

REVIEW Article

Advances in the prevention and prenatal treatment of spina bifida

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ABSTRACT

Spina bifida is a neural tube defect (NTD) that arises when the neural tube fails to close properly during early development. This review focuses on myelomeningocele (MMC), the most common severe form of spina bifida, which often leads to motor and sensory impairments, including lower limb weakness or paralysis, as well as renal, urological, orthopedic, developmental, and psychosocial challenges. We explore the etiology, pathogenesis, prevention, diagnosis, and management of spina bifida, with a special emphasis on in-utero surgical repair. Over the past several decades, researchers and clinicians have made remarkable strides across all stages of care from prevention to postnatal outcomes. Widespread use of folic acid supplementation has significantly reduced the number of new cases. Advances in prenatal imaging and diagnostics now allow for earlier and more accurate detection, enabling timely intervention. In-utero surgical techniques continue to evolve, with innovative hybrid approaches that combine the strengths of open and minimally invasive methods. The momentum in this field shows no sign of slowing. Promising developments in stem cell therapy, biomaterials, robotic-assisted surgery, 3D printing, and enhanced imaging are redefining

treatment goals in spina bifida. With each advance, clinicians gain better tools to improve outcomes for both mother and child, minimizing risks and maximizing long-term health and quality of life for both patients.

Keywords

Spina bifida, myelomeningocele, prenatal surgery, fetal surgery, fetal therapy, stem cells, biomaterials, neural tube defects.

Abbreviations

American College of Obstetricians and Gynecologists (ACOG); body mass index (BMI); cerebrospinal fluid (CSF); Children's Hospital of Philadelphia (CHOP); Cellular Therapy for In Utero Repair of Myelomeningocele Trial (CuRe Trial); European cardiovascular magnetic resonance (EuroCMR); Fetoscopic Robotic Open Spina Bifida Treatment (FROST); Myelomeningocele (MMC); Myeloschisis (MS); North American Fetal Therapy Network (NAFTNet); Neural tube defect (NTD); Placental mesenchymal stromal cell (PMSC); Preterm premature rupture of membranes (PPROM); Society for Cardiovascular Magnetic Resonance (SCMR); Transamniotic stem cell therapy (TRASCET).

SUMMARY

1. *Introduction*
2. *Spina Bifida Prognosis*
3. *Spina Bifida Prevention: Folic Acid and NTDs*
4. *Spina Bifida Diagnosis*
5. *In-Utero Treatment for Spina Bifida*
6. *In-Utero Treatment for Spina Bifida: Recent Advancements of Note*
7. *Unresolved Challenges*
8. *Conclusions*

1. Introduction

Spina bifida is a serious congenital condition caused by the incomplete closure of the neural tube during early embryonic development. Its prevalence varies widely across regions, with a global average ranging from 3.52 to 24.31 per 10,000 births.¹ In the United States, spina bifida affects approximately 3.59 per 10,000 live births.² Over the past three decades, early diagnosis and surgical innovation, especially prenatal intervention, have transformed the outlook for children with spina bifida. The landmark MOMS (Management of Myelomeningocele Study) trial and follow-up studies have demonstrated that fetal repair of myelomeningocele (MMC) can significantly improve motor outcomes and reduce the need for cerebrospinal fluid (CSF) shunting.^{3,4} Today, clinicians recognize the fetus as a patient in their own right, with a wide range of prenatal diagnostic tools and treatment options available. Advances in fetal medicine now allow healthcare teams not only to plan the timing, mode, and place of delivery—but in many cases, to treat the condition before birth. For families facing a spina bifida diagnosis, fetal surgery presents a hopeful alternative to termination and the potential to dramatically improve quality of life. This review synthesizes the current state of knowledge on open spina bifida, with a focus on prenatal surgery and outcome optimization.

Spina Bifida, a Neural Tube Defect

Spina bifida is a neural tube defect (NTD) that arises when the neural tube fails to close properly at the caudal (lower) end during early embryonic development.⁵ The neural plate, which formed from ectoderm around 17 days after conception,⁶ undergoes primary and secondary neurulation to form the brain and spinal cord. Primary neurulation creates a hollow tube through folding and separation from the

surface around day 22, while secondary neurulation forms the lower spinal cord through the hollowing of a solid cell cord around day 26.^{5,7,8} Neural tube closure occurs in a zipper-like fashion across multiple regions, guided by signals from the notochord and surrounding tissues⁹ (see **Figure 1**). Neural tube defects are classified as open or closed depending on whether neural tissue is exposed to the intrauterine environment. In closed defects, neural elements are sealed off from the intrauterine environment; in open defects, neural elements, or a cyst containing neural elements, remain exposed to the amniotic fluid.⁸

In MMC, the most common severe form of spina bifida, neural elements protrude through an opening in the spine. In closed MMC, the meninges and spinal cord remain sealed off from the intrauterine environment inside a cyst. In contrast, in open MMC, the neural elements are directly exposed to amniotic fluid, which causes further injury.⁸ The commonly accepted “two-hit hypothesis” explains MMC pathogenesis: first a failure of neurulation, followed by progressive in-utero injury to exposed neural tissue.¹⁰ In the most severe, but rare form of spina bifida, myeloschisis (MS), the spinal cord is completely open on the back, forming a flat plate of neural tissue, or a “placode”.^{11,12} In both MMC and MS the caudal end of the spinal cord may remain fixed or tethered to the vertebral column, pulling the hindbrain into the spinal cord, known as hindbrain herniation, and blocking the flow of CSF resulting in hydrocephalus.¹⁰

2. Spina Bifida Prognosis

Myelomeningocele involves a wide range of motor, sensory, and developmental challenges. Children with MMC may experience lower limb weakness or paralysis, loss of sensation, and orthopedic conditions such as clubfoot, scoliosis, kyphosis (outward curvature of the spine), or hip dislocations.¹¹⁻¹³ Many also face complications involving the renal and urological systems, including infections, and incontinence.¹¹⁻¹³ Neurologically, MMC is often associated with hindbrain herniation, also known as Chiari II malformation, and hydrocephalus, which frequently requires cerebrospinal fluid diversion through ventriculoperitoneal shunting or endoscopic third ventriculostomy.¹¹⁻¹³ Some individuals may also experience seizures, along with cognitive and psychosocial difficulties.¹¹⁻¹³ Despite these challenges, survival rates have steadily improved, with 20-year survival ranging from 50% to 87%

depending on the presence of hydrocephalus.¹⁴ Clinicians can now manage MMC more effectively thanks to advances in both prenatal and postnatal surgical techniques. Once viewed primarily as a life-threatening condition, MMC is now increasingly treated as a quality-of-life issue. The advent of fetal surgery has shifted treatment goals toward maximizing independence and developmental potential for affected children.¹⁵ As medicine and technology have improved outcomes for children with spina bifida, one of the most powerful advances has been preventing the condition in the first place through folic acid supplementation.

3. Spina Bifida Prevention: Folic Acid and NTDs

Folic acid deficiency is a major contributor to the development of NTDs. Because folates cannot be synthesized from scratch, they must be obtained in the diet.¹⁶ Folic acid plays a critical role in synthesizing thymidylates, purines, and other components essential to the creation of DNA and RNA.¹⁶ In this role, folate plays a key role in cell proliferation and neurulation, processes vital for neural tube closure.⁸ This makes adequate folic acid intake especially important during early pregnancy, when the neural tube is forming. Pregnant women require five to ten times more folate than their age-matched peers.¹⁷

Early studies in the 1980s suggested that folic acid supplementation reduced the risk of NTDs, though initial trials were limited by small sample sizes¹⁸ and methodological concerns.¹⁹ In response, the UK Medical Research Council conducted a large, randomized controlled trial in 1991, which confirmed that a daily 4,000 µg dose of folic acid significantly reduced NTD recurrence.²⁰ More recently, a large meta-analysis found that folic acid supplementation reduces the risk of NTDs by approximately two-thirds, although it did not demonstrate a clear protective effect for other congenital anomalies.²¹ Furthermore, a multicenter randomized controlled trial showed that supplementation with 4 mg of folic acid per day, compared with 0.4 mg, significantly reduced the occurrence of multiple birth defects, including NTDs.²² Globally, a daily intake of at least 400 µg of folic acid is recommended periconceptionally.²³

Because neural tube closure often occurs before pregnancy is recognized (see **Figure 1**), relying solely on prenatal vitamin use may fail to protect many pregnancies. This recognition prompted

widespread public health interventions, including mandatory folic acid fortification of cereal grain products in the United States beginning in 1998.⁸ Researchers predict that about 1300 fewer babies are given NTD diagnoses in the United States every year because of this folic acid fortification.²⁴ Similar reductions in NTDs have been observed in other countries including Chile, Canada, Brazil and South Africa upon the introduction of folic acid enrichment programs.^{25–28} Currently, countries with folic acid enrichment programs have fewer cases of spina bifida (35.2 vs. 52.3 per 100,000 live births), even after accounting for stillbirth and termination of pregnancy.¹ Despite these successes, not all countries have adopted fortification policies, leaving continued opportunities for global prevention.^{1,8,29} While broader access to folic acid could prevent many cases of spina bifida worldwide, accurate and timely diagnosis remains crucial for babies who are still affected.

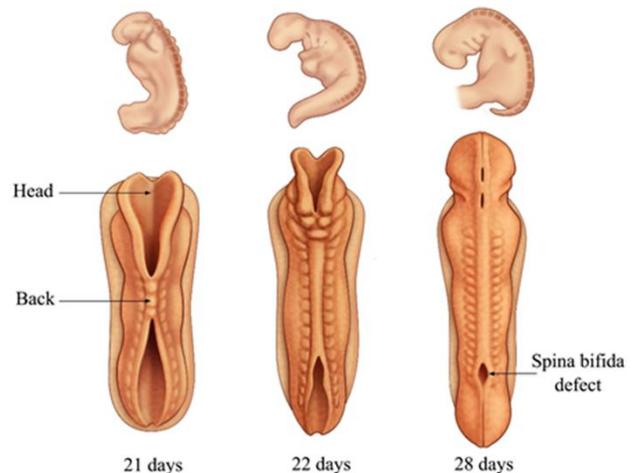


Figure 1. Folic acid makes major contributions to the development of the neural tube. Neural tube formation is often complete before a woman knows that she is pregnant because she first expects her period about 15 days after conception. Countries such as the United States started adding 140 µg of folic acid to every 100 grams of enriched grains in the 1990s, to ensure that women have enough folic acid during the critical first month of pregnancy. This reduced nationwide cases of spina bifida by up to 2/3.⁸ Image reproduced from reference 13, with permission.

4. Spina Bifida Diagnosis

MMC is primarily diagnosed using prenatal ultrasound. Ultrasound interpretation is inherently

subjective and dependent on fetal positioning; however, recent advancements have improved diagnostic accuracy. Computerized support software systems, such as those developed by Cengizler et al. allow for the algorithmic identification of the fetal spine, limiting subjectivity.^{30,31} Machine learning-based analyses have also been done to study the sonographer workflow, finding that each scan is a “non-ordered multistep process of anatomical structure acquisition” due in part to a need to take advantage of fetal position.³² These recent findings have helped standardize techniques while highlighting variables that can be improved upon. Ultrasound has also been shown to be a reliable diagnostic approach for MMC regardless of maternal body mass index (BMI),³³ and cranial, spinal, ventricular, muscular, and post-surgical markers of MMC have been identified which further enhance diagnostic accuracy.^{34–39} In many cases, ultrasound is also often supplemented with molecular and biochemical testing, magnetic resonance imaging, and echocardiography^{39–47} which help to confirm or clarify findings. Novel imaging and diagnostic techniques have also greatly advanced the capability of clinicians to identify these defects,⁴⁸ including next-generation whole-exome sequencing technologies,⁴⁹ which have led to the identification of several novel mutations associated with NTDs.^{50–57} Advances in prenatal diagnosis made it possible not only to detect spina bifida earlier, but also to consider surgical repair before birth.

5. In-Utero Treatment for Spina Bifida

History

Historically, open spina bifida has been managed with postnatal surgical repair, which involves removing exposed tissue, dissecting a connective strand at the base of the spinal cord called the filum terminale, and surgically folding the spinal cord into a tube through a process known as tubularization. While this approach protects the spinal cord and prevents further damage, it does not reverse neurological deficits. Recognizing the limitations of postnatal repair, researchers began exploring the potential of earlier intervention, specifically in-utero surgery, to improve outcomes by halting or reducing neurological injury before birth.

Initial experiments in the 1980s and 1990s demonstrated the feasibility of fetal repair in animal models, including primates,⁵⁸ rats,^{59,60} pigs,⁶⁰ and lambs.^{61,62} These promising results paved the way for human trials. In 1997, Meuli-Simmen and colleagues reported the first successful technical attempt at in-utero MMC repair in humans.^{63,64} Early cases faced challenges: the first two fetal surgeries used a maternal skin graft to cover the exposed neural tissue - one baby survived to one year, while the other died due to extreme prematurity.⁶⁵ However, rapid progress followed. By 1998 and 1999, teams at the Children’s Hospital of Philadelphia (CHOP) and Vanderbilt University reported improved outcomes with refined surgical techniques.^{66–69} Later, in 2003, Dr. Harrison’s team at the University of California, San Francisco reported their attempt at fetoscopic MMC repair.⁷⁰ Complete fetoscopic repair was only accomplished in one case, with two other fetuses undergoing partial fetoscopic repair. Dr. Harrison’s team then temporarily abandoned the fetoscopic approach for the remaining 10 fetuses, opting instead to utilize an open approach.⁷⁰ Efforts to optimize fetal surgical techniques culminated in the landmark Management of Myelomeningocele Study (MOMS) trial, published in 2011,³ which confirmed that prenatal surgery significantly improves motor outcomes and reduces the need for CSF shunting, marking a turning point in the standard of care. Notable clinical trials advancing prenatal spina bifida repair can be found in **Table 1**.

The MOMS Trial & Open Hysterotomy Spina Bifida Repair

The MOMS Trial showed that prenatal surgery vastly improved the child’s mental and motor outcomes compared to postnatal surgery.³ In the study, clinicians utilized an open-hysterotomy prenatal closure approach, comparing outcomes to children treated postnatally. In this approach, a laparotomy incision is made to expose and exteriorize the uterus. A uterine incision is then made to expose the MMC, using ultrasound to guide the location of the hysterotomy. The neural placode is dissected and dropped into the spinal canal. The dura, skin, and uterus are then closed.⁷² During the MOMS Trial, participating centers coordinated with non-participating centers, who agreed not to perform

Table 1. Notable clinical trials advancing prenatal spina bifida repair

Clinical Trial	Outcomes	Year	Ref
Management of Myelomeningocele Study (MOMS) & Follow-up	Prenatal (open hysterotomy) repair is superior to postnatal repair in terms of shunt rate, reversal of hindbrain herniation, ambulation, and other metrics.	2011-2022	3,71-77
Cirurgia Endoscópica para Correção Antenatal da Meningomielocele (CECAM)	Minimally invasive fetoscopic repair is feasible, albeit with risks such as membrane rupture.	2016	78
Fetoscopic Meningomyelocele Repair Study (fMMC) & Follow-up	Hybrid laparotomy-assisted fetoscopic repair is feasible, with similar benefits to open repair and lowered risk of prematurity.	2017-2025	79-81
Fetoscopic NEOX Cord 1K® Spina Bifida Repair	Investigating feasibility of human umbilical cord patches for defect coverage.	Ongoing	82
Cellular Therapy for In Utero Repair of Myelomeningocele - The Cure Trial (CuRe)	Investigating application of stem cells to defect. Ongoing but thus far positive outcomes.	Ongoing	83,84

prenatal MMC surgery while the trial was ongoing and instead referred to MOMS Trial centers, allowing robust data to be obtained. Surgically repaired children have since been followed, with updates being published at 12 months, 30 months, and 5-10 years, providing ample evidence of the short- and long-term benefits of in-utero spina bifida surgery to the unborn child.

The MOMS Trial demonstrated remarkable results, and in fact was stopped early due to the efficacy of prenatal MMC repair over postnatal repair, particularly in terms of hindbrain herniation reversal and hydrocephalus. Prenatally repaired children required hydrocephalus management by CSF diversion via shunt placement at less than half the rate of postnatally repaired children (40% vs. 82%). Partial or total hindbrain herniation reversal was also markedly different, with 75% of prenatally repaired children showing no moderate or severe hindbrain herniation and 36% showing complete hindbrain herniation resolution, compared to 33% and 4% of postnatally repaired children, respectively. The incidence of brainstem kinking was also lower in prenatally repaired children (20% vs. 49%), as was abnormal 4th ventricular location (46% vs. 72%) and rates of spinal cord cysts called syringomyelia (39% vs. 58%).³ An update was published outlining all 12-

month outcome data and expanding upon the initially presented findings. The reduced need for CSF shunting was maintained (44% vs. 83.6%), and this follow-up data revealed that prenatally repaired children also needed fewer shunt revisions after placement (15.4% vs. 40.2%).⁷³ Amazingly, even at the 5–10-year follow-up, prenatally repaired children required fewer shunts (49% vs. 85%) and fewer shunt revisions (23% vs. 60%), and the rate of total hindbrain herniation reversal had climbed and remained markedly improved (39% vs. 13%).⁷⁴ Rates of syringomyelia also remained starkly different (59% vs. 81%). Most critically, neonatal death rates remained low and comparable between groups across this time range, with a 95% survival rate in the prenatal surgery group (5 total deaths of 91) and a 97% survival rate in the postnatal surgery group (3 total deaths of 92).^{71,74}

As reduced lower body motor function and paralysis are particularly detrimental impacts of spina bifida, MOMS Trial investigators also longitudinally measured motor function improvements, and the results were similarly impressive. 44.8% of prenatally repaired children were walking independently at their 30-month follow-up exam, 27.6% were walking with the assistance of orthotics or devices, and only 27.6% were not walking. In marked contrast, the postnatally

repaired group had rates of 23.9% (independent), 35.3% (assisted), and 40.9% (not walking).⁷⁵ Prenatally repaired children also exhibited lower rates of leg-length discrepancy, and required fewer orthopedic treatments with casts or braces at both 12 and 30 months.⁷⁷ In addition, prenatally repaired children demonstrated greater object manipulation, general locomotion and mobility, and stationary motor function than postnatally repaired children. In fact, these improvements in motor function were more often greater than expected and less often worse than expected based on anatomical measures, in comparison to postnatally repaired children.^{3,75} At the 5–10-year follow-up, these differences persisted, and prenatally repaired children were more likely to walk independently, less likely to require a wheelchair, and more likely to be capable of walking outdoors than their postnatally repaired counterparts.^{71,74} These physical improvements translated into greater self-care capabilities. Prenatally repaired children were more competent chewing and swallowing, using a fork, brushing their teeth, washing and drying their hands, removing their socks and shoes, and zipping, showcasing the real-world benefits to these children.⁷¹ Prenatally repaired children were even found to be taller than their postnatally repaired counterparts.⁷⁴

Besides physical improvements, prenatally repaired children in the MOMS Trial exhibited several mental, developmental, and social improvements, particularly in comparison to the postnatally repaired children. Prenatally repaired children exhibited greater psychomotor development than those repaired postnatally,^{3,75} and greater verbal learning, nonverbal reasoning, and reading scores.⁷⁴ In fact, on cognitive and neurological measures for which prenatally repaired children did not outperform postnatally repaired children, no robust differences were noted besides fine motor control.⁷⁴ Socially and developmentally, prenatally repaired children experienced a higher quality of life with less negative impact on their families, with the differences being maintained through the 5–10-year follow-up.^{74,76} Outside of the MOMS Trial, comparisons of outcomes between postnatal and prenatal repair are still relatively limited.⁸⁵ However, several non-MOMS studies have validated these findings, demonstrating similarly improved neonatal outcomes including a reduced need for a shunt or shunt revision, higher rates of hindbrain herniation resolution, improved neonatal functional level, and shorter

length of stay in neonatal intensive care units than postnatal repair.^{86–88} Another study showed that fetal spina bifida repair may also help normalize cerebral blood flow.⁸⁹ Similar benefits to ambulation have been shown in non-MOMS studies as well,^{80,90,91} with one study even showing prenatally repaired children participating in sports activities at later time points.⁹¹ Non-MOMS studies have found prenatally repaired children exhibit adaptive behavior, executive functioning abilities, neurocognitive outcomes, psychoeducational achievement, and memory scores within the age-matched population norms, albeit with delays or deficits, particularly for children with shunts.^{91–93}

Open hysterotomy-based prenatal spina bifida surgery is not without its drawbacks, however (see **Table 2**). As with many other prenatal surgeries, open MMC repair is more strongly associated with preterm delivery, low birth weight, and other obstetric issues such as uterine rupture, membrane separation or rupture (including preterm premature rupture of membranes or PPRM), maternal pulmonary edema, placental abruption, and oligohydramnios (reduced amniotic fluid) than postnatal repair.^{3,67,80,86–88,94,95} However, the risk of some of these complications is notably similar to the risk following classical c-section.^{94,96,97} Prenatal surgery has also been more strongly associated with certain perinatal complications, such as respiratory distress syndrome and bradycardia.^{3,80} Other findings, particularly on urological function post-repair, are mixed. Both MOMS and non-MOMS studies have shown positive findings, such as prenatally repaired children being more likely to void volitionally and have lower rates of urinary tract infection, but there are also negative findings such as a higher prevalence of bladder muscle overactivity compared to postnatally repaired children.^{98,99} As the field of fetal surgery continues to evolve, advancements will likely improve the outlook for mothers and children following open prenatal MMC repair.¹⁰⁰

Advances in Open Hysterotomy-Based Prenatal MMC Repair

Following the MOMS Trial, inclusion criteria for open prenatal MMC repair have expanded. Although high BMI was initially an exclusion factor, later studies have shown comparable perioperative outcomes in women with higher BMI.¹⁰¹ Access has also expanded to include women with gestational

diabetes, certain fetal abnormalities, maternal infections and Rh alloimmunization,^{4,102} and discussion continues to be had about further loosening eligibility criteria.¹⁰³ CHOP has also developed a new closure technique, which uses a needlepoint monopolar cautery to raise myofascial flaps, that has been shown to result in reduced rates of CSF diversion, more significant closure, reductions in hindbrain herniation and shunting, and overall improved results.^{4,104} Due to these technical modifications and increased experience, outcomes reported post-MOMS by CHOP have improved. As of 2017, PPRM rates had dropped from the 46% observed in MOMS to 32%, repair site dehiscence (rupture) rates had dropped from 13% to 3.6%, and pulmonary edema, the need for maternal blood transfusion, and severe preterm birth rates all dropped substantially as well.^{4,88}

Even postnatal repair continues to improve. Intraoperative neurophysiological monitoring, which has been used extensively in spinal surgery already, has been recently shown to be useful in postnatal neonatal spina bifida repair as well.¹⁰⁵

Several other approaches in addition to the open hysterotomy-based repair technique have also developed, which offer unique benefits over postnatal and open prenatal repair. These include percutaneous fetoscopic repair, laparotomy-assisted fetoscopic repair, and a percutaneous/mini-laparotomy fetoscopic technique.^{78,81,106–111} Anesthetics and analgesics are administered to both the mother and the fetus during all these procedures.^{78,108,112}

Percutaneous Fetoscopic Repair

Percutaneous fetoscopic repair of MMC is a minimally invasive alternative to open fetal surgery that has evolved significantly over time. The procedure involves inserting instruments through small punctures in the maternal abdomen and uterus, dissecting the neural placode, covering it with a patch, and closing the defect, all without a large uterine incision. Unlike open hysterotomy, which requires cesarean section, percutaneous fetoscopic repair allows for vaginal delivery, and has been associated with a lower risk of maternal complications such as uterine thinning, rupture, and pulmonary edema (see **Table 2**).^{113–116} However, percutaneous fetoscopy carries higher rates of placental abruption, PPRM, and earlier gestational age at delivery, contributing to complications like respiratory distress syndrome, site

dehiscence, sepsis, and increased need for CSF diversion.^{107,115–118} Despite these risks, some studies have reported better neurological outcomes, including higher rates of hindbrain herniation reversal, urinary continence, and independent walking compared to open repair.^{115,118}

Technological refinements have improved outcomes over time. The technique has been adapted to close larger spinal defects with comparable success, and the gestational age window for surgery has been extended slightly later (to 27+6 weeks) without compromising outcomes, which helps reduce the risk of extreme prematurity.^{119,120} Most recently, hybrid surgical approaches have emerged, aiming to leverage the benefits of both open and fetoscopic techniques while limiting the drawbacks, reflecting the continued drive to optimize safety and effectiveness for both mother and child.

Hybrid Approaches

The hybrid laparotomy-assisted fetoscopic approach, developed in part by Texas Children's Fetal Center,⁷⁹ combines elements of open and fetoscopic MMC repair. Surgeons begin with a maternal abdominal incision to partially exteriorize the uterus. The membranes are secured to the uterine wall, amniotic fluid is replaced with carbon dioxide, and a fetoscope is inserted. The neural placode is dissected and closed fetoscopically.^{81,112} This hybrid repair offers several benefits over earlier techniques. Unlike open surgery, it allows for vaginal delivery and achieves higher gestational age at birth than both open and percutaneous repairs.^{80,87,107,116,121–123} Vaginal delivery following laparotomy-assisted fetoscopic repair has also been associated with shorter length of stay in neonatal intensive care units.¹²² While maternal complication rates (e.g., PPRM, edema, preeclampsia) are generally similar to open repair, laparotomy-assisted fetoscopic is associated with lower rates of uterine thinning or dehiscence.⁸⁰ Neonatal complication rates are similarly very comparable.^{80,121} Though laparotomy-assisted fetoscopic repair takes longer than open repair and initially raised concerns about carbon dioxide use, studies have shown no adverse impact on fetal growth or long-term development.¹²⁴ Importantly, when factors such as gestational age at delivery, age at time of outcome measurement, and presence of hydrocephalus treatment are controlled for, long term

motor and neurodevelopmental outcomes are also comparable to open repair.^{80,121,125,126}

Over time, technical refinements in the laparotomy-assisted fetoscopic approach have improved results. A multilayered closure technique reduced CSF leakage, skin dehiscence, and tethered cord, while improving rates of hindbrain herniation reversal.⁸⁰ More recently, a California team introduced a “percutaneous/mini-laparotomy” hybrid approach, which uses a small incision and camera port for fetoscopic closure. Early results suggest a lower risk of prematurity than percutaneous repair and the possibility of vaginal delivery, though PPROM remains a challenge.¹⁰⁸ Together, these innovations reflect the ongoing evolution of fetal surgery, balancing safety and precision while expanding treatment options for spina bifida.

As of 2017, ACOG and NAFTNet recommend providing women with a fetal MMC diagnosis the option to receive open fetal repair. They do not yet recommend fetoscopic fetal repair due to limited data.¹²⁷ Although open fetal surgery is currently recommended by ACOG, emerging bioengineering strategies are introducing new tools and techniques that could further improve outcomes. Several surgical approaches are shown in **Table 2**.

6. In-Utero Treatment for Spina Bifida: Recent Advancements of Note

Additional improvements to open and fetoscopic approaches, as well as to postnatal repair procedures not outlined above have improved the outcomes and processes further. Modified closure techniques have also been tested since the MOMS trial. Two-layer closure (myofascial and skin) has been shown to improve watertightness and outcomes,¹²⁸ and another more recently developed 3-port, 3-layer fetoscopic repair technique may further improve outcomes and lead to higher rates of watertight closure.^{129,130} MOMS3, a follow-up of MOMS participants in teen and young adult years, is also currently planned and enrolling study participants.¹³¹ There are also numerous groundbreaking advancements being developed that make use of stem cells and bioengineered tissues, and which leverage other technological modalities such as robotics, 3D printing, and plastic surgery.

Stem Cell and Biomaterials Based Approaches

Stem cells, tissue engineering, and biomaterials-based advances have continued to drive fetal spina

Table 2. Benefits and drawbacks of different surgical approaches

Approach	Key Benefits	Key Drawbacks
Postnatal Repair	<ul style="list-style-type: none"> • Non-invasive (post-birth) • Decreased risk of maternal morbidity • Decreased logistical complexity 	<ul style="list-style-type: none"> • Worst postnatal outcomes • Little reversal of neurological defects
Open Hysterotomy	<ul style="list-style-type: none"> • Well-established • Improved postnatal outcomes 	<ul style="list-style-type: none"> • Invasive • Necessitates c-section delivery • Risk of maternal morbidity
Percutaneous Fetoscopic	<ul style="list-style-type: none"> • Least invasive • Allows vaginal birth • Improved postnatal outcomes 	<ul style="list-style-type: none"> • More time-consuming • Increased risk of prematurity • Limited data
Laparotomy-Assisted Fetoscopic	<ul style="list-style-type: none"> • Less invasive • Allows vaginal birth • Improved postnatal outcomes • Lowest risk of prematurity among prenatal techniques 	<ul style="list-style-type: none"> • More time-consuming • Limited data
Percutaneous/Mini-Laparotomy	<ul style="list-style-type: none"> • Less invasive • Allows vaginal birth • Lowered risk of prematurity 	<ul style="list-style-type: none"> • More time-consuming • Limited data

bifida repair forward.^{132–134} In 2008, the concept of applying neural stem cells to spina bifida defects to aid in repair was validated in an animal model. Authors showed that application of stem cells to induced MMC defects in a lamb model led to the local production of neurotrophic factors.¹³⁵ This approach has since been built upon with the use of early gestational placental mesenchymal stromal cells (PMSCs) in prenatal MMC repair. These PMSCs are isolated from placental chorionic villus tissue and can be subsequently expanded and banked.¹³⁶ The cells can then be seeded into an extracellular matrix delivery vehicle which is applied to the defect. In mouse¹³⁷ and sheep^{138–143} models, this approach has shown promising results. Now, a clinical trial is currently underway at UC Davis Health called the “CuRe Trial: Cellular Therapy for In Utero Repair of Myelomeningocele”, which uses the approach outlined above to complement fetal surgery for spina bifida treatment. Robbie, the first baby who received this treatment, moved her legs and wiggled her toes after birth, and is thus far in very great health. Robbie is now crawling and kicking, and other babies have undergone the same treatment as the trial continues.^{83,84} See **Supplemental video 1** for an animation of the procedure used in the CuRe Trial.¹⁴⁴ Another stem cell-based advancement in development is the use of transamniotic stem cell therapy (TRASCET)¹⁴⁵ as an alternative to open fetal MMC repair. TRASCET involves harnessing and augmenting biological roles of specific fetal stem cells – in the context of spina bifida, placental-, amniotic fluid-, or bone marrow-derived stem cells – to exert therapeutic benefits. Rat and rabbit models utilizing this approach have shown promising results, with higher rates of partial or complete defect coverage and reductions in hindbrain herniation rates.^{146–151} Other stem cell-based advancements under investigation include differentiation of human induced pluripotent stem cells into neural crest stem cells for implantation into MMC defects, which has also been shown to be feasible in a lamb model;¹⁵² differentiation of amniotic fluid-derived stem cells into keratinocytes;¹⁵³ and use of fibroblasts and keratinocytes to bioengineer lab-grown fetal skin, which has been demonstrated in lamb models as well.¹⁵⁴

Other advancements in patch development and defect coverage are also being investigated. One such approach involves the use of basic fibroblast growth factors, delivered via an extracellular gelatin matrix

for coverage of the spina bifida defect. In 2010, researchers at CHOP demonstrated preliminary success in adhering their growth factor-crosslinked hydrogel sponge scaffold to the MMC defect in a rat model, with evidence of tissue ingrowth and angiogenesis.¹⁵⁵ They then went on to show equivalent effectiveness between this gelatin sponge and gelatin microspheres.¹⁵⁶ More recently, in 2016, these researchers reported feasibility of this approach in a sheep model,¹⁵⁷ complementing their previous work in rats. Another group at Yale is also investigating growth factor-based defect coverage and has shown success in rat models.¹⁵⁸ The feasibility of covering the defect using collagen-, small intestinal submucosa-, silicone, polypropylene or high-density polyethylene-, amniotic membrane-, biosynthetic cellulose-, and nanofiber-based scaffolds/patches, has also been shown in experimental animal models, and in some cases human patients.^{110,159–171} Defect coverage using umbilical cord-derived patches has also been shown to be successful in both rat¹⁷² and sheep^{173–177} models and in human fetal spina bifida repair.¹⁷⁸ There is currently an active clinical trial attempting to show feasibility of coverage of spina bifida defects with these patches as well.⁸²

Plastic Surgery, Robotic Surgery, and 3D Printed Materials

Outside of biomaterials, technological and engineering innovations have steadily enhanced prenatal MMC repair. In Zurich, researchers successfully demonstrated the first in-utero use of pedicled random pattern transposition flaps - which have been commonly used in adult and pediatric surgeries - for closing spinal defects, with minimal complications.¹⁷⁹ Robotic-assisted fetal surgery is another promising frontier.^{15,180} Although not yet standard, multiple animal studies have shown the feasibility of robot-assisted endoscopic repair,^{181–184} including novel methods using customized magnetic catheters to improve precision.¹⁸⁴ A major milestone came with the 2025 “Fetoscopic Robotic Open Spina Bifida Treatment” (FROST) study, which showcased the integration of robotics and 3D printing. Researchers created lifelike, 3D-printed uterine models, including a silicone fetus and placenta, to train surgeons in robot-assisted MMC repair. After 15–21 simulations, the surgeons could competently perform mock procedures.¹⁸⁵ Possible use of 3D

printed materials in training contexts such as these was discussed at a teaching session at the 2018 EuroCMR/SCMR joint congress, at which clinicians agreed that there were potential applications for these materials.¹⁸⁶ But use cases have expanded beyond training, and 3D printing is now being used to assess patch requirements and build patient-specific models for surgical planning.^{187,188} These advancements highlight how technology is driving safer, more effective, and increasingly personalized approaches to fetal spina bifida repair. The fruits of this can be seen not only in the United States, but internationally as well.

International Adoption

International adoption of prenatal spina bifida repair has also progressed, though variability exists in terms of inclusion and diagnostic criteria, surgical techniques, perioperative management strategies, and neonatal resuscitation practices.^{189–192} Despite this variability, there has also been increasing collaboration between prenatal MMC repair centers in different countries, and the development of more multidisciplinary approaches and teams.¹⁹³ Several non-U.S. countries are continuing to expand access to prenatal MMC repair and have seen improvements in outcomes over time, including (but certainly not limited to) Germany,^{194–198} Brazil,^{78,111} Taiwan,¹⁹⁹ Canada,^{200,201} France,^{202,203} and Israel.²⁰⁴ As prenatal repair of MMC expands to more countries and healthcare systems, it also raises important ethical questions about risk, access, and responsibility.

7. Unresolved Challenges

Limitations of Recent Advances

The advances outlined above have resulted in remarkable progress. However, they are not without limitations. Some, but not all the new techniques and technologies have been validated in humans, raising questions about translatability. For those which have shown applicability in human patients, limited data and small sample sizes of existing trials and studies must also be considered. As additional data are collected demonstrating safety and efficacy, it is likely that these novel interventions will need to roll out in waves. Eligibility criteria will start relatively narrow and expand over time, as has been the case for prenatal surgical approaches outlined above.

Regulatory hurdles should also be considered and foreseen. These approaches not only utilize biomaterials but also involve treatment of the most vulnerable group of people, unborn children, and therefore will understandably be met with scrutiny by regulatory agencies. While this scrutiny is crucial in ensuring that pregnant women and their children are protected, it will likely slow progress towards therapeutic advancements. Unresolved challenges exist not only pertaining to recent advances, but also to existing treatments and their availability.

Limited Domestic Access to Prenatal MMC Repair

Although adoption of in-utero MMC repair has grown in the United States, access remains limited. A 2024 national survey found that only 31% of U.S. hospitals offered both prenatal and postnatal MMC repair, while the majority provided postnatal repair only.²⁰⁵ One way by which access could be increased would be expansion of insurance coverage to support maternal travel to specialized centers offering prenatal MMC repair. Given the compelling evidence that prenatal repair significantly improves neurodevelopmental outcomes and mobility while reducing the long-term need for interventions such as shunting and catheterization, these benefits would translate not only to enhanced quality of life for affected children and their families but also to substantial healthcare cost savings over a lifetime.

Maternal Complications vs. Fetal Benefit

The main drawback of in-utero spina bifida repair is the incidence of negative maternal outcomes, as outlined above. The ethics of fetal surgery have been discussed at greater length elsewhere,^{206,207} but frequently discussed considerations include the weighing of risks to the mother against the benefits to the fetus. As with many other fetal surgical interventions, fetal MMC repair comes with risks of preterm birth, ruptured membranes, dehiscence, and other obstetric complications. Though as noted, the risk of some of these complications such as rupture are similar to typical c-section rates.^{94,96,97} Each of the approaches outlined above come with benefits and drawbacks, with some (such as the percutaneous/mini-laparotomy approach) having much lower risks of uterine dehiscence, and others (such as the laparotomy-assisted fetoscopic approach) having lower risks of premature birth. As these

- Clinicians now recognize the fetus as a patient in their own right.
- Folic acid supplementation plays an important role in spina bifida prevention worldwide.
- Prenatal surgical approaches for myelomeningocele (MMC) have evolved and continue to evolve, expanding treatment options and improving outcomes for both mother and child.

techniques continue to develop further, it is possible that new, hybrid approaches will be able to further limit drawbacks while continuing to draw on the strengths of different techniques. However, these obstetric complications must also be viewed considering the reality that during a pregnancy, doctors and surgeons are treating two patients – the mother and her unborn child.²⁰⁸⁻²¹⁰ In the words of the “Father of Fetal Surgery” himself, “The fetus is no longer a medical recluse hidden inside an opaque womb. The fetus is a patient with problems that cannot only be examined by an array of prenatal tests, but also can be actively managed by arranging the timing, mode, and place of delivery. In a few cases, the fetal problem can even be treated before birth”.²⁰⁹ For this second patient, prenatal surgery offers an option besides termination and a markedly improved quality of life.

8. Conclusions

Our understanding of spina bifida has greatly improved over the past several decades, with advances being made from prevention to diagnosis and management. A better grasp on the preventative roles of folic acid has allowed fewer cases of spina bifida to manifest in the first place. This highlights that clinicians worldwide should encourage folic acid supplementation prior to and during pregnancy. Improved imaging and diagnostics have allowed for more rapid and accurate identification of those cases which do manifest. Clinicians are now equipped with a multitude of techniques to repair the defect in utero, as new, hybrid approaches continue to develop to harness the benefits and limit the drawbacks of each. In practice, the findings of this review and the broader literature will inform the selection of surgical techniques by clinicians based upon each woman’s circumstances and risk profile. As the field moves forward, improvements will continue to be made. Stem cells and other biomaterials, robotic surgeries, 3D printing, improved imaging and diagnostic markers, and other refinements in technology and

technique have consistently pushed the limits of what is possible. Further work is needed to expand treatment availability so that more expecting mothers and their unborn children can access the fruits of this advancing field and pursue positive outcomes for both patients.

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Conflict of Interest

The authors have no conflicts of interest to disclose.

Author Contributions

Conceptualization, Z.B.S.; writing - original draft preparation Z.B.S.; writing - review and editing, Z.B.S. and K.E.F.; figure preparation, K.E.F.; project administration, Z.B.S. All authors have read and agreed to the published version of the manuscript.

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References

- 1 Atta CAM, Fiest KM, Frolkis AD, Jette N, Pringsheim T, St Germaine-Smith C et al. Global birth prevalence of spina bifida by folic acid fortification status: A systematic review and meta-analysis. *Am. J. Public Health.* 2016; 106: e24–e34.

- 2 Stallings EB, Isenburg JL, Rutkowski RE, Kirby RS, Nembhard WN, Sandidge T et al. National population-based estimates for major birth defects, 2016–2020. *Birth Defects Res* 2024; 116: e2301.
- 3 Adzick NS, Thom EA, Spong CY, Brock JW, Burrows PK, Johnson MP et al. A Randomized Trial of Prenatal versus Postnatal Repair of Myelomeningocele. *New England Journal of Medicine* 2011; 364: 993–1004.
- 4 Moldenhauer JS, Adzick NS. Fetal surgery for myelomeningocele: After the Management of Myelomeningocele Study (MOMS). *Semin Fetal Neonatal Med* 2017; 22: 360–366.
- 5 Singh R, Munakomi S. *Embryology, Neural Tube*. StatPearls Publishing, 2025. <http://www.ncbi.nlm.nih.gov/pubmed/28196803> (accessed 25 Jan. 2026).
- 6 Torchia MG, Persaud TVN. *The Developing Human: Clinically Oriented Embryology*. 12th ed. Elsevier, 2024.
- 7 Shaikh F, Sanadhya M, Kaleem S, Verma T, Jayaraj RL, Ahmad F. Critical appraisal on neural tube defects and their complexities. *Pediatr Neonatol* 2025; 66: 515–521.
- 8 Ledet III LF, Plaisance CJ, Daniel CP, Wagner MJ, Alvarez I, Burroughs CR et al. Spina Bifida Prevention: A Narrative Review of Folic Acid Supplements for Childbearing Age Women. *Cureus* 2024; 16: e53008.
- 9 Isaković J, Šimunić I, Jagečić D, Hribljan V, Mitrečić D. Overview of Neural Tube Defects: Gene–Environment Interactions, Preventative Approaches and Future Perspectives. *Biomedicines* 2022; 10: 965.
- 10 Aydin E, Peiro JL, Habli M. Closing the Gap: Prenatal Repair and the Reimagined Future of Spina Bifida. *Clin Obstet Gynecol* 2025; 68: 383–386.
- 11 Karsonovich T, Alruwaili AA, Das JM. Myelomeningocele. *StatPearls Publishing*, 2025. <http://www.ncbi.nlm.nih.gov/pubmed/2828669>.
- 12 Copp AJ, Adzick NS, Chitty LS, Fletcher JM, Holmbeck GN, Shaw GM. Spina bifida. *Nat Rev Dis Primers* 2015; 1: 15007.
- 13 Parvin A, Hasan MM. An Overview of Spina Bifida. *Open J Orthop* 2023; 13: 443–456.
- 14 Tennant PW, Pearce MS, Bythell M, Rankin J. 20-year survival of children born with congenital anomalies: a population-based study. *The Lancet* 2010; 375: 649–656.
- 15 Boswell TC, Ahn ES, Ruano R, Gargollo PC. Robotic Fetal Surgery: The Next Frontier? In: *Minimally Invasive and Robotic-Assisted Surgery in Pediatric Urology*. Springer International Publishing: Cham, 2020, pp 359–379.
- 16 Leung K-Y, Pai YJ, Chen Q, Santos C, Calvani E, Sudiwala S et al. Partitioning of One-Carbon Units in Folate and Methionine Metabolism Is Essential for Neural Tube Closure. *Cell Rep* 2017; 21: 1795–1808.
- 17 Sato K. Why is folate effective in preventing neural tube closure defects? *Med Hypotheses* 2020; 134: 109429.
- 18 Laurence KM, James N, Miller MH, Tennant GB, Campbell H. Double-blind randomised controlled trial of folate treatment before conception to prevent recurrence of neural-tube defects. *BMJ* 1981; 282: 1509–1511.
- 19 Smithells RW, Seller MJ, Harris R, Fielding DW, Schorah CJ, Nevin NC et al. Further Experience of Vitamin Supplementation for Prevention of Neural Tube Defect Recurrences. *The Lancet* 1983; 321: 1027–1031.
- 20 MRC Vitamin Study Research Group. Prevention of neural tube defects: Results of the Medical Research Council Vitamin Study. *The Lancet* 1991; 338: 131–137.
- 21 De-Regil LM, Peña-Rosas JP, Fernández-Gaxiola AC, Rayco-Solon P. Effects and safety of periconceptional oral folate supplementation for preventing birth defects. *Cochrane Database of Systematic Reviews* 2015; 2015: CD007950.
- 22 Bortolus R, Filippini F, Cipriani S, Trevisanuto D, Cavallin F, Zanconato G et al. Efficacy of 4.0 mg versus 0.4 mg Folic Acid Supplementation on the Reproductive Outcomes: A Randomized Controlled Trial. *Nutrients* 2021; 13: 4422.
- 23 Gomes S, Lopes C, Pinto E. Folate and folic acid in the periconceptional period: recommendations from official health organizations in thirty-six countries worldwide and WHO. *Public Health Nutr* 2016; 19: 176–189.
- 24 Williams J, Mai CT, Mulinare J, Isenburg J, Flood TJ, Ethen M et al. Updated Estimates of Neural Tube Defects Prevented by Mandatory Folic Acid Fortification–United States, 1995–2011. *MMWR Morb Mortal Wkly Rep* 2015; 64: 1–5.
- 25 Sayed AR, Bourne D, Pattinson R, Nixon J, Henderson B. Decline in the prevalence of neural tube defects following folic acid fortification and its cost-benefit in South Africa. *Birth Defects Res A Clin Mol Teratol* 2008; 82: 211–216.
- 26 Rodrigues VB, da Silva EN, dos Santos AM, Santos LMP. Prevented cases of neural tube defects and cost savings after folic acid fortification of flour in Brazil. *PLoS One* 2023; 18: e0281077.

- 27 De Wals P, Van Allen MI, Uh S-H, Lowry RB, Sibbald B, Evans JA et al. Reduction in Neural-Tube Defects after Folic Acid Fortification in Canada. *N Engl J Med* 2007; 357: 135–142.
- 28 López-Camelo JS, Castilla EE, Orioli IM. Folic acid flour fortification: Impact on the frequencies of 52 congenital anomaly types in three South American countries. *Am J Med Genet A* 2010; 152A: 2444–2458.
- 29 Yacob A, Carr CJ, Foote J, Scullen T, Werner C, Mathkour M et al. The Global Burden of Neural Tube Defects and Disparities in Neurosurgical Care. *World Neurosurg* 2021; 149: e803–e820.
- 30 Cengizler Ç, Ün MK, Büyükkurt S. A Nature-Inspired Search Space Reduction Technique for Spine Identification on Ultrasound Samples of Spina Bifida Cases. *Sci Rep* 2020; 10: 9280.
- 31 Cengizler C, Kerem Ün M, Buyukkurt S. A novel evolutionary method for spine detection in ultrasound samples of spina bifida cases. *Comput Methods Programs Biomed* 2021; 198: 105787.
- 32 Drukker L, Sharma H, Karim JN, Droste R, Noble JA, Papageorghiou AT. Clinical workflow of sonographers performing fetal anomaly ultrasound scans: deep-learning-based analysis. *Ultrasound in Obstetrics & Gynecology* 2022; 60: 759–765.
- 33 Barnes KS, Singh S, Barkley A, Lepard J, Hopson B, Cawyer CR et al. Determination of anatomic level of myelomeningocele by prenatal ultrasound. *Child's Nervous System* 2022; 38: 985–990.
- 34 Zhu X, Zhao S, Yang X, Feng Q, Zhang X, Yang F et al. First-Trimester Cranial Ultrasound Markers of Open Spina Bifida. *Journal of Ultrasound in Medicine* 2021; 40: 1155–1162.
- 35 Ungureanu DR, Comănescu MC, Istrate-Ofițeru A-M, Zorilă G-L, Drăgușin RC, Iliescu DG. Open Spina Bifida: The Role of Ultrasound Markers in the First Trimester and Morphopathology Correlation. *Curr Health Sci J* 2023; 49: 445–456.
- 36 Volpe N, Bovino A, Di Pasquo E, Corno E, Taverna M, Valentini B et al. First-trimester ultrasound of the cerebral lateral ventricles in fetuses with open spina bifida: a retrospective cohort study. *Am J Obstet Gynecol MFM* 2024; 6: 101445.
- 37 Milani HJF, de Sá Barreto EQ, Araujo Júnior E, Cavalheiro S, Barbosa MM, Moron AF. Measurement of the Area and Circumference of the Leg: Preliminary Results of a New Method for Estimating Leg Muscle Trophism in Fetuses With Open Lumbosacral Spina Bifida. *Journal of Ultrasound in Medicine* 2022; 41: 377–388.
- 38 Barreto EQ de S, Cavalheiro S, Milani HJF, Barbosa MM, Araujo Júnior E, Nardoza LMM et al. Cerebellar herniation demonstrated by the occipitum-dens line: Ultrasonography assessment of normal fetuses, fetuses with myelomeningocele, and fetuses that underwent antenatal myelomeningocele surgery. *Prenat Diagn* 2018; 38: 280–285.
- 39 Yin J, Wang Y, Wang S, Li G, Gu H, Chen L. Research progress on ultrasound and molecular markers for prenatal diagnosis of neural tube defects. *Heliyon* 2024; 10: e36060.
- 40 Chao A-S, Jhang L-S, Hsieh PC-C. Prenatal Diagnosis and Outcomes of Cervical Meningocele and Myelomeningocele. *J Med Ultrasound* 2024; 32: 21–24.
- 41 Di Mascio D, Greco F, Rizzo G, Khalil A, Buca D, Sorrentino F et al. Diagnostic accuracy of prenatal ultrasound in identifying the level of the lesion in fetuses with open spina bifida: A systematic review and meta-analysis. *Acta Obstet Gynecol Scand* 2021; 100: 210–219.
- 42 Nagaraj UD, Bierbrauer KS, Stevenson CB. Imaging Fetal Spine Malformations in the Context of In Utero Surgery. *Magn Reson Imaging Clin N Am* 2024; 32: 431–442.
- 43 Nagaraj UD, Bierbrauer KS, Stevenson CB, Peiro JL, Lim FY, Habli MA et al. Prenatal and postnatal MRI findings in open spinal dysraphism following intrauterine repair via open versus fetoscopic surgical techniques. *Prenat Diagn* 2020; 40: 49–57.
- 44 Tang X, Bai G, Wang H, Guo F, Yin H. A comparison of the accuracy of fetal magnetic resonance imaging and ultrasonography for the diagnosis of fetal congenital malformations of the spine and spinal cord. *Prenat Diagn* 2022; 42: 1295–1302.
- 45 Garel J, Rossi A, Blondiaux E, Cassart M, Hoffmann C, Garel C. Prenatal imaging of the normal and abnormal spinal cord: recommendations from the Fetal Task Force of the European Society of Paediatric Radiology (ESPR) and the European Society of Neuroradiology (ESNR) Pediatric Neuroradiology Committee. *Pediatr Radiol* 2023; 54: 548–561.
- 46 Meller C, Covini D, Aiello H, Izbizky G, Medina SP, Otano L. Update on prenatal diagnosis and fetal surgery for myelomeningocele. *Arch Argent Pediatr* 2021; 119: e215–e228.
- 47 Walter U. What can fetal neurosonography reveal about the future of an unborn child? *Ultraschall in der Medizin - European Journal of Ultrasound* 2025; 46: 418–424.

- 48 Royal C, Chertin L, Alfawzan M, Killian ME. Novel Techniques in Antenatal Imaging of Spinal Dysraphisms. *Curr Urol Rep* 2025; 26: 31.
- 49 Lei Y, Finnell RH. New Techniques for the Study of Neural Tube Defects. *Adv Tech Biol Med* 2015; 04: 157.
- 50 Lemay P, De Marco P, Traverso M, Merello E, Dionne-Laporte A, Spiegelman D et al. Whole exome sequencing identifies novel predisposing genes in neural tube defects. *Mol Genet Genomic Med* 2019; 7: e00467.
- 51 Lemay P, De Marco P, Emond A, Spiegelman D, Dionne-Laporte A, Laurent S et al. Rare deleterious variants in GRHL3 are associated with human spina bifida. *Hum Mutat* 2017; 38: 716–724.
- 52 Lemay P, Guyot M-C, Tremblay É, Dionne-Laporte A, Spiegelman D, Henrion É et al. Loss-of-function de novo mutations play an important role in severe human neural tube defects. *J Med Genet* 2015; 52: 493–497.
- 53 Azzarà A, Rendeli C, Crivello AM, Brugnoletti F, Rumore R, Ausili E et al. Identification of new candidate genes for spina bifida through exome sequencing. *Child's Nervous System* 2021; 37: 2589–2596.
- 54 Lei Y, Zhu H, Duhon C, Yang W, Ross ME, Shaw GM et al. Mutations in Planar Cell Polarity Gene SCRIB Are Associated with Spina Bifida. *PLoS One* 2013; 8: e69262.
- 55 Lei Y, Kim S, Chen Z, Cao X, Zhu H, Yang W et al. Variants identified in PTK7 associated with neural tube defects. *Mol Genet Genomic Med* 2019; 7: e00584.
- 56 Robinson A, Escuin S, Doudney K, Vekemans M, Stevenson RE, Greene NDE et al. Mutations in the planar cell polarity genes CELSR1 and SCRIB are associated with the severe neural tube defect craniorachischisis. *Hum Mutat* 2012; 33: 440–447.
- 57 Beaumont M, Akloul L, Carré W, Quélin C, Journal H, Pasquier L et al. Targeted panel sequencing establishes the implication of planar cell polarity pathway and involves new candidate genes in neural tube defect disorders. *Hum Genet* 2019; 138: 363–374.
- 58 Michejda M. Intrauterine Treatment of Spina Bifida: Primate Model. *European Journal of Pediatric Surgery* 1984; 39: 259–261.
- 59 Heffez DS, Aryanpur J, Hutchins GM, Freeman JM. The paralysis associated with myelomeningocele: clinical and experimental data implicating a preventable spinal cord injury. *Neurosurgery* 1990; 26: 987–92.
- 60 Heffez DS, Aryanpur J, Rotellini NAC, Hutchins GM, Freeman JM. Intrauterine Repair of Experimental Surgically Created Dysraphism. *Neurosurgery* 1993; 32: 1005–1010.
- 61 Meuli M, Meuli-Simmen C, Hutchins GM, Yingling CD, Hoffman KM, Harrison MR et al. In utero surgery rescues neurological function at birth in sheep with spina bifida. *Nat Med* 1995; 1: 342–347.
- 62 Meuli M, Meuli-Simmen C, Yingling CD, Hutchins GM, Timmel GB, Harrison MR et al. In utero repair of experimental myelomeningocele saves neurological function at birth. *J Pediatr Surg* 1996; 31: 397–402.
- 63 Meuli M, Meuli-Simmen C, Hutchins GM, Seller MJ, Harrison MR, Adzick NS. The spinal cord lesion in human fetuses with myelomeningocele: Implications for fetal surgery. *J Pediatr Surg* 1997; 32: 448–452.
- 64 Meuli-Simmen C, Meuli M, Adzick NS, Harrison MR. Latissimus dorsi flap procedures to cover myelomeningocele in utero: A feasibility study in human fetuses. *J Pediatr Surg* 1997; 32: 1154–1156.
- 65 Bruner JP, Tulipan NE, Richards WO. Endoscopic coverage of fetal open myelomeningocele in utero. *Am J Obstet Gynecol* 1997; 176: 256–257.
- 66 Adzick NS, Sutton LN, Crombleholme TM, Flake AW. Successful fetal surgery for spina bifida. *The Lancet* 1998; 352: 1675–1676.
- 67 Bruner JP, Tulipan N, Paschall RL, Boehm FH, Walsh WF, Silva SR et al. Fetal surgery for myelomeningocele and the incidence of shunt-dependent hydrocephalus. *JAMA* 1999; 282: 1819–25.
- 68 Sutton LN, Adzick NS, Bilaniuk LT, Johnson MP, Crombleholme TM, Flake AW. Improvement in hindbrain herniation demonstrated by serial fetal magnetic resonance imaging following fetal surgery for myelomeningocele. *JAMA* 1999; 282: 1826–31.
- 69 Tulipan N, Hernanz-Schulman M, Bruner JP. Reduced Hindbrain Herniation after Intrauterine Myelomeningocele Repair: A Report of Four Cases. *Pediatr Neurosurg* 1998; 29: 274–278.
- 70 Farmer D I. In utero repair of myelomeningocele. *Archives of Surgery* 2003; 138: 872.
- 71 Houtrow AJ, MacPherson C, Jackson-Coty J, Rivera M, Flynn L, Burrows PK et al. Prenatal Repair and Physical Functioning Among Children With Myelomeningocele: A Secondary Analysis of a Randomized Clinical Trial. *JAMA Pediatr* 2021; 175: e205674.
- 72 Adzick NS, Thom EA, Spong CY, Brock JW, Burrows PK, Johnson MP et al. A Randomized Trial of Prenatal versus Postnatal Repair of Myelomeningocele:

- Supplementary Appendix. *New England Journal of Medicine* 2011; 364: 993–1004.
- 73 Tulipan N, Wellons JC, Thom EA, Gupta N, Sutton LN, Burrows PK et al. Prenatal surgery for myelomeningocele and the need for cerebrospinal fluid shunt placement. *J Neurosurg Pediatr* 2015; 16: 613–620.
- 74 Houtrow AJ, Thom EA, Fletcher JM, Burrows PK, Adzick NS, Thomas NH et al. Prenatal Repair of Myelomeningocele and School-age Functional Outcomes. *Pediatrics* 2020; 145: e20191544.
- 75 Farmer DL, Thom EA, Brock JW, Burrows PK, Johnson MP, Howell LJ et al. The Management of Myelomeningocele Study: full cohort 30-month pediatric outcomes. *Am J Obstet Gynecol* 2018; 218: 256.e1-256.e13.
- 76 Antiel RM, Adzick NS, Thom EA, Burrows PK, Farmer DL, Brock JW et al. Impact on family and parental stress of prenatal vs postnatal repair of myelomeningocele. *Am J Obstet Gynecol* 2016; 215: 522.e1-522.e6.
- 77 Swarup I, Talwar D, Howell LJ, Adzick NS, Horn BD. Orthopaedic outcomes of prenatal versus postnatal repair of myelomeningocele. *Journal of Pediatric Orthopaedics B* 2022; 31: 87–92.
- 78 Pedreira DAL, Zanon N, Nishikuni K, Moreira de Sá RA, Acacio GL, Chmait RH et al. Endoscopic surgery for the antenatal treatment of myelomeningocele: the CECAM trial. *Am J Obstet Gynecol* 2016; 214: 111.e1-111.e11.
- 79 National Library of Medicine. Fetoscopic Meningomyelocele Repair Study (fMMC). <https://clinicaltrials.gov/study/NCT02230072> (accessed 8 Feb. 2026).
- 80 Sanz-Cortes M, Whitehead WE, Johnson RM, Aldave G, Castillo H, Desai NK et al. Laparotomy-assisted, two-port fetoscopic myelomeningocele repair: infant to preschool outcomes. *J Neurosurg Pediatr* 2025; 35: 10–21.
- 81 Belfort MA, Whitehead WE, Shamshirsaz AA, Batani ZH, Olutoye OO, Olutoye OA et al. Fetoscopic Open Neural Tube Defect Repair: Development and Refinement of a Two-Port, Carbon Dioxide Insufflation Technique. *Obstetrics and gynecology* 2017; 129: 734–743.
- 82 National Library of Medicine. Fetoscopic NEOX Cord 1K® Spina Bifida Repair (Trial ID NCT04243889). <https://clinicaltrials.gov/study/NCT04243889> (accessed 26 Jan. 2026).
- 83 National Library of Medicine. Cellular Therapy for In Utero Repair of Myelomeningocele - The Cure Trial (CuRe) (Trial ID NCT04652908). <https://clinicaltrials.gov/study/NCT04652908> (accessed 26 Jan. 2026).
- 84 UC Davis Health Children’s Hospital. The CuRe Trial. <https://health.ucdavis.edu/children/cure-trial/> (accessed 26 Jan. 2026).
- 85 Grivell RM, Andersen C, Dodd JM. Prenatal versus postnatal repair procedures for spina bifida for improving infant and maternal outcomes. *Cochrane Database of Systematic Reviews* 2014; 2014: CD008825.
- 86 Flanders TM, Heuer GG, Madsen PJ, Buch VP, Mackell CM, Alexander EE et al. Detailed Analysis of Hydrocephalus and Hindbrain Herniation After Prenatal and Postnatal Myelomeningocele Closure: Report From a Single Institution. *Neurosurgery* 2020; 86: 637–645.
- 87 Munoz JL, Kelling E, Johnson RM, Buskmiller C, Whitehead WE, Joyeux L et al. Impact of Prenatal Repair for Fetal Myelomeningocele on Gastrointestinal Function. *J Pediatr* 2025; 282: 114573.
- 88 Moldenhauer JS, Soni S, Rintoul NE, Spinner SS, Khalek N, Martinez-Poyer J et al. Fetal Myelomeningocele Repair: The Post-MOMS Experience at the Children’s Hospital of Philadelphia. *Fetal Diagn Ther* 2015; 37: 235–240.
- 89 Vonzun L, Ruegg L, Zepf J, Strübing N, Grethen P, Meuli M et al. Middle Cerebral Artery Doppler before and after Fetal Spina Bifida Repair: An Indirect Sign of Hindbrain Compression and Decompression? *Fetal Diagn Ther* 2024; 52: 1–9.
- 90 Danzer E, Gerdes M, Bebbington MW, Koh J, Adzick SN, Johnson MP. Fetal myelomeningocele surgery: preschool functional status using the Functional Independence Measure for children (WeeFIM). *Child’s Nervous System* 2011; 27: 1083–1088.
- 91 Danzer E, Thomas NH, Thomas A, Friedman KB, Gerdes M, Koh J et al. Long-term neurofunctional outcome, executive functioning, and behavioral adaptive skills following fetal myelomeningocele surgery. *Am J Obstet Gynecol* 2016; 214: 269.e1-269.e8.
- 92 Danzer E, Gerdes M, Bebbington MW, Zarnow DM, Adzick NS, Johnson MP. Preschool neurodevelopmental outcome of children following fetal myelomeningocele closure. *Am J Obstet Gynecol* 2010; 202: 450.e1-450.e9.
- 93 Johnson MP, Gerdes M, Rintoul N, Pasquariello P, Melchionni J, Sutton LN et al. Maternal-fetal surgery for myelomeningocele: Neurodevelopmental outcomes at 2 years of age. *Am J Obstet Gynecol* 2006; 194: 1145–1150.

- 94 Goodnight WH, Bahtiyar O, Bennett KA, Emery SP, Lillegard JB, Fisher A et al. Subsequent pregnancy outcomes after open maternal-fetal surgery for myelomeningocele. *Am J Obstet Gynecol* 2019; 220: 494.e1-494.e7.
- 95 Johnson MP, Bennett KA, Rand L, Burrows PK, Thom EA, Howell LJ et al. The Management of Myelomeningocele Study: obstetrical outcomes and risk factors for obstetrical complications following prenatal surgery. *Am J Obstet Gynecol* 2016; 215: 778.e1-778.e9.
- 96 Wilson RD, Johnson MP, Flake AW, Crombleholme TM, Hedrick HL, Wilson J et al. Reproductive outcomes after pregnancy complicated by maternal-fetal surgery. *Am J Obstet Gynecol* 2004; 191: 1430–1436.
- 97 Wilson RD, Lemerand K, Johnson MP, Flake AW, Bebbington M, Hedrick HL et al. Reproductive outcomes in subsequent pregnancies after a pregnancy complicated by open maternal-fetal surgery (1996–2007). *Am J Obstet Gynecol* 2010; 203: 209.e1-209.e6.
- 98 Brock JW, Thomas JC, Baskin LS, Zderic SA, Thom EA, Burrows PK et al. Effect of Prenatal Repair of Myelomeningocele on Urological Outcomes at School Age. *Journal of Urology* 2019; 202: 812–818.
- 99 Parizi JLG, Leal da Cruz M, Andrade MC, Garrone G, Ottoni SL, Cavalheiro S et al. A Comparative Analysis of Bladder Pattern of Patients who Underwent In Utero Versus Postnatal Myelomeningocele Repair. *Journal of Urology* 2020; 203: 194–199.
- 100 Evangelista A, Ruccolo L, Friuli V, Benazzo M, Conti B, Pisani S. Advances in Fetal Repair of Spina Bifida Integrating Prenatal Surgery, Stem Cells, and Biomaterials. *Biomedicines* 2026; 14: 136.
- 101 Moldenhauer JS, Soni S, Jatres J, Gebb J, Khalek N, Paidas Teefey C et al. Open Fetal Surgical Outcomes for Myelomeningocele Closure Stratified by Maternal Body Mass Index in a Large Single-Center Cohort. *Fetal Diagn Ther* 2020; 47: 889–893.
- 102 Yamashiro KJ, Farmer DL. Fetal myelomeningocele repair: a narrative review of the history, current controversies and future directions. *Transl Pediatr* 2021; 10: 1497–1505.
- 103 Jouannic J-M, Dugas A, Maurice P, Dhombres F, Garel C, Blondiaux É et al. Should we modify eligibility criteria for fetal surgery for open spinal dysraphism? *European Journal of Obstetrics & Gynecology and Reproductive Biology* 2026; 318: 114912.
- 104 Flanders TM, Madsen PJ, Pisapia JM, Hudgins ED, Mackell CM, Alexander EE et al. Improved Postoperative Metrics with Modified Myofascial Closure in Fetal Myelomeningocele Repair. *Operative Neurosurgery* 2020; 18: 158–165.
- 105 Krause M, Leibnitz F, Knüpfer MM, Merckenschlager A, Griessenauer CJ, Gburek-Augustat J. The potential impact of intraoperative neurophysiological monitoring on neurological function outcomes after postnatal spina bifida repair. *Child’s Nervous System* 2025; 41: 119.
- 106 Keil C, Sass B, Schulze M, Köhler S, Axt-Fliedner R, Bedei I. The intrauterine treatment of open spinal dysraphism. *Dtsch Arztebl Int* 2025; 122: 33–37.
- 107 Zargarzadeh N, Sambatur E, Abiad M, Rojhani E, Javinani A, Northam W et al. Gestational age at birth varies by surgical technique in prenatal open spina bifida repair: a systematic review and meta-analysis. *Am J Obstet Gynecol* 2025; 232: 524–537.
- 108 Chmait RH, Monson MA, Pham HQ, Chu JK, Van Speybroeck A, Chon AH et al. Percutaneous/mini-laparotomy fetoscopic repair of open spina bifida: a novel surgical technique. *Am J Obstet Gynecol* 2022; 227: 375–383.
- 109 Danzer E, Joyeux L, Flake AW, Deprest J. Fetal surgical intervention for myelomeningocele: lessons learned, outcomes, and future implications. *Dev Med Child Neurol* 2020; 62: 417–425.
- 110 Stevenson CB, Fletcher S, Larrew T, Chu JK. In-utero repair of open neural tube defects, lesion closure techniques and the choice of patch. *Best Pract Res Clin Obstet Gynaecol* 2025; 103: 102677.
- 111 Pedreira DAL, Zanon N, de Sá RAM, Acacio GL, Ogeda E, Belem TMLOU et al. Fetoscopic single-layer repair of open spina bifida using a cellulose patch: preliminary clinical experience. *The Journal of Maternal-Fetal & Neonatal Medicine* 2014; 27: 1613–1619.
- 112 Naus CA, Mann DG, Andropoulos DB, Belfort MA, Sanz-Cortes M, Whitehead WE et al. “This is how we do it” Maternal and fetal anesthetic management for fetoscopic myelomeningocele repairs: the Texas Children’s Fetal Center protocol. *Int J Obstet Anesth* 2025; 61: 104316.
- 113 Sanz Cortes M, Chmait RH, Lapa DA, Belfort MA, Carreras E, Miller JL et al. Experience of 300 cases of prenatal fetoscopic open spina bifida repair: report of the International Fetoscopic Neural Tube Defect Repair Consortium. *Am J Obstet Gynecol* 2021; 225: 678.e1-678.e11.
- 114 Sanz Cortes M, Lapa DA, Acacio GL, Belfort M, Carreras E, Maiz N et al. Proceedings of the First Annual Meeting of the International Fetoscopic

- Myelomeningocele Repair Consortium. *Ultrasound in Obstetrics & Gynecology* 2019; 53: 855–863.
- 115 Kunpalin Y, Karadjole VS, Medeiros ESB, Domínguez-Moreno M, Sichitiu J, Abbasi N et al. Benefits and complications of fetal and postnatal surgery for open spina bifida: systematic review and proportional meta-analysis. *Ultrasound in Obstetrics & Gynecology* 2025; 66: 135–146.
- 116 Kabagambe SK, Jensen GW, Chen YJ, Vanover MA, Farmer DL. Fetal Surgery for Myelomeningocele: A Systematic Review and Meta-Analysis of Outcomes in Fetoscopic versus Open Repair. *Fetal Diagn Ther* 2018; 43: 161–174.
- 117 Fareed A, Farhat S, Kerhani AA, Choudhary A, Raza SSM. Fetal in-utero management of myelomeningocele: a mini-review on history, challenges, management gap, and recommendations. *Annals of Medicine & Surgery* 2024; 86: 3196–3198.
- 118 Paslaru FG, Panaitescu AM, Iancu G, Veduta A, Gica N, Paslaru AC et al. Myelomeningocele Surgery over the 10 Years Following the MOMS Trial: A Systematic Review of Outcomes in Prenatal versus Postnatal Surgical Repair. *Medicina (B Aires)* 2021; 57: 707.
- 119 Lapa (Pedreira) DA, Acacio GL, Gonçalves RT, Sá RAM, Brandt RA, Chmait RH et al. Percutaneous fetoscopic closure of large open spina bifida using a bilaminar skin substitute. *Ultrasound in Obstetrics & Gynecology* 2018; 52: 458–466.
- 120 Chmait RH, Chu JK, Van Speybroeck AL, Llanes MsAS, Korst LM, Nguyen HN et al. Fetoscopic repair of open spina bifida between 26 0/7 and 27 6/7 gestational weeks and postnatal cerebrospinal fluid diversion. *The Journal of Maternal-Fetal & Neonatal Medicine* 2025; 38: 2477770.
- 121 Corroenne R, Rangwani S, Whitehead WE, Johnson RM, Nassr AA, Buskmiller C et al. Neurodevelopmental Outcomes after Fetoscopic Myelomeningocele Repair. *J Pediatr* 2025; 279: 114472.
- 122 Kohn JR, Rao V, Sellner AA, Sharhan D, Espinoza J, Shamshirsaz AA et al. Management of Labor and Delivery After Fetoscopic Repair of an Open Neural Tube Defect. *Obstetrics & Gynecology* 2018; 131: 1062–1068.
- 123 Miranda ML, Ximenes R, Andrade KC, Baldo C, Villarreal M, Caetano MR et al. Safety and Effectiveness of Fetal Myelomeningocele Repair: Case Series Analysis Using an Exteriorized Uterus and a Fetoscopic Approach. *Fetal Diagn Ther* 2025; 52: 521–531.
- 124 Sanz Cortes M, Davila I, Torres P, Yopez M, Lee W, Guimaraes C V. et al. Does fetoscopic or open repair for spina bifida affect fetal and postnatal growth? *Ultrasound in Obstetrics & Gynecology* 2019; 53: 314–323.
- 125 Sanz Cortes M, Corroenne R, Pyarali M, Johnson RM, Whitehead WE, Espinoza J et al. Ambulation after in-utero fetoscopic or open neural tube defect repair: predictors for ambulation at 30 months. *Ultrasound in Obstetrics & Gynecology* 2024; 64: 203–213.
- 126 Duron V, Miller R, Feldstein N, Schmoke N, Wu YS, Shirel T et al. Outcomes Following Fetoscopic Repair of Myelomeningocele: A Prospective Single-Center Experience. *Prenat Diagn* 2025; 45: 658–667.
- 127 American College of Obstetricians & Gynecologists (ACOG). Committee Opinion No. 720: Maternal–Fetal Surgery for Myelomeningocele. *Obstetrics & Gynecology* 2017; 130: e164–e167.
- 128 Giné C, Arévalo S, Maíz N, Rodó C, Manrique S, Poca A et al. Fetoscopic two-layer closure of open neural tube defects. *Ultrasound in Obstetrics & Gynecology* 2018; 52: 452–457.
- 129 Bowman R, Alhajjat A, Muller R, Scoville J, Shaaban A. Achieving water-tight open spina bifida closure through a novel three-port three-layer fetoscopic repair. *Am J Obstet Gynecol MFM* 2025; 7: 101697.
- 130 Cruz SM, Hameedi S, Sbragia L, Ogunleye O, Diefenbach K, Isaacs AM et al. Fetoscopic Myelomeningocele (MMC) Repair: Evolution of the Technique and a Call for Standardization. *J Clin Med* 2025; 14: 1402.
131. Children's Hospital of Philadelphia. MOMS3: Follow-up in the Teen and Young Adult Years to the Management of Myelomeningocele Study. 2023. <https://www.chop.edu/news/moms3-follow-management-myelomeningocele-study> (accessed 19 Feb. 2026).
132. Dhaulakhandi DB, Rohilla S, Rattan KN. Neural Tube Defects: Review of Experimental Evidence on Stem Cell Therapy and Newer Treatment Options. *Fetal Diagn Ther* 2010; 28: 72–78.
- 133 Watanabe M, Kim AG, Flake AW. Tissue Engineering Strategies for Fetal Myelomeningocele Repair in Animal Models. *Fetal Diagn Ther* 2015; 37: 197–205.
- 134 Winkler SM, Harrison MR, Messersmith PB. Biomaterials in fetal surgery. *Biomater Sci* 2019; 7: 3092–3109.
- 135 Fauza DO, Jennings RW, Teng YD, Snyder EY. Neural stem cell delivery to the spinal cord in an ovine model of fetal surgery for spina bifida. *Surgery* 2008; 144: 367–373.

- 136 Lankford L, Chen YJ, Saenz Z, Kumar P, Long C, Farmer D et al. Manufacture and preparation of human placenta-derived mesenchymal stromal cells for local tissue delivery. *Cytotherapy* 2017; 19: 680–688.
- 137 Jackson JE, Pivetti C, Stokes SC, Theodorou CM, Kumar P, Paxton ZJ et al. Placental Mesenchymal Stromal Cells: Preclinical Safety Evaluation for Fetal Myelomeningocele Repair. *Journal of Surgical Research* 2021; 267: 660–668.
- 138 Theodorou CM, Stokes SC, Jackson JE, Pivetti CD, Kumar P, Yamashiro KJ et al. Efficacy of clinical-grade human placental mesenchymal stromal cells in fetal ovine myelomeningocele repair. *J Pediatr Surg* 2022; 57: 753–758.
- 139 Stokes SC, Theodorou CM, Jackson JE, Pivetti C, Kumar P, Yamashiro KJ et al. Long-term safety evaluation of placental mesenchymal stromal cells for in utero repair of myelomeningocele in a novel ovine model. *J Pediatr Surg* 2022; 57: 18–25.
- 140 Wang A, Brown EG, Lankford L, Keller BA, Pivetti CD, Sitkin NA et al. Placental Mesenchymal Stromal Cells Rescue Ambulation in Ovine Myelomeningocele. *Stem Cells Transl Med* 2015; 4: 659–669.
- 141 Vanover M, Pivetti C, Lankford L, Kumar P, Galganski L, Kabagambe S et al. High density placental mesenchymal stromal cells provide neuronal preservation and improve motor function following in utero treatment of ovine myelomeningocele. *J Pediatr Surg* 2019; 54: 75–79.
- 142 Kabagambe S, Keller B, Becker J, Goodman L, Pivetti C, Lankford L et al. Placental mesenchymal stromal cells seeded on clinical grade extracellular matrix improve ambulation in ovine myelomeningocele. *J Pediatr Surg* 2018; 53: 178–182.
- 143 Galganski LA, Kumar P, Vanover MA, Pivetti CD, Anderson JE, Lankford L et al. In utero treatment of myelomeningocele with placental mesenchymal stromal cells — Selection of an optimal cell line in preparation for clinical trials. *J Pediatr Surg* 2020; 55: 1941–1946.
- 144 ProLifedoc Inc. CuRe Trial Animation.
- 145 Lazow SP, Fauza DO. Transamniotic Stem Cell Therapy. In: Turksen, K. (eds) *Cell Biology and Translational Medicine, Volume 7. Advances in Experimental Medicine and Biology()*. Springer, 2019, pp 61–74.
- 146 Shieh HF, Tracy SA, Hong CR, Chalphin A V, Ahmed A, Rohrer L et al. Transamniotic stem cell therapy (TRASCET) in a rabbit model of spina bifida. *J Pediatr Surg* 2019; 54: 293–296.
- 147 Dionigi B, Ahmed A, Brazzo J, Connors JP, Zurakowski D, Fauza DO. Partial or complete coverage of experimental spina bifida by simple intra-amniotic injection of concentrated amniotic mesenchymal stem cells. *J Pediatr Surg* 2015; 50: 69–73.
- 148 Dionigi B, Brazzo JA, Ahmed A, Feng C, Wu Y, Zurakowski D et al. Trans-amniotic stem cell therapy (TRASCET) minimizes Chiari-II malformation in experimental spina bifida. *J Pediatr Surg* 2015; 50: 1037–1041.
- 149 Feng C, D. Graham C, Connors JP, Brazzo J, Zurakowski D, Fauza DO. A comparison between placental and amniotic mesenchymal stem cells for transamniotic stem cell therapy (TRASCET) in experimental spina bifida. *J Pediatr Surg* 2016; 51: 1010–1013.
- 150 Li X, Yuan Z, Wei X, Li H, Zhao G, Miao J et al. Application potential of bone marrow mesenchymal stem cell (BMSCs) based tissue-engineering for spinal cord defect repair in rat fetuses with spina bifida aperta. *J Mater Sci Mater Med* 2016; 27: 77.
- 151 Ma W, Wei X, Gu H, Li H, Guan K, Liu D et al. Sensory neuron differentiation potential of in utero mesenchymal stem cell transplantation in rat fetuses with spina bifida aperta. *Birth Defects Res A Clin Mol Teratol* 2015; 103: 772–779.
- 152 Saadai P, Wang A, Nout YS, Downing TL, Lofberg K, Beattie MS et al. Human induced pluripotent stem cell-derived neural crest stem cells integrate into the injured spinal cord in the fetal lamb model of myelomeningocele. *J Pediatr Surg* 2013; 48: 158–163.
- 153 Basler M, Pontiggia L, Biedermann T, Reichmann E, Meuli M, Mazzone L. Bioengineering of Fetal Skin: Differentiation of Amniotic Fluid Stem Cells into Keratinocytes. *Fetal Diagn Ther* 2020; 47: 198–204.
- 154 Mazzone L, Moehrlen U, Ochsenbein-Kölbl N, Pontiggia L, Biedermann T, Reichmann E et al. Bioengineering and in utero transplantation of fetal skin in the sheep model: A crucial step towards clinical application in human fetal spina bifida repair. *J Tissue Eng Regen Med* 2020; 14: 58–65.
- 155 Watanabe M, Jo J, Radu A, Kaneko M, Tabata Y, Flake AW. A Tissue Engineering Approach for Prenatal Closure of Myelomeningocele with Gelatin Sponges Incorporating Basic Fibroblast Growth Factor. *Tissue Eng Part A* 2010; 16: 1645–1655.
- 156 Watanabe M, Li H, Roybal J, Santore M, Radu A, Jo J-I et al. A Tissue Engineering Approach for Prenatal Closure of Myelomeningocele: Comparison of Gelatin Sponge and Microsphere Scaffolds and Bioactive Protein Coatings. *Tissue Eng Part A* 2011; 17: 1099–1110.

- 157 Watanabe M, Li H, Kim AG, Weilerstein A, Radu A, Davey M et al. Complete tissue coverage achieved by scaffold-based tissue engineering in the fetal sheep model of Myelomeningocele. *Biomaterials* 2016; 76: 133–143.
- 158 Farrelly JS, Bianchi AH, Ricciardi AS, Buzzelli GL, Ahle SL, Freedman-Weiss MR et al. Alginate microparticles loaded with basic fibroblast growth factor induce tissue coverage in a rat model of myelomeningocele. *J Pediatr Surg* 2019; 54: 80–85.
- 159 Eggink AJ, Roelofs LAJ, Feitz WFJ, Wijnen RMH, Mullaart RA, Grotenhuis JA et al. In utero Repair of an Experimental Neural Tube Defect in a Chronic Sheep Model Using Biomatrices. *Fetal Diagn Ther* 2005; 20: 335–340.
- 160 Eggink AJ, Roelofs LAJ, Lammens MMY, Feitz WFJ, Wijnen RMH, Mullaart RA et al. Histological Evaluation of Acute Covering of an Experimental Neural Tube Defect with Biomatrices in Fetal Sheep. *Fetal Diagn Ther* 2006; 21: 210–216.
- 161 Eggink AJ, Roelofs LAJ, Feitz WFJ, Wijnen RMH, Lammens MMY, Mullaart RA et al. Delayed Intrauterine Repair of an Experimental Spina Bifida with a Collagen Biomatrix. *Pediatr Neurosurg* 2008; 44: 29–35.
- 162 Fontecha CG, Peiro JL, Sevilla JJ, Aguirre M, Soldado F, Fresno L et al. Fetoscopic coverage of experimental myelomeningocele in sheep using a patch with surgical sealant. *European Journal of Obstetrics & Gynecology and Reproductive Biology* 2011; 156: 171–176.
- 163 Fontecha CG, Peiro JL, Aguirre M, Soldado F, Añor S, Fresno L et al. Inert patch with bioadhesive for gentle foetal surgery of myelomeningocele in a sheep model. *European Journal of Obstetrics & Gynecology and Reproductive Biology* 2009; 146: 174–179.
- 164 Peiro JL, Fontecha CG, Ruano R, Esteves M, Fonseca C, Marotta M et al. Single-Access Fetal Endoscopy (SAFE) for myelomeningocele in sheep model I: amniotic carbon dioxide gas approach. *Surg Endosc* 2013; 27: 3835–3840.
- 165 Brown EG, Saadai P, Pivetti CD, Beattie MS, Bresnahan JC, Wang A et al. In utero repair of myelomeningocele with autologous amniotic membrane in the fetal lamb model. *J Pediatr Surg* 2014; 49: 133–138.
- 166 Saadai P, Nout YS, Encinas J, Wang A, Downing TL, Beattie MS et al. Prenatal repair of myelomeningocele with aligned nanofibrous scaffolds—a pilot study in sheep. *J Pediatr Surg* 2011; 46: 2279–2283.
- 167 Oliveira R de CS e, Valente PR, Abou-Jamra RC, Araújo A, Saldiva PH, Pedreira DAL. Biosynthetic cellulose induces the formation of a neoduramater following pre-natal correction of meningocele in fetal sheep. *Acta Cir Bras* 2007; 22: 174–181.
- 168 Herrera SRF, Leme RJ de A, Valente PR, Caldini ÉG, Saldiva PHN, Pedreira DAL. Comparison between two surgical techniques for prenatal correction of meningocele in sheep. *Einstein (São Paulo)* 2012; 10: 455–461.
- 169 Kunpalin Y, Vergote S, Joyeux L, Telli O, David AL, Belfort M et al. Local host response of commercially available dural patches for fetal repair of spina bifida aperta in rabbit model. *Prenat Diagn* 2023; 43: 370–381.
- 170 Dasargyri A, Reichmann E, Moehrlen U. Bio-engineering of fetal cartilage for in utero spina bifida repair. *Pediatr Surg Int* 2020; 36: 25–31.
- 171 Gansevoort M, Oostendorp C, Bouwman LF, Tiemessen DM, Geutjes PJ, Feitz WFJ et al. Collagen-Heparin-FGF2-VEGF Scaffolds Induce a Regenerative Gene Expression Profile in a Fetal Sheep Wound Model. *Tissue Eng Regen Med* 2024; 21: 1173–1187.
- 172 Snowise S, Mann L, Morales Y, Moise KJ, Johnson A, Fletcher S et al. Cryopreserved human umbilical cord versus biocellulose film for prenatal spina bifida repair in a physiologic rat model. *Prenat Diagn* 2017; 37: 473–481.
- 173 Mann LK, Won JH, Patel R, Bergh EP, Garnett J, Bhattacharjee MB et al. Allografts for Skin Closure during In Utero Spina Bifida Repair in a Sheep Model. *J Clin Med* 2021; 10: 4928.
- 174 Papanna R, Moise KJ, Mann LK, Fletcher S, Schniederjan R, Bhattacharjee MB et al. Cryopreserved human umbilical cord patch for in-utero spina bifida repair. *Ultrasound in Obstetrics & Gynecology* 2016; 47: 168–176.
- 175 Papanna R, Mann L, Snowise S, Morales Y, Prabhu S, Tseng S et al. Neurological Outcomes after Human Umbilical Cord Patch for In Utero Spina Bifida Repair in a Sheep Model. *American Journal of Perinatology Reports* 2016; 06: e309–e317.
- 176 Athiel Y, Jouannic J, Mauffré V, Dehan C, Adam C, Blot S et al. Allogenic umbilical cord-derived mesenchymal stromal cells improve motor function in prenatal surgical repair of myelomeningocele: An ovine model study. *BJOG* 2024; 131: 759–767.
- 177 Guilbaud L, Dugas A, Weber M, Deflers C, Lallemand P, Lilin T et al. In utero treatment of myelomeningocele with allogenic umbilical cord-derived mesenchymal stromal cells in an ovine model. *Curr Res Transl Med* 2022; 70: 103314.

- 178 Papanna R, Fletcher S, Moise KJ, Mann LK, Tseng SCG. Cryopreserved Human Umbilical Cord for In Utero Myeloschisis Repair. *Obstetrics & Gynecology* 2016; 128: 325–330.
- 179 Meuli M, Meuli-Simmen C, Mazzone L, Tharakan SJ, Zimmermann R, Ochsenbein N et al. In utero Plastic Surgery in Zurich: Successful Use of Distally Pedicled Random Pattern Transposition Flaps for Definitive Skin Closure during Open Fetal Spina Bifida Repair. *Fetal Diagn Ther* 2018; 44: 173–178.
- 180 Berris M, Shoham M. Febotics – a marriage of fetal surgery and robotics. *Computer Aided Surgery* 2006; 11: 175–180.
- 181 Aaronson OS, Tulipan NB, Cywes R, Sundell HW, Davis GH, Bruner JP et al. Robot-Assisted Endoscopic Intrauterine Myelomeningocele Repair: A Feasibility Study. *Pediatr Neurosurg* 2002; 36: 85–89.
- 182 Knight CG, Lorincz A, Johnson A, Gidell K, Rabah R, Klein MD et al. Robot-enhanced fetoscopic surgery. *J Pediatr Surg* 2004; 39: 1463–1465.
- 183 Kohl T, Hartlage MG, Kiehitz D, Westphal M, Buller T, Achenbach S et al. Percutaneous fetoscopic patch coverage of experimental lumbosacral full-thickness skin lesions in sheep. *Surg Endosc* 2003; 17: 1218–23.
- 184 Gervasoni S, Lussi J, Viviani S, Boehler Q, Ochsenbein N, Moehrlen U et al. Magnetically Assisted Robotic Fetal Surgery for the Treatment of Spina Bifida. *IEEE Trans Med Robot Bionics* 2022; 4: 85–93.
- 185 Kunpalin Y, Kik CC, Lebouthillier F, Abbasi N, Ryan G, Spoor J et al. Fetoscopic Robotic Open Spina Bifida Treatment (FROST): A Preclinical Feasibility and Learning Curve Study. *BJOG* 2025; 132: 1259–1268.
- 186 Biglino G, Milano EG, Capelli C, Wray J, Shearn AI, Caputo M et al. Three-dimensional printing in congenital heart disease: Considerations on training and clinical implementation from a teaching session. *Int J Artif Organs* 2019; 42: 595–599.
- 187 Fils AJ, Kasmirski J, Okpaise O, Reynolds JM, Tonni G, Werner H et al. The Use of 3D Printing in Fetal Surgery for Surgical Planning: A Scoping Review. *J Clin Med* 2024; 13: 4999.
- 188 Miller JL, Ahn ES, Garcia JR, Miller GT, Satin AJ, Baschat AA. Ultrasound-based three-dimensional printed medical model for multispecialty team surgical rehearsal prior to fetoscopic myelomeningocele repair. *Ultrasound in Obstetrics & Gynecology* 2018; 51: 836–837.
- 189 Kik CC, Kunpalin Y, Kulkarni A V., DeKoninck PLJ, Spoor JKH, Van Mieghem T. Global variability in fetal spina bifida surgery: a survey of neurosurgical strategies. *J Neurosurg Pediatr* 2025; 36: 3–10.
- 190 Keil C, Wiora N, Krispin E, Windhorst A, Axt-Fliedner R, Bedei I. Evolving Practices in Prenatal Open Spinal Dysraphism: A Global Survey of Selection Criteria, Surgical Techniques, and Diagnostic Trends. *Prenat Diagn* 2026; 46: 75–83.
- 191 Nulens K, Kunpalin Y, Nijs K, Carvalho JCA, Pollard L, Abbasi N et al. Enhanced recovery after fetal spina bifida surgery: global practice. *Ultrasound in Obstetrics & Gynecology* 2024; 64: 669–677.
- 192 Gallagher K, Crombag N, Prashar K, Deprest J, Ourselin S, David AL et al. Global Policy and Practice for Intrauterine Fetal Resuscitation During Fetal Surgery for Open Spina Bifida Repair. *JAMA Netw Open* 2023; 6: e239855.
- 193 Castillo J, Locastro MM, Corroenne R, Malhotra A, Van Speybroeck A, Lai G et al. Maternal–fetal surgery for myelomeningocele longitudinal follow-up model: Mitigation of care fragmentation through care coordination and outcomes reporting. *J Pediatr Rehabil Med* 2025; 18: 146–154.
- 194 Kohl T, Tchatcheva K, Merz W, Wartenberg HC, Heep A, Müller A et al. Percutaneous fetoscopic patch closure of human spina bifida aperta: advances in fetal surgical techniques may obviate the need for early postnatal neurosurgical intervention. *Surg Endosc* 2009; 23: 890–895.
- 195 Degenhardt J, Schürg R, Winarno A, Oehmke F, Khaleeva A, Kaweckı A et al. Percutaneous minimal-access fetoscopic surgery for spina bifida aperta. Part II: maternal management and outcome. *Ultrasound in Obstetrics & Gynecology* 2014; 44: 525–531.
- 196 Kohl T. Percutaneous minimally invasive fetoscopic surgery for spina bifida aperta. Part I: surgical technique and perioperative outcome. *Ultrasound in Obstetrics & Gynecology* 2014; 44: 515–524.
- 197 El Damaty A, Elsässer M, Pfeifer U, Kotzaeridou U, Gille C, Spratte J et al. The first experience with 16 open microsurgical fetal surgeries for myelomeningocele in Germany. *European Journal of Paediatric Neurology* 2025; 55: 79–86.
- 198 Vonzun L, Kahr M, Noll F, Mazzone L, Moehrlen U, Meuli M et al. Systematic classification of maternal and fetal intervention-related complications following open fetal myelomeningocele repair – results from a large prospective cohort. *BJOG* 2021; 128: 1184–1191.
- 199 Feng M, Chen P-C, Lin G-R, Lin T-Y, Hsieh T-T, Shaw SW. The clinical experience of fetoscopic repair of myelomeningocele in Taiwan: The dilemma in prenatal decision-making and first successful case. *Taiwan J Obstet Gynecol* 2024; 63: 904–908.

- 200 Pruthi V, Abbasi N, Ryan G, Drake J, Kulkarni A V., Kwan-Wong T et al. Fetal Surgery for Open Spina Bifida in Canada: Initial Results. *Journal of Obstetrics and Gynaecology Canada* 2021; 43: 733-739.e1.
- 201 Kik CC, Kunpalin Y, Kulkarni A V., Varghese A, Abbasi N, Ryan G et al. Contemporary Outcomes of a National Fetal Spina Bifida Surgery Service. *Prenat Diagn* 2024; 44: 1635–1640.
- 202 Friszer S, Dhombres F, Di Rocco F, Rigouzzo A, Garel C, Guilbaud L et al. Résultats préliminaires de l'étude PRIUM : programme de réparation in utero des myéloméningocèles. *J Gynecol Obstet Biol Reprod (Paris)* 2016; 45: 738–744.
- 203 Guilbaud L, Maurice P, Lallemand P, De Saint-Denis T, Maisonneuve E, Dhombres F et al. Open fetal surgery for myelomeningocele repair in France. *J Gynecol Obstet Hum Reprod* 2021; 50: 102155.
- 204 Hadassah International. Hadassah Medical Center Performs Israel's First Spina Bifida Fetal Surgery to Lessen Crippling Spinal Damage. <https://hadassahinternational.org/hadassah-medical-center-performs-israels-first-spina-bifida-fetal-surgery-lessen-crippling-spinal-damage/> (accessed 26 Jan. 2026).
- 205 Garg SP, Shah K V., Lentskevich M, Yau A, Gosain AK. Prenatal Spina Bifida Repair: A Survey of Current Practice in the United States. *Plast Reconstr Surg Glob Open* 2024; 12: e6377.
- 206 Cavolo A, Gastmans C, Crombag N. Ethical challenges in conducting maternal-fetal surgery trials. A systematic review. *Pediatr Res* 2025; 98: 479–490.
- 207 Austin MT, Cole TR, McCullough LB, Chervenak FA. Ethical challenges in invasive maternal-fetal intervention. *Semin Pediatr Surg* 2019; 28: 150819.
- 208 HARRISON MR, ADZICK NS. The Fetus as a Patient Surgical Considerations. *Ann Surg* 1991; 213: 279–291.
- 209 Harrison MR. The Unborn Patient: Prenatal Diagnosis and Treatment. *Journal of the Japanese Society of Pediatric Surgeons* 1991; 27: 402–403.
- 210 Harrison MR, Evans ML, Adzick NS, Holzgreve W. The unborn patient: the art and science of fetal therapy. 3rd ed. W.B. Saunders Company, 2001.

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