)⁶⁶.

CD36-mediated fatty acid transport improves the availability of exogenous lipids, which tumor cells use to support proliferation, survival, and adaptation to metabolic stress. This lipid dependency is especially significant in bladder cancer, where changes in lipid metabolism are increasingly recognized as a hallmark of tumor progression⁶⁷.

CD36 also influences the tumor microenvironment and contributes to aggressive disease behaviour. Clinical studies indicate that CD36

expression is associated with advanced pathological stages in MIBC, supporting its role in tumor progression and dissemination⁶⁸. NMIBC tumor samples exhibit increased expression of the fatty acid transporters FATP4, CD36, and ACSL1⁶⁹.

Regarding molecular changes in BC after radiotherapy, Shang et al. sought to identify the mechanisms and signal transduction pathways. They investigated the profile of messenger RNA (mRNA) and long non-coding RNA (lncRNA). The expression of matrix metalloproteinase MMP-3, MMP-10, MMP-12, and MMP-13 was significantly increased in BC after RTx, whereas the expression of CD36 was decreased⁷⁰.

We also perform a study that examines a panel of 26 dysregulated microRNAs (miRNAs) in BC, several of which may be associated with tumor aggressiveness and increased risk of disease progression⁷¹.

Collectively, current evidence demonstrates that CD36 plays multiple roles in bladder cancer by promoting lipid-driven metabolic reprogramming, sustaining cancer stemness, and facilitating tumor progression.

4. Potential prognostic markers for ccRCC

Identifying molecular markers linked to poor prognosis in renal cell carcinoma is crucial for elucidating tumor heterogeneity and informing personalized therapeutic strategies (Table 2).

Table 2. Markers that are potentially associated with a worse prognosis for ccRCC

Markers	
Human B7 homolog 1 (B7H1) and 4 (B7H4) expression	72
Low levels of carbonic anhydrase IX (CAIX)	73
High levels of the proliferation marker Ki-67	73
Higher levels of hypoxia-inducible factor (HIF)-1 alpha expression	74
Expression of the U3 small nucleolar ribonucleoprotein (IMP3)	69,75,76
Deletion of chromosome 9p	77
Mutations of tumor suppressor genes on chromosome 3p21, including mutations of breast cancer type 1 (BRCA1)-associated protein 1 (BAP1) and SET domain containing 2 (SETD2)	78
PD-L1/PD-1	79

We found evidence for a continuous crosstalk between these markers and CD36 in other malignant

pathologies. Co-expression of PD-L1/PD-1 with CXCR3/ CD36 in circulating lymphocytes and serum IL-19 levels explain poor prognosis and can be considered potential markers for extranodal involvement in lymphoma. Co-expression was evaluated by flow cytometry in seventy-eight lymphoma patients before and after therapy. There also was 50 healthy controls in the study⁷⁹.

The relationship between Ki67 and CD36 was demonstrated in oral squamous cell carcinoma. CD36-positive cells demonstrated increased expression of Ki-67 and migration activity compared with CD36-negative cells⁸⁰.

Hypoxia and hypoxia-inducible factors (HIFs) play essential and multiple roles in renal ischemia-reperfusion injury (IRI). Qu et al. demonstrated a new role for the HIF-2 α / CD36 regulatory axis in rewiring dendritic cell (DC) lipid metabolism under IRI-associated hypoxia. This is a potential therapeutic target to resolve long-standing obstacles in the treatment of this severe complication⁸¹.

4.1 The role of CD36 in tumor progression and dissemination

In bladder cancer, elevated CD36 expression is linked to increased tumor growth, invasiveness, and the development of aggressive phenotypes. CD36 has been known to be associated with altered lipid metabolism and initiation of metastasis, contributing to cancer progression^{29,67,75}. High expression of FATP4, CD36, and long-chain acyl-CoA synthetase 1 (ACSL1) has been associated with metastasis. Patients with NMIBC who demonstrate high expression of these fatty acid transporters need to be closely monitored and treated more aggressively⁶⁹. Additionally, CD36 is implicated in the maintenance of cancer stem cell populations, which are essential for tumor initiation, therapy resistance, and metastatic spread⁸².

Overexpression of CD36 in cell lines or patient-derived cells with low metastatic potential greatly increased their potential to metastasize to lymph nodes, with penetrance increasing from less than 20% to 75-80%²⁹. The link between CD36 and the initiation of the metastatic process was also demonstrated, without being able to specify exactly how this occurs, because CD36+ cells are not only capable of initiating metastasis but can also recapitulate their molecular and cellular heterogeneity from the primary origin²⁹. Clinical

evidence further supports this role, as elevated CD36 expression correlates with advanced tumor stage and poorer prognosis in bladder cancer patients⁶⁸.

Current evidence regarding the role of CD36 investigates the molecular mechanism and signal transduction pathways implicated in BC after Rtx. They analysed the mRNA and lncRNA profile. One of the study’s conclusions was that several genes were down-regulated in BC after Rtx, including CD36⁷⁰. Additionally, endostatin gene therapy in RCC can also have a role in treatment, reducing the number of lung tumor nodules, reducing metastasis and resulting in a higher survival rate^{76,83}.

Collectively, these findings support a role for CD36 in the regulation of metastatic processes, both through tumor-intrinsic mechanisms and via modulation of the tumor microenvironment.

4.2. The prognostic value of CD36

In several types of cancers, there is a positive correlation between CD36 expression in tumor cells and poor clinical outcome, as illustrated in **Table 3**.

The CD36 receptor has gained increasing attention as a prognostic biomarker in urological malignancies, including both bladder cancer and renal cell carcinoma, due to its involvement in lipid metabolism, tumor progression and immune regulation.

Although direct clinical data remain limited in RCC, emerging evidence indicates that CD36 expression correlates with increased tumor aggressiveness and unfavourable clinical outcomes. We identified 2 studies with CD36 expression that support the above, one of which demonstrated a correlation between mRNA responsible for the CD36 cluster and visceral adipose tissue (VAT), based on the premise that a higher VAT rate leads to a more aggressive tumor progression or a worse prognosis for patients with RCC⁹¹ and the other suggest that the transport of FA through the cell membrane, facilitated by CD36 and FATP4, could be pathognomonic for RCC. Also, it demonstrated a strong correlation between oncogenesis and tumor progression and the simultaneous overexpression of both FATP4 and CD36⁹².

In addition to protein-level alterations, transcriptomic analyses have identified regulatory RNA networks associated with RCC prognosis. Seven mRNA targets of miRNA-21 and 12 mRNA targets of miRNA-155 were identified as key

prognostic factors for patients with RCC. These two miRNAs were significantly related to 5-year survival ($p < 0.05$). The analysis of the competitive endogenous RNA (ceRNA) regulatory network provided new and useful perspectives for the development of further therapeutic strategies for RCC⁹³.

Table 3. The role of CD36 in various types of cancers

Type of cancer	Role of CD36	Ref.
Oral squamous cell carcinoma	CD36 depletion greatly reduced the size of lymph node metastases in all tumor lines	29
Human melanoma	Impairs metastasis – inhibition of CD36	29
Breast cancer	Impairs metastasis – inhibition of CD36	29,84
Cervical cancer	Poor tumor differentiation, lymph node metastasis	85
Gastric cancer	Contribute to the tumor proliferation	27,86
Bladder cancer	Promots bladder cancer	45
Prostate cancer	Associated with reduced relapse-free survival and increased incidence in metastases	87
Cholangiocarcinoma	Cancer relapse	88
Hepatocellular carcinoma	CD36 knockdown - impairs cell proliferation, decreases migration and invasion	82
Colorectal cancer	Associated with metastatic tumors, reduced 5-year survival	89
Leukemias and lymphomas	CD36 knockdown - decreases FA uptake and increases chemotherapeutic-induced apoptosis	90
Glioblastoma	Elevated capacity to form tumorspheres in culture	82

Clinical studies in bladder cancer have shown that CD36 expression correlates with increased tumor aggressiveness and poorer clinical outcomes. For instance, immunohistochemical analyses in muscle-invasive bladder cancer revealed that CD36-positive tumors correlate with advanced pathological stage

- Elevated CD36 expression, in tissue or circulation, is an indicator of disease progression and metastatic risk, correlating with increased invasiveness, metastatic potential, and poor prognosis, particularly in clear cell renal cell carcinoma and bladder cancer.
- Targeting CD36-mediated lipid metabolism can reduce tumor growth and metastasis in preclinical models, being promising for the treatment of renal and bladder cancer.

and a trend toward lymph node involvement. Importantly, patients with CD36-positive tumors exhibited significantly shorter disease-free survival compared to those with CD36-negative tumors, supporting its role as a negative prognostic marker⁶⁸. Furthermore, alterations in lipid transport pathways involving CD36 are associated with tumor progression and increased metastatic potential in bladder cancer. Elevated expression of CD36 and other fatty acid transporters enhances lipid uptake, thereby promoting tumor growth, invasion, and resistance to therapy. These findings further underscore the prognostic significance of CD36⁶⁹. It was also demonstrated that the fatty acid uptake by tumor cells increased, and the fatty acid was depleted within the tumor microenvironment through the interaction of circZNF609 with IGF2BP2 and CD36⁹⁴.

Additionally, there is a connection between time to progression (TTP) and individual gene expression, adjusting for clinical covariates, in a specific type of bladder cancer. The study refers to a high-risk non-muscle-invasive (HG1) micropapillary variant of bladder cancer (MPBC). Furthermore, high expression of FABP3 and CD36 was associated with shorter TTP⁹⁵.

Collectively, these findings indicate that CD36 represents a promising prognostic biomarker in both BC and RCC, reflecting its central role in lipid-driven tumor progression and immune regulation. Its expression may help stratify patients by risk and could serve as a potential therapeutic target in the future.

5. Conclusion

CD36 has recently been identified as a promising therapeutic target across multiple cancer types, offering significant clinical implications. In kidney and bladder cancer, high levels of CD36 could be considered a marker for poor prognosis. Even if the mechanisms underlying CD36's role in oncogenesis

remain poorly understood, CD36 inhibitors could be considered for cancer therapy in the near future. Targeting CD36 may offer therapeutic benefit, but given its systemic metabolic roles, strategies such as tumor-specific delivery or pathway-selective inhibition will be essential.

Further clinical trials are needed to investigate the regulation and downstream signalling pathways of CD36 during the manipulation of the target in precise clinical translation.

Although evidence increasingly supports the role of CD36, significant gaps persist in understanding the molecular pathways that regulate its activity and its interactions with stromal and immune components. Recognising CD36 at the intersection of metabolic regulation and tumor ecology underscores its potential as both a biomarker and a therapeutic target.

Conflict of Interest

The authors have no conflicts of interest to disclose.

Acknowledgements

Ethical approval is not required for this type of study. This study was funded by Program 1 - Development of the national RD system, Subprogram 1.2 - Institutional Performance - RDI excellence funding projects.; Contract PFE 33/2021, PN: 23.16.01.02.

Author Contributions

Conceptualization, M.I.P, V.L.M and M.C.P; writing - original draft preparation, M.I.P, V.L.M and M.C.P writing - review and editing, M.I.P, V.L.M, M.C.P, L.C.C and M.E.H; figure preparation, M.I.P and M.C.P; supervision, M.E.H and L.C.C. All authors have read and agreed to the published version of the manuscript.

Generative AI statement

The author(s) declared that generative AI was used in the creation of this manuscript, solely for the purpose of grammar and language correction. No data or text were generated using AI.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Asch AS, Barnwell J, Silverstein RL, Nachman RL. Isolation of the thrombospondin membrane receptor. *J Clin Invest.* 1987;79(4):1054-1061. doi:10.1172/JCI112918
2. Krieger M. The other side of scavenger receptors: pattern recognition for host defense: *Curr Opin Lipidol.* 1997;8(5):275-280. doi:10.1097/00041433-199710000-00006
3. Whelan FJ, Meehan CJ, Golding GB, McConkey BJ, E Bowdish DM. The evolution of the class A scavenger receptors. *BMC Evol Biol.* 2012;12(1):227. doi:10.1186/1471-2148-12-227
4. Lehner R, Quiroga AD. Fatty Acid Handling in Mammalian Cells. In: *Biochemistry of Lipids, Lipoproteins and Membranes.* Elsevier; 2016:149-184. doi:10.1016/B978-0-444-63438-2.00005-5
5. Yang R, Liu Q, Zhang M. The Past and Present Lives of the Intraocular Transmembrane Protein CD36. *Cells.* 2022;12(1):171. doi:10.3390/cells12010171
6. Yang X, Okamura DM, Lu X, et al. CD36 in chronic kidney disease: novel insights and therapeutic opportunities. *Nat Rev Nephrol.* 2017;13(12):769-781. doi:10.1038/nrneph.2017.126
7. Cao D, Luo J, Chen D, et al. CD36 regulates lipopolysaccharide-induced signaling pathways and mediates the internalization of *Escherichia coli* in cooperation with TLR4 in goat mammary gland epithelial cells. *Sci Rep.* 2016;6(1):23132. doi:10.1038/srep23132
8. Xing Q, Feng Y, Sun H, et al. Scavenger receptor MARCO contributes to macrophage phagocytosis and clearance of tumor cells. *Exp Cell Res.* 2021;408(2):112862. doi:10.1016/j.yexcr.2021.112862
9. Thorne RF, Ralston KJ, De Bock CE, et al. Palmitoylation of CD36/FAT regulates the rate of its post-transcriptional processing in the endoplasmic reticulum. *Biochim Biophys Acta BBA - Mol Cell Res.* 2010;1803(11):1298-1307. doi:10.1016/j.bbamcr.2010.07.002
10. Hao JW, Wang J, Guo H, et al. CD36 facilitates fatty acid uptake by dynamic palmitoylation-regulated endocytosis. *Nat Commun.* 2020;11(1):4765. doi:10.1038/s41467-020-18565-8
11. Zeng S, Wu F, Chen M, et al. Inhibition of Fatty Acid Translocase (FAT/CD36) Palmitoylation Enhances Hepatic Fatty Acid β -Oxidation by Increasing Its Localization to Mitochondria and Interaction with Long-Chain Acyl-CoA Synthetase 1. *Antioxid Redox Signal.* 2022;36(16-18):1081-1100. doi:10.1089/ars.2021.0157
12. Fujimoto E, Kobayashi T, Fujimoto N, Akiyama M, Tajima S, Nagai R. AGE-Modified Collagens I and III Induce Keratinocyte Terminal Differentiation through AGE Receptor CD36: Epidermal-Dermal Interaction in Acquired Perforating Dermatitis. *J Invest Dermatol.* 2010;130(2):405-414. doi:10.1038/jid.2009.269
13. Ohgami N, Nagai R, Ikemoto M, et al. CD36, a Member of the Class B Scavenger Receptor Family, as a Receptor for Advanced Glycation End Products. *J Biol Chem.* 2001;276(5):3195-3202. doi:10.1074/jbc.M006545200
14. Febbraio M, Hajjar DP, Silverstein RL. CD36: a class B scavenger receptor involved in angiogenesis, atherosclerosis, inflammation, and lipid metabolism. *J Clin Invest.* 2001;108(6):785-791. doi:10.1172/JCI14006
15. Horiuchi S, Sakamoto Y, Sakai M. Scavenger receptors for oxidized and glycated proteins. *Amino Acids.* 2003;25(3-4):283-292. doi:10.1007/s00726-003-0029-5
16. Park YM. CD36, a scavenger receptor implicated in atherosclerosis. *Exp Mol Med.* 2014;46(6):e99-e99. doi:10.1038/emm.2014.38
17. Abumrad NA, el-Maghrabi MR, Amri EZ, Lopez E, Grimaldi PA. Cloning of a rat adipocyte membrane protein implicated in binding or transport of long-chain fatty acids that is induced during preadipocyte differentiation. Homology with human CD36. *J Biol Chem.* 1993;268(24):17665-17668.
18. Stewart CR, Stuart LM, Wilkinson K, et al. CD36 ligands promote sterile inflammation through assembly of a Toll-like receptor 4 and 6 heterodimer. *Nat Immunol.* 2010;11(2):155-161. doi:10.1038/ni.1836
19. Philips JA, Rubin EJ, Perrimon N. *Drosophila* RNAi Screen Reveals CD36 Family Member Required for Mycobacterial Infection. *Science.* 2005;309(5738):1251-1253. doi:10.1126/science.1116006

20. Ren Y, Silverstein RL, Allen J, Savill J. CD36 gene transfer confers capacity for phagocytosis of cells undergoing apoptosis. *J Exp Med.* 1995;181(5):1857-1862. doi:10.1084/jem.181.5.1857
21. Sun M, Finnemann SC, Febbraio M, et al. Light-induced Oxidation of Photoreceptor Outer Segment Phospholipids Generates Ligands for CD36-mediated Phagocytosis by Retinal Pigment Epithelium. *J Biol Chem.* 2006;281(7):4222-4230. doi:10.1074/jbc.M509769200
22. Dobri AM, Dudău M, Enciu AM, Hinescu ME. CD36 in Alzheimer's Disease: An Overview of Molecular Mechanisms and Therapeutic Targeting. *Neuroscience.* 2021;453:301-311. doi:10.1016/j.neuroscience.2020.11.003
23. Hale JS, Li M, Sinyuk M, Jahnen-Dechent W, Lathia JD, Silverstein RL. Context Dependent Role of the CD36 - Thrombospondin - Histidine-Rich Glycoprotein Axis in Tumor Angiogenesis and Growth. Addison CL, ed. *PLoS ONE.* 2012;7(7):e40033. doi:10.1371/journal.pone.0040033
24. DeFilippis RA, Chang H, Dumont N, et al. CD36 Repression Activates a Multicellular Stromal Program Shared by High Mammographic Density and Tumor Tissues. *Cancer Discov.* 2012;2(9):826-839. doi:10.1158/2159-8290.CD-12-0107
25. Kitamura T, Doughty-Shenton D, Cassetta L, et al. Monocytes Differentiate to Immune Suppressive Precursors of Metastasis-Associated Macrophages in Mouse Models of Metastatic Breast Cancer. *Front Immunol.* 2018;8:2004. doi:10.3389/fimmu.2017.02004
26. Ladanyi A, Mukherjee A, Kenny HA, et al. Adipocyte-induced CD36 expression drives ovarian cancer progression and metastasis. *Oncogene.* 2018;37(17):2285-2301. doi:10.1038/s41388-017-0093-z
27. Pan J, Fan Z, Wang Z, et al. CD36 mediates palmitate acid-induced metastasis of gastric cancer via AKT/GSK-3 β / β -catenin pathway. *J Exp Clin Cancer Res.* 2019;38(1):52. doi:10.1186/s13046-019-1049-7
28. Hale JS, Otvos B, Sinyuk M, et al. Cancer Stem Cell-Specific Scavenger Receptor CD36 Drives Glioblastoma Progression. *Stem Cells.* 2014;32(7):1746-1758. doi:10.1002/stem.1716
29. Pascual G, Avgustinova A, Mejetta S, et al. Targeting metastasis-initiating cells through the fatty acid receptor CD36. *Nature.* 2017;541(7635):41-45. doi:10.1038/nature20791
30. Takaichi M, Tachinami H, Takatsuka D, et al. Targeting CD36-Mediated Lipid Metabolism by Selective Inhibitor-Augmented Antitumor Immune Responses in Oral Cancer. *Int J Mol Sci.* 2024;25(17):9438. doi:10.3390/ijms25179438
31. Wang J, Li Y. CD36 tango in cancer: signaling pathways and functions. *Theranostics.* 2019;9(17):4893-4908. doi:10.7150/thno.36037
32. Susztak K, Ciccone E, McCue P, Sharma K, Böttinger EP. Multiple Metabolic Hits Converge on CD36 as Novel Mediator of Tubular Epithelial Apoptosis in Diabetic Nephropathy. Shulman G, ed. *PLoS Med.* 2005;2(2):e45. doi:10.1371/journal.pmed.0020045
33. Feng WW, Zuppe HT, Kurokawa M. The Role of CD36 in Cancer Progression and Its Value as a Therapeutic Target. *Cells.* 2023;12(12):1605. doi:10.3390/cells12121605
34. Zhou X, Su M, Lu J, Li D, Niu X, Wang Y. CD36: The Bridge between Lipids and Tumors. *Molecules.* 2024;29(2):531. doi:10.3390/molecules29020531
35. Yang YL, Lin SH, Chuang LY, et al. CD36 is a novel and potential anti-fibrogenic target in albumin-induced renal proximal tubule fibrosis. *J Cell Biochem.* 2007;101(3):735-744. doi:10.1002/jcb.21236
36. Mitrofanova A, Burke G, Merscher S, Fornoni A. New insights into renal lipid dysmetabolism in diabetic kidney disease. *World J Diabetes.* 2021;12(5):524-540. doi:10.4239/wjdv12.i5.524
37. Cao W, Xu J, Zhou ZM, Wang GB, Hou FF, Nie J. Advanced Oxidation Protein Products Activate Intrarenal Renin-Angiotensin System via a CD36-Mediated, Redox-Dependent Pathway. *Antioxid Redox Signal.* 2013;18(1):19-35. doi:10.1089/ars.2012.4603
38. Zhang M, Gao X, Wu J, et al. Oxidized high-density lipoprotein enhances inflammatory activity in rat mesangial cells. *Diabetes Metab Res Rev.* 2010;26(6):455-463. doi:10.1002/dmrr.1102
39. Goto K, Iso T, Hanaoka H, et al. Peroxisome Proliferator-Activated Receptor- γ in Capillary Endothelia Promotes Fatty Acid Uptake by Heart During Long-Term Fasting. *J Am Heart Assoc.* 2013;2(1):e004861. doi:10.1161/JAHA.112.004861
40. Kennedy DJ, Chen Y, Huang W, et al. CD36 and Na/K-ATPase- α 1 Form a Proinflammatory Signaling Loop in Kidney. *Hypertension.* 2013;61(1):216-224. doi:10.1161/HYPERTENSIONAHA.112.198770
41. Gao X, Wu J, Qian Y, et al. Oxidized high-density lipoprotein impairs the function of human renal proximal tubule epithelial cells through CD36. *Int J Mol Med.* 2014;34(2):564-572. doi:10.3892/ijmm.2014.1799

42. O. Apostolov E, Ok E, Burns S, et al. Carbamylated-Oxidized LDL: Proatherosclerotic Effects on Endothelial Cells and Macrophages. *J Atheroscler Thromb.* 2013;20(12):878-892. doi:10.5551/jat.14035
43. Blevins HM, Xu Y, Biby S, Zhang S. The NLRP3 Inflammasome Pathway: A Review of Mechanisms and Inhibitors for the Treatment of Inflammatory Diseases. *Front Aging Neurosci.* 2022;14:879021. doi:10.3389/fnagi.2022.879021
44. Hou Y, Wang Q, Han B, Chen Y, Qiao X, Wang L. CD36 promotes NLRP3 inflammasome activation via the mtROS pathway in renal tubular epithelial cells of diabetic kidneys. *Cell Death Dis.* 2021;12(6):523. doi:10.1038/s41419-021-03813-6
45. Yang L, Sun J, Li M, et al. Oxidized Low-Density Lipoprotein Links Hypercholesterolemia and Bladder Cancer Aggressiveness by Promoting Cancer Stemness. *Cancer Res.* 2021;81(22):5720-5732. doi:10.1158/0008-5472.CAN-21-0646
46. Yang H, Li Q, Wang C, et al. Cytotoxic Necrotizing Factor 1 Downregulates CD36 Transcription in Macrophages to Induce Inflammation During Acute Urinary Tract Infections. *Front Immunol.* 2018;9:1987. doi:10.3389/fimmu.2018.01987
47. Byun JH, Lebeau PF, Platko K, et al. Inhibitory Antibodies against PCSK9 Reduce Surface CD36 and Mitigate Diet-Induced Renal Lipotoxicity. *Kidney360.* 2022;3(8):1394-1410. doi:10.34067/KID.0007022021
48. Pepino MY, Kuda O, Samovski D, Abumrad NA. Structure-Function of CD36 and Importance of Fatty Acid Signal Transduction in Fat Metabolism. *Annu Rev Nutr.* 2014;34(1):281-303. doi:10.1146/annurev-nutr-071812-161220
49. Bensadoun A, Mottler CD, Pelletier C, et al. A new monoclonal antibody, 4-1a, that binds to the amino terminus of human lipoprotein lipase. *Biochim Biophys Acta BBA - Mol Cell Biol Lipids.* 2014;1841(7):970-976. doi:10.1016/j.bbalip.2014.03.008
50. Silverstein RL, Febbraio M. CD36, a Scavenger Receptor Involved in Immunity, Metabolism, Angiogenesis, and Behavior. *Sci Signal.* 2009;2(72). doi:10.1126/scisignal.272re3
51. Kuroda N, Sugawara E, Kusano H, Yuba Y, Yorita K, Takeuchi K. A review of ALK-rearranged renal cell carcinomas with a focus on clinical and pathobiological aspects. *Pol J Pathol.* 2018;69(2):109-113. doi:10.5114/pjp.2018.76693
52. Grünwald V. Risikoadaptierte (Immun-)Therapie beim Nierenzellkarzinom. *Urol.* 2018;57(11):1326-1333. doi:10.1007/s00120-018-0791-3
53. Ding M, Lu X, Wang C, et al. The E2F1-miR-520/372/373-SPOP Axis Modulates Progression of Renal Carcinoma. *Cancer Res.* 2018;78(24):6771-6784. doi:10.1158/0008-5472.CAN-18-1662
54. Osmanov YI, Kogan EA, Gadzhieva ZK, Prochenko DD. [Ultrastructural features of histological variants of renal cell carcinoma]. *Urol Mosc Russ* 1999. 2022;(1):113-116.
55. Patard JJ, Leray E, Rioux-Leclercq N, et al. Prognostic Value of Histologic Subtypes in Renal Cell Carcinoma: A Multicenter Experience. *J Clin Oncol.* 2005;23(12):2763-2771. doi:10.1200/JCO.2005.07.055
56. Thoenes W, Störkel St, Rumpelt HJ. Histopathology and Classification of Renal Cell Tumors (Adenomas, Oncocytomas and Carcinomas). *Pathol - Res Pract.* 1986;181(2):125-143. doi:10.1016/S0344-0338(86)80001-2
57. Yang H, Zhao H, Ren Z, et al. Overexpression CPT1A reduces lipid accumulation via PPAR α /CD36 axis to suppress the cell proliferation in ccRCC. *Acta Biochim Biophys Sin.* 2022;54(2):220-231. doi:10.3724/abbs.2021023
58. Ackerman D, Tumanov S, Qiu B, et al. Triglycerides Promote Lipid Homeostasis during Hypoxic Stress by Balancing Fatty Acid Saturation. *Cell Rep.* 2018;24(10):2596-2605.e5. doi:10.1016/j.celrep.2018.08.015
59. Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg Effect: The Metabolic Requirements of Cell Proliferation. *Science.* 2009;324(5930):1029-1033. doi:10.1126/science.1160809
60. Qiu B, Ackerman D, Sanchez DJ, et al. HIF2 α -Dependent Lipid Storage Promotes Endoplasmic Reticulum Homeostasis in Clear-Cell Renal Cell Carcinoma. *Cancer Discov.* 2015;5(6):652-667. doi:10.1158/2159-8290.CD-14-1507
61. Seo J, Jeong DW, Park JW, Lee KW, Fukuda J, Chun YS. Fatty-acid-induced FABP5/HIF-1 reprograms lipid metabolism and enhances the proliferation of liver cancer cells. *Commun Biol.* 2020;3(1):638. doi:10.1038/s42003-020-01367-5
62. Liao M, Li Y, Xiao A, et al. HIF-2 α -induced upregulation of CD36 promotes the development of ccRCC. *Exp Cell Res.* 2022;421(2):113389. doi:10.1016/j.yexcr.2022.113389
63. Van Der Mijn JC, Fu L, Khani F, et al. Combined Metabolomics and Genome-Wide Transcriptomics Analyses Show Multiple HIF1 α -Induced Changes in Lipid Metabolism in Early Stage Clear Cell Renal Cell

- Carcinoma. *Transl Oncol.* 2020;13(2):177-185. doi:10.1016/j.tranon.2019.10.015
64. Bouraoui Y, Said R, Bruss C, et al. Increased expression of CD36 and CD163 in clear cell renal cell carcinoma suggests an association between lipid transport and an “M2-like” macrophage phenotype. *Front Immunol.* 2026;17:1773666. doi:10.3389/fimmu.2026.1773666
65. Su P, Wang Q, Bi E, et al. Enhanced Lipid Accumulation and Metabolism Are Required for the Differentiation and Activation of Tumor-Associated Macrophages. *Cancer Res.* 2020;80(7):1438-1450. doi:10.1158/0008-5472.CAN-19-2994
66. Wallace DMA, Raghavan D, Kelly KA, et al. Neoadjuvant (Pre-emptive) Cisplatin Therapy in Invasive Transitional Cell Carcinoma of the Bladder. *Br J Urol.* 1991;67(6):608-615. doi:10.1111/j.1464-410X.1991.tb15225.x
67. Xia L, Zhou Z, Chen X, et al. Ligand-dependent CD36 functions in cancer progression, metastasis, immune response, and drug resistance. *Biomed Pharmacother.* 2023;168:115834. doi:10.1016/j.biopha.2023.115834
68. Pardo JC, Sanhueza T, Ruiz De Porras V, et al. Prognostic Impact of CD36 Immunohistochemical Expression in Patients with Muscle-Invasive Bladder Cancer Treated with Cystectomy and Adjuvant Chemotherapy. *J Clin Med.* 2022;11(3):497. doi:10.3390/jcm11030497
69. Jeong H, Oh HE, Kim H, et al. Upregulation of Fatty Acid Transporters is Associated With Tumor Progression in Non-Muscle-Invasive Bladder Cancer. *Pathol Oncol Res.* 2021;27:594705. doi:10.3389/pore.2021.594705
70. Shang D, Zheng T, Zhang J, Tian Y, Liu Y. Profiling of mRNA and long non-coding RNA of urothelial cancer in recipients after renal transplantation. *Tumor Biol.* 2016;37(9):12673-12684. doi:10.1007/s13277-016-5148-1
71. Pavalean MI, Dobre M, Pelisenco IA, Madan VL, Milanese E, Hinescu ME. Association of miRNA-17-92 Cluster with Muscle Invasion in Bladder Cancer. *Int J Mol Sci.* 2025;26(15):7546. doi:10.3390/ijms26157546
72. Thompson RH, Kwon ED. Significance of B7-H1 Overexpression in Kidney Cancer. *Clin Genitourin Cancer.* 2006;5(3):206-211. doi:10.3816/CGC.2006.n.038
73. Bui MHT, Visapaa H, Seligson D, et al. Prognostic Value Of Carbonic Anhydrase Ix And Ki67 As Predictors Of Survival For Renal Clear Cell Carcinoma. *J Urol.* 2004;171(6 Part 1):2461-2466. doi:10.1097/01.ju.0000116444.08690.e2
74. Klatte T, Seligson DB, Riggs SB, et al. Hypoxia-Inducible Factor 1 α in Clear Cell Renal Cell Carcinoma. *Clin Cancer Res.* 2007;13(24):7388-7393. doi:10.1158/1078-0432.CCR-07-0411
75. Li Z, Kang Y. Lipid Metabolism Fuels Cancer’s Spread. *Cell Metab.* 2017;25(2):228-230. doi:10.1016/j.cmet.2017.01.016
76. Foguer K, Braga MDS, Peron JPS, Bortoluci KR, Bellini MH. Endostatin gene therapy inhibits intratumoral macrophage M2 polarization. *Biomed Pharmacother.* 2016;79:102-111. doi:10.1016/j.biopha.2016.01.035
77. Presti JC, Wilhelm M, Reuter V, Russo P, Motzer R, Waldman F. Allelic loss on chromosomes 8 and 9 correlates with clinical outcome in locally advanced clear cell carcinoma of the kidney. *J Urol.* 2002;167(3):1464-1468.
78. Hakimi AA, Ostrovnaya I, Reva B, et al. Adverse Outcomes in Clear Cell Renal Cell Carcinoma with Mutations of 3p21 Epigenetic Regulators BAP1 and SETD2 : A Report by MSKCC and the KIRC TCGA Research Network. *Clin Cancer Res.* 2013;19(12):3259-3267. doi:10.1158/1078-0432.CCR-12-3886
79. Saber MM. Coexpression of PD-L1/PD-1 with CXCR3/CD36 and IL-19 Increase in Extranodal Lymphoma. *Tang B, ed. J Immunol Res.* 2023;2023:1-17. doi:10.1155/2023/4556586
80. Sakurai K, Tomihara K, Yamazaki M, et al. CD36 expression on oral squamous cell carcinoma cells correlates with enhanced proliferation and migratory activity. *Oral Dis.* 2020;26(4):745-755. doi:10.1111/odi.13210
81. Qu J, Li D, Jin J, et al. Hypoxia-Inducible Factor 2 α Attenuates Renal Ischemia-Reperfusion Injury by Suppressing CD36-Mediated Lipid Accumulation in Dendritic Cells in a Mouse Model. *J Am Soc Nephrol.* 2023;34(1):73-87. doi:10.1681/ASN.0000000000000027
82. Guerrero-Rodríguez SL, Mata-Cruz C, Pérez-Tapia SM, Velasco-Velázquez MA. Role of CD36 in cancer progression, stemness, and targeting. *Front Cell Dev Biol.* 2022;10:1079076. doi:10.3389/fcell.2022.1079076
83. De Góes Rocha FG, Chaves KCB, Chammas R, et al. Endostatin gene therapy enhances the efficacy of IL-2 in suppressing metastatic renal cell carcinoma in mice. *Cancer Immunol Immunother.* 2010;59(9):1357-1365. doi:10.1007/s00262-010-0865-6
84. Yang P bo, Hou P pei, Liu F yuan, et al. Blocking PPAR γ interaction facilitates Nur77 interdiction of fatty acid uptake and suppresses breast cancer

- progression. Proc Natl Acad Sci. 2020;117(44):27412-27422. doi:10.1073/pnas.2002997117
85. Deng M, Cai X, Long L, et al. CD36 promotes the epithelial–mesenchymal transition and metastasis in cervical cancer by interacting with TGF- β . J Transl Med. 2019;17(1):352. doi:10.1186/s12967-019-2098-6
86. Li C, Zhang L, Qiu Z, Deng W, Wang W. Key Molecules of Fatty Acid Metabolism in Gastric Cancer. Biomolecules. 2022;12(5):706. doi:10.3390/biom12050706
87. Watt MJ, Clark AK, Selth LA, et al. Suppressing fatty acid uptake has therapeutic effects in preclinical models of prostate cancer. Sci Transl Med. 2019;11(478):eaau5758. doi:10.1126/scitranslmed.aau5758
88. Raggi C, Taddei ML, Rae C, Braconi C, Marra F. Metabolic reprogramming in cholangiocarcinoma. J Hepatol. 2022;77(3):849-864. doi:10.1016/j.jhep.2022.04.038
89. Drury J, Rychahou PG, He D, et al. Inhibition of Fatty Acid Synthase Upregulates Expression of CD36 to Sustain Proliferation of Colorectal Cancer Cells. Front Oncol. 2020;10:1185. doi:10.3389/fonc.2020.01185
90. Zhang Y, Guo H, Zhang Z, Lu W, Zhu J, Shi J. IL-6 promotes chemoresistance via upregulating CD36 mediated fatty acids uptake in acute myeloid leukemia. Exp Cell Res. 2022;415(1):113112. doi:10.1016/j.yexcr.2022.113112
91. Xu WH, Qu YY, Wang J, et al. Elevated CD36 expression correlates with increased visceral adipose tissue and predicts poor prognosis in ccRCC patients. J Cancer. 2019;10(19):4522-4531. doi:10.7150/jca.30989
92. Kim YS, Jung J, Jeong H, et al. High Membranous Expression of Fatty Acid Transport Protein 4 Is Associated with Tumorigenesis and Tumor Progression in Clear Cell Renal Cell Carcinoma. Dis Markers. 2019;2019:1-7. doi:10.1155/2019/5702026
93. Yang Q, Chu W, Yang W, et al. Identification of RNA Transcript Makers Associated With Prognosis of Kidney Renal Clear Cell Carcinoma by a Competing Endogenous RNA Network Analysis. Front Genet. 2020;11:540094. doi:10.3389/fgene.2020.540094
94. Li K, Lv J, Wang J, et al. CircZNF609 inhibited bladder cancer immunotherapy sensitivity via enhancing fatty acid uptake through IGF2BP2/CD36 pathway. Int Immunopharmacol. 2024;137:112485. doi:10.1016/j.intimp.2024.112485
95. Guo CC, Dadhania V, Zhang L, et al. Gene Expression Profile of the Clinically Aggressive Micropapillary Variant of Bladder Cancer. Eur Urol. 2016;70(4):611-620. doi:10.1016/j.eururo.2016.02.056

This article is an Open Access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited and it is not used for commercial purposes; 2026, Pavalean et al., Applied Systems and Discoveries Journals.